An Integrative View on Intra- and Inter-Cellular Cooperation Mechanisms in Alzheimer’s Disease

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Introduction

Alzheimer’s disease (AD) has been identified as central nervous system pathology more than 100 years ago and for a long time the diagnostic criteria remained the same, based on the anatomopathological findings: amyloid plaques and fibrillary tangles. For almost half of century no significant progress has been made for this immutable disease, until electron micrograph studies identified amyloid fibrils in both Alzheimer’s disease and Down syndrome patients [1]. Combined with genetic studies, the Amyloid Precursor Protein (APP) gene and splicing proteins were identified and soon, familial forms of the disease and subsequent mutations were reported [2–4]. Familial studies, as well as molecular studies of late-onset AD led to identification of other risk factors, such as apolipoprotein E [5,6]. The new wave of knowledge came from attempts to identify the proteases involved in fibril generation and AD pathology. Observation that mutated presenilins (enzymes involved in Notch signaling) are related to aggressive forms of early onset AD [7] led to a demonstrated relationship between these gamma secretase and APP [8] followed by identification of alpha secretase [9] and beta secretase [10]. Beta secretase has soon become the main point of interest in the field of AD research for almost half a century, while in AD a β secretase cleaves the extracellular domain of APP, generating the C99 fragment, further cleaved by a gamma-secretase complex, with presenilins as main enzymatic component. In AD, a β secretase cleaves the extracellular domain of APP, generating the C99 fragment and creating the premises to yield Aβ peptides 40 and 42, with the latter exhibiting high propensity to aggregate in the extracellular space and create amyloid plaques. Besides these two main Aβ species, several other truncated species have been identified Cerelosporin Fluid (CSF), plasma or interstitial fluid [13]. Very recently, through nuclear magnetic resonance and electron paramagnetic resonance spectroscopy, it was shown that C99 has a flexible transmembrane domain with a binding site for cholesterol [14]. It is now known

Abstract

Alzheimer’s disease (AD) has been identified as central nervous system pathology more than 100 years ago and for a long time the diagnostic criteria remained the same, based on the anatomopathological findings. Failure of clinical trials in almost every field of AD therapy forced a widening of perspective on molecular pathologies, to the point which, now, AD is considered a multifactorial disease. Recent advancement in AD cell biology uncovered new data regarding amyloid precursor protein and its enzymatic cleavage products, the amyloid beta peptides, such as oligomerization and membrane pore formation. More intracellular deregulated events have been highlighted, e.g. endoplasmic reticulum stress and mitochondrial dysfunction, possibly related through the newly discovered Mitochondria-Associated Endoplasmic Reticulum Membrane (MAM). During the last few years, non-neuronal cell populations came into focus in AD research, such as glial cells, endothelial cells forming the blood-brain barrier or brain non-neuronal stem cells of mesenchymal nature. All these cellular players interact or react to what was considered the central dogma of AD — the amyloid cascade — to the point which this dogma is about to be overthrown.

Keywords: Alzheimer’s disease; Amyloid pores; Endoplasmic reticulum stress; Mitochondrial dysfunction; Blood-brain barrier; Brain mesenchymal stem cells

Intracellular Pathogenic Links

Amyloid cascade and tau hyperphosphorylation- a 5 years update

Amyloid cascade: Amyloid cascade is a two-step enzymatic cleavage, initiated normally by a class of proteases (ADAM), also called α secretases, that shed the extracellular domain of APP and generate a C-terminal fragment, further cleaved by a gamma-secretase complex, with presenilins as main enzymatic component. In AD, a β secretase cleaves the extracellular domain of APP, generating the C99 fragment and creating the premises to yield Aβ peptides 40 and 42, with the latter exhibiting high propensity to aggregate in the extracellular space and create amyloid plaques. Besides these two main Aβ species, several other truncated species have been identified Cerebrospinal Fluid (CSF), plasma or interstitial fluid [13]. Very recently, through nuclear magnetic resonance and electron paramagnetic resonance spectroscopy, it was shown that C99 has a flexible transmembrane domain with a binding site for cholesterol [14]. It is now known

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that amyloid peptides are detrimental to synaptic activity even in oligomeric forms [15], or intracellular deposits [16]. Up to this moment, all three classes of secretase are fairly well described along with the secretory pathway, alternative enzymatic cleaving sites and subsequent peptide products. Cleavage occurs with optimum efficiency in the endosomal compartment, characterized by a lower pH than the rest of cellular compartments.

