Case Report

Dermatophytic Disease: Report of A Tunisian Case Caused by a Sterile Dermatophyte Identified as Trichophyton Rubrum by ITS1 and ITS4 Sequencing

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Abstract
Dermatophytic disease is a rare chronic mycosis characterized by polymorphic cutaneous lesions and possible visceral invasion, and is associated with immunodeficiency and refractoriness to drug treatment. We report herein a dermatophytic disease Tunisian case caused by a sterile dermatophyte further identified as Trichophyton rubrum by ITS1 and ITS4 sequencing.

The case we report herein highlights the usefulness of molecular techniques in identification of causative agent when the mycological criteria are lacking.

Keywords: Dermatophytic disease; Sterile dermatophyte; Trichophyton rubrum; Sequencing

Introduction
Subcutaneous mycoses, which are much less common than superficial fungal infections, include a heterogeneous group of infections that often result from direct penetration of the fungus into the dermis and subcutaneous tissue through traumatic injury [1]. Dermatophytic disease is a rare chronic mycosis characterized by polymorphic cutaneous lesions and possible visceral invasion, and is associated with immunodeficiency and refractoriness to drug treatment. Dermatophytic disease has mainly described in North Africa [2]; in Tunisia, most previously reported cases are caused by Trichophyton violaceum or Microsporum canis [3,4]. The problematic identification of filamentous fungi when classical morphological criteria are missing is now feasible by DNA sequencing [5].

We report herein a dermatophytic disease Tunisian case caused by a sterile dermatophyte further identified as Trichophyton rubrum by ITS1 and ITS4 sequencing.

Case Presentation
The patient is a 43 year-old Tunisian man and no known immunodeficiency. He was addressed to the lab of Mycology-Parasitology of Farhat Hached hospital, Sousse, Tunisia, for cutaneous lesions very evocative of tinea corporis. The lesions first appeared on the abdomen (Figure 1) and progressively spread to the back and the members. Later, papulo-nodules and vegetating plaques appeared and involved large parts of the body (Figure 2). On examination, neither onychomycosis nor tinea capitis were associated to skin lesions and no lymph nodes nor visceral involvement could be demonstrated.

History of the patient revealed that his father and sister were suffering of the same disease (later confirmed as dermatophytic disease caused by Trichophyton violaceum on mycological ground). On the other hand, the patient reported previous similar episodes which poorly responded to various drugs including antifungal agents (Fluconazole and itraconazole) with no improvement till now. Later, a genetic study was performed in all members of his family. All the patients with deep dermatophytosis had autosomal recessive CARD9 (caspase recruitment domain-containing protein 9) deficiency [6].

Direct mycological examination of skin and nodular lesions showed 3 to 5µm septated branching hyphae. Culture on Sabouraud dextrose agar of both kinds of samples yielded non-sporulating sterile colonies that couldn’t be identified at the specific level (Figure 3). In an effort to provide a definitive identification, cultures of the isolates were subjected to PCR targeting the ribosomal internal transcribed space (ITS) domain. Portions of fungal growth were treated for DNA extraction by using QIAamp® DNA Blood Mini Kit (Qiagen SA, Courtaboeuf, France).

Nucleic acid extraction was performed according to the manufacturers’ instructions. At the end of the procedure, the DNA pellet was dissolved in 50 to 70µl hydration solution. The following oligonucleotides were used as primers:

**ITS1: 5’ CAGGCCGGAGGCTGGCCCC 3’**
**ITS4: 5’ CGAGGACAACCAGAAAAGAT 3’**

**Figure 1:** Abdomen cutaneous lesions.
These primers amplify a 608bp fragment of the ITS regions of rDNA.

PCR was performed on DNA extracts from fungal isolates under the following conditions: the amplification reaction was performed in a total volume of 40µl; the PCR mixture contained 5µl of 10X reaction buffer (Hot star buffer; Qiagen SA, France), 5µl of 2mmol l⁻¹ desoxynucleoside triphosphates containing an equimolar mixture of dATP, dCTP, dGTP and dTTP (Qiagen), 1µl (10 µmol l⁻¹) of each primer, 0.2µl of Hot star Taq polymerase (Qiagen) and 50ng of template DNA. Samples were amplified through 45 cycles in a thermocycler (Thermolyne Amplitron II Series 1091, Barnstead Thermolyne Corporation, Dubuque, IA, USA) as follows: initial denaturation for 15min at 95°C, denaturation for 1mn at 95°C, annealing for 1mn at 54°C and extension for 2mn at 72°C. This was followed by a final extension step for 7min at 72°C.

The resulting amplicons were separated electrophoretically on 2% agarose gel and visualized under UV light after ethidium bromide staining (Figure 4).

The sequence alignment of the strain showed 100% homology with Trichophyton rubrum registered in Database Trichophyton rubrum strain ATCC MYA-4607 18S.

**Discussion**

Dermatophytic disease is a rare chronic mycosis characterized by polymorphic cutaneous lesions and possible visceral invasion. It is associated with immunodeficiency, in particular, various inborn errors of interleukin-17 immunity [6] and known to be resistant to antifungals [7]. Correct diagnosis of dermatophytic disease and identification of the causal agent is a major importance as it allows appropriate antifungal treatment to be promptly instituted. Usually, identification of the causing species is based on mycological criteria. The case we report herein shows that identification on the basis of the phenotypical characters may be difficult or impossible when culture yields sterile colonies. In this situation, genome sequencing is needed.

Molecular techniques are much beneficial for fungal identification owing to their rapidity, sensitivity and specificity. Moreover, these methods rely on genetic characters which are more constant than phenotypic ones. In addition, they help to characterize atypical or sterile fungi that are difficult to identify by mycological techniques [8,9,10]. Specific molecular-based techniques for identification of species or even strain in severe fungal infections can be used as possible alternatives for routine identification of fungi [11].

In our study, the identification of the isolate as Trichophyton rubrum could only be achieved after sequencing of ITS region. The pathogenic potential of Trichophyton rubrum is dependent on a variety of local and systemic factors that affect the natural host resistance to dermatophytic infection [12]. In nearly all instances, chronic cutaneous Trichophyton rubrum infections preclude deep invasion; tissue invasion is attributed to a presumed alteration of host resistance [13].

The case we report herein highlights the usefulness of molecular techniques in identification of causative agent when the mycological criteria are lacking.

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**References**