**Amyloid precursor protein and cholesterol metabolism:** Although the relationship between amyloid plaque and cholesterol has been long reported, only recently, functional cholesterol-binding domains in several amyloidogenic proteins have been identified. For APP, although cholesterol binding domains are found in both C99 and Aβ peptides, they are only partially overlapping, leading to greater affinity of cholesterol for Aβ. Furthermore, cholesterol binding enables Aβ peptides to form membrane pores by oligomerization. This data offered the perspective of an original therapeutic strategy using cholesterol competing-drugs, such as bezarotene for the treatment of Alzheimer’s and other neurodegenerative diseases that involve cholesterol-dependent toxic oligomers [17].

But not only cholesterol and apolipo proteins are shown to influence Aβ deposition and cognition, recently other players involved in cholesterol metabolisms were studied and reported to exert varia influences on AD pathology.

Hemizygosity of ATP-binding cassette transporter A1 (ABCA1) transporter (that regulates cholesterol efflux and formation of High-Density Lipoprotein (HDL)) increases Aβ deposition, but only when associated with ApoE4 phenotype [18].

In turn, overexpression of human apoA-1 in the circulation prevents learning and memory deficits in APP/PS1 mice, but apparently without changing the burden of Aβ deposition in the brain. The protective effect was supposed due partly to anti neuroinflammatory effect and diminished cerebral amyloid angiopathy [19].

**Hormonal signaling:** AD has become today the “diabetes of the brain”, or type 3 diabetes, due to its well characterized resistance to insulin signaling, accompanied by IGF-1 resistance and closely associated with IRS-1 dysfunction, potentially triggered by Aβ oligomers [20]. “Resistance to insulin” in AD brain has grossomodo the same significance as in diabetes – defective downstream insulin signaling, in spite of sufficient hormone levels. Mechanisms responsible for brain insulin resistance, reported so far are: i) increased IGF-1R levels with aberrant distribution in AD cortex [21]; ii) phosphorylation of insulin receptor substrate 1 and 2 (IRS 1/2) [22], acting a negative feedback exerted by various kinases (ERK2, glycogensynthase kinase–3 (GSK-3), mammalian target of rapamycin/S6K1 (mTOR/S6K1)and certain isoforms of Protein Kinase C (PKC)) and from feed-forward inhibition exerted by Iκβ kinase β (IKKβ) and JNK1/2/20; iii) reduced cytosolic and/or membranous levels of PI3K [23]; iv) reduced GLUT-1 expression at blood-brain barrier in AD patients [24].

In turn, early hyperinsulinemia is enough to exacerbate AD pathology observed in APP/PS1 mice [25]. Intranasal administration of insulin seems to improve cognition in both healthy and MCI patients [26].

Circulating leptin was associated with a reduced incidence of dementia and AD [27], but excess Aβ can potentially lead to a pathologically low leptin state, early in the disease process, that progressively worsens as the amyloid burden increases [28].

**Anti-pathogen activity of Aβ:** Possible roles played by pathogens in AD were repeatedly reported, from viral [29,30] to bacterial [31] and fungal [32] infections, as well as in other neurodegenerative diseases, such as Parkinson’s Disease and amyotrophic lateral sclerosis [33]. Most notably, recent evidence indicate the herpes simplex virus-1 to be a strong presence in the pathogenesis of AD, leading to generation of multiple APP fragments with neurotoxic potentials [34], induces accumulation of intracellular Ca(2+) and downstream signaling to Ca(2+)-dependent APP phosphorylation and intracellular accumulation of Aβ42 [35] and abnormally hyperphosphorylated tau [36].

In 2010, Socisa et al proposed Aβ as an antimicrobial peptide and proved it to have antimicrobial effect in vitro, comparable with LL-37, an archetypical human antimicrobial peptide. The authors also showed that brain homogenates of AD patients have higher antimicrobial activity than aged matched non-AD samples and that antimicrobial action correlates with tissue Abeta levels [37]. Further on, other peptides generated by enzymatic cleavage from the extracellular domain of APP exert antimicrobial effects in vitro against Gram-negative and Gram-positive bacteria, putatively via a membrane permeabilising activity [38]. Aβ42 was also shown to reduce uptake of IAV by epithelial cells and appeared to possess direct antiviral effect, possibly by promoting viral aggregation and further helping of phagocytic viral clearance [39].

**Tau hyperphosphorylation:** The second main pathogenic link involved in AD is hyperphosphorylation of tau – a microtubule associated protein, involved in cytoskeleton stability. Unlike APP, able to induce by itself a complete AD phenotype (both amyloid plaques and tau hyperphosphorylation) when overexpressed in animal models, abnormal tau phosphorylation leads to tautopathies. In AD tau pathology is most likely secondary to amyloid cascade, the main link between the two being glycogen synthase kinase 3, activated by the Aβ peptides via insulin and Wnt signaling pathways [40].

Intraneuronal interaction between Aβ peptides and tau has been proposed more than 5 years ago to occur on multiple tau peptides, especially those in exons 7 and 9, enhancing tau phosphorylation by GSK3beta [41] and reconfirmed lately in postmortem brains from AD patients at different stages of disease progression and animal models [42]. Furthermore, recent evidence suggests that tau protein may mediate amyloid-β peptide (Aβ) toxicity by modulating the tyrosine kinase Fyn [43]. Animal models with a heterozygous Fyn phenotype showed increased soluble Aβ accumulation and worsened spatial learning in the absence of changes in tau phosphorylation [44]. Tau-Fyn interaction has been recently investigated in order to map Fyn binding site on Tau and target interaction inhibitors by high-throughput screening [45]. Phosphorylation of tau is also influenced by Cdk5, a cyclin kinase which, unlike its other family members is not involved in cell cycle progression but in phosphorylation of cytoskeletal proteins and synaptic formation. In AD was reported as one of the major players causing aberrant hyperphosphorylation of tau through phosphorylation of specific repeats in neurofilaments of
heavy and medium molecular weight [46]. Silencing of CDK5 reduces the phosphorylation of tau in primary neuronal cultures and in the brain of wild-type mice [47]. Recent data report, however, that there is a possibility that a therapeutic effect would be attained only by dual kinase inhibition [48].

Another recent input on tau biology is the relationship between the Unfolded Protein Response (UPR) and early tau phosphorylation [49] as well as tau involvement in mitochondrial dysfunction [50]. Truncated tau induced significant mitochondrial fragmentation in neurons and when combined with Aβ at sublethal concentrations, increased the levels of oxidative stress. It also enhanced Aβ-induced mitochondrial potential loss in primary neurons [51].

Tau phosphorylation seems to be modulated also by neurotransmitters input, as specific serotogenic denervation increased tau phosphorylation in denervated cortex, without affecting amyloid-beta (Aβ) pathology [52].

**Posttranslational control of main players in AD by microRNAs**

MicroRNAs are small non-coding RNAs, than can pair, perfectly or imperfectly, with the 3' untranslated region (UTR) of different mRNAs. A perfect complementarity between the mRNA and miRNA will lead to mRNA degradation, while an imperfect match will stop the protein translation. Either way, microRNAs act as downregulators of protein synthesis, for one, several or, in some cases, tens of mRNAs. It is also possible for one mRNA to be targeted by several different microRNAs that regulate its expression at different time points in the life of the cell, or under different environmental conditions.

As this posttranslational regulatory pathway has been identified no more than two decades ago, the bulk of data regarding the main actors in AD pathology (APP, beta and gamma- secretases) regulated by microRNA is no older than 5-10 years. The research in the field of microRNA involvement in AD pathology is expanding, along with discovery of new microRNA sequences. Up-to-date, in one microRNA data base (http://www.mirbase.org) are listed over 1800 sequences of identified microRNAs in humans, out of which many species or clusters are expressed in the central nervous system in a both time-dependent manner [53] and cell-specific manner [54]. Many of microRNAs expressed in the human brain are not conserved between different families of primates, suggesting that, phylogenetically, they are a recent acquisition [55].

From the full range of miAR Nuri identified in the brain, so far only a few have been associated with neuro degenerative processes: miR-133b [56], miR433 [57] in Parkinson’s disease, miR-9 in Huntington’s disease [58,59], miR-132, miR-124a, miR-125b, miR-107, miR-219 andmiR-128 in Alzheimer’s disease [60-62].

One of the first reports of a miRNA profile modification in patients with AD belongs to Walter Lukiew, studying human hippocampus from fetal and adult patients with AD. The results indicate the abnormal expression of miR-9, miR-125b andmiR-128 in the hippocampus of AD brains [61]. Another study by Wang W X et al. On AD patients’ brains, compared with control subjects without cognitive impairment, showed a marked decrease of miARN-107, even in the early stages. Wang et al. demonstrated that the 3'-UTR sequence of BACE1 mRNA is a possible site of attachment of them iARN-107, regulating the expression of the enzyme. In addition to the decrease observed in the cortex of the temporal, the same trend was observed in the motor cortex. These changes indicate a global change miR-107 levels in the whole brain, even in area sun affected by AD pathology [63]. The 3' UTR of mRNA of BACE1 is also targeted by miRNA-29a/b-1cluster, as decreased expression of these microRNAs was demonstrated in patients with sporadic AD. Consistent with the literature [60,64-68] an increase of BACE1 expression was demonstrated through quantitative RT-PCR. Decreased expression miR-29a/b-1 and increased BACE1 is not specific to a particular cortical area, as demonstrated by cerebellum assessment [69]. Cogswell metal, Quantified the expression of over 300 mRNA sites in the hippocampus, medial front algyrus and cerebellum taken from different stages of AD compared with age-matched controls. These data show modifications of certain species of miRNA – out of which miR-9 and miR-132 were repressed in the hippocampus and frontal gyrus, correlating with the progress and location of pathological lesions. In addition they report detecting miRNAs in the CSF, with different levels from controls [70]. Over expression of miR-125b causes tau hyperphosphorylation and targets the phosphatases DUSP6 and PPP1CA and the anti-apoptotic factor Bcl-W [71], whereas miR-922 increased the levels of phosphorylated tau by regulating ubiquitin carboxy-terminal hydrolase L1 [72].

Recently, other species of miRNAs are found dysregulated in animal models of AD, related not to amyloid cascade but to other pathogenic links such as synaptic plasticity (upregulation of miR-181 [73]), or immune-related (miR-155, upregulated simultaneously with an increase of microglia and astrocyte activation [74]).

**Mitochondrial dysfunction**

Alteration of mitochondrial metabolism in AD patients has been well documented in the literature and confirmed in vitro studies showing that Aβ affects mitochondrial DNA and proteins, leading to impairments of the Electronic Transport Chain (ETC) and ultimately mitochondrial dysfunction [75]. In a triple transgenic mouse AD model, mitochondrial dysfunction was detectable from embryonic stage, continues throughout the reproductive period and is exacerbated during reproductive senescence, unlike control wild type littersmates, in which oxidative stress and a significant decline in mitochondrial function was demonstrated only with reproductive senescence [76]. A significant decrease in mitochondrial membrane potential was also noted in two cell models of AD, also accompanied by a decrease in ATP synthesis [77]. Interesting results have also been reported in selected nerve cell lines [78], along with already consecrated in vitro neuronal models PC-12 [79] and SHSY-5Y [80] cell lines.

The relationship between mitochondrial dysfunctions and other pathogenic links in AD was under intense scrutiny, most efforts concentrated on relationship between this organelle and aberrant APP processing. Correlating with the newly demonstrated ability of Aβ to oligomerize at cell membrane site, increasing evidence suggested that both APP and Aβ peptides accumulate also in mitochondrial membranes, forming pores that increase permeability and further promote the excess accumulation of Ca(2+) [81]. Both full length APP and C99 were shown to target to the mitoplast (inner membrane and matrix compartments) in brains of an AD transgenic...
mice, which seemed to be almost completely dependent on BACE 1 activity [82]. As for Aβ peptides, they were found at mitoplast level in both monomeric and oligomeric forms, even as early as 2 months of age in transgenic AD mice, generating free radicals, ultimately leading to oxidative damage [75]. Mass spectrometry studies identified heat shock protein 60 (HSP60) to be responsible for Swedish mutated APP (KM670/671NL) translocation to mitochondrial matrix, along with critical components of gamma-secretase complex [83].

Studies on the integrity of the inner mitochondrial membranes and functional status of electron transfer complexes reported that APP treatment negatively influences the complex IV activity, while the activities of complexes I and II did not change. Furthermore, activity of complex III was significantly enhanced in APP expressing cells, as compensatory response, in order to balance the defect of complex IV, compensatory mechanisms that was, however, unable to prevent the strong impairment of total mitochondrial electron transport chain [84].

Another pathogenic interaction proposed recently to explain the mitochondrial dysfunction under AD burden was upregulation of voltage-dependent anion channel 1 protein (VDAC1) levels, found in the cortical tissues from the brains of patients with AD, relative to controls, as well as in transgenic laboratory mice [85].

The association between mutant APP and mitochondrial dysfunction in not cell-type specific, as it has been very recently reported to also occur in the striated muscle fibres of a transgenic animal used as an AD model [86].

Last, but not least, mitochondrial dynamics have also been reported to be altered in AD, with a shift of balance towards organelle fission and decreased mitochondrial anterograde movement, to accompany the defective functions [87]. Furthermore, downregulation of Akt signaling, also possibly related to insulin resistance in AD, was shown to diminish mitochondrial biogenesis, with subsequent memory impairment in laboratory animals [88]. Another pathologic link relating mitochondria and AD is Aβ-binding alcohol dehydrogenase (ABAD) - a direct molecular link from Aβ to mitochondrial toxicity. Aβ was shown to interact with ABAD in the mitochondria of AD patients and transgenic mice overexpressing ABAD, with the latter manifesting exaggerated neuronal oxidative stress and impaired memory in an Aβ-rich environment [89].

**Endoplasmic reticulum stress in AD**

Endoplasmic Reticulum (ER) stress is manifested by several functional dysregulations such as calcium release or accumulation of unfolded proteins in the lumen of the organelle. This loss of homeostasis is sensed by specific transmembrane proteins that trigger the Unfolded Protein Response (UPR) - three different signaling pathway that regulate gene transcription to increase chaperone levels into assisting ER to properly fold proteins, or even to downregulate production of mRNA of unfolded proteins [90] (Figure 1).

At first, endoplasmic reticulum was related to AD pathogenesis through mutated presenilins and perturbed calcium release [91]. Soon enough was shown that mutated PS-1 downregulates the Unfolded Protein Response (UPR) signaling in ER, possibly through inhibited activation of stress transducers at the ER membrane [92]. In sporadic AD experimental data showed that the aberrant splicing isoform of PS-2 gene, generated by exon 5 skipping, is responsible for downregulation of the signaling pathway of the UPR, in a similar fashion to that reported for mutants of PS1 linked to familial AD. Prolonged ER stress leads ultimately to apoptosis, which in AD neurons was mediated by caspase-4 [93].

The relationship between ER stress and pathologic accumulation of Aβ peptides and hyperphosphorylated tau protein in the AD brain is still under study, as controversial results have been so far reported by different group studies (reviewed in [94]). ER stress triggered by Aβ was shown to promote cholesterol synthesis and mitochondrial cholesterol influx, resulting in mitochondrial glutathione depletion [95]. Sustained activation of the endoplasmic Reticulum (ER) stress response by amyloid-beta (Aβ) peptide was shown to induce apoptotic cell death not only in neurons but also in brain endothelial cells [96]. Downregulation of ER membrane transducers was confirmed in AD human brains and an interesting hypothesis was recently proposed, that linked one of this transducers with the expression of an α secretase (ADAM10) involved in non-amyloidogenic processing of APP [97].

Not only APP seems to be linked to ER stress, but also tau protein and its level of hyperphosphorylation. A strong association between the presence of UPR activation markers and p-tau was observed in the hippocampus of front temporal dementia cases, occurring independently from Aβ deposits [98]. In turn, it was proposed that tau increased the levels of ubiquitinated proteins in the brain and triggered activation of UPR by interfering with protein quality control in the ER. Increased levels of ubiquitinated proteins were accompanied by increased levels of UPR activation [99].

**Non-Neuronal Cells Involved in AD Pathogenesis**

**Microglial activation and oxidative stress**

Based on current data one cannot establish whether inflammation is a cause, a promoter, or a secondary phenomenon in AD [100], but with accumulating evidence for oxidative stress as an important pathogenic factor in AD, theories have emerged speculating that it is involved in the initiation of the disease. In fact, oxidative damage was found to be one of the first events in the disease progression [101], it has been highlighted in MCI and AD patients and it gets more intense as the disease advances [102]. An increase in microglial activation has...
been observed in very early stages of AD, with a number of studies reporting aggregation of activated microglia around amyloid plaques in animal [103] and human brain [104-106] and, interestingly, it does not seem to be preserved over time [107]. Inflammation-related gene profiling in AD indicate microglia to be in an alternative, "reconstructive activation state, rather than the classical, destructive state" [108], but in the complex environment of neuroinflammation, there is not a clear cut between "destructive" and "regenerative" actions of microglial subpopulations. The neuroprotective effects of activated microglia in inflammatory environments are partially overlapping its phagocytic, destructive ones and may be triggered by signalling molecules produced by apoptotic neurons [109].

Like most of AD pathogenic links, there seems to be a dual relationship between reactive oxygen species and Aβ accumulation. Aβ-stimulated microglia produce and secrete proinflammatory molecules and neurotoxic factors [110], such as IL-1α, IL-8 and TNFα, that were proposed as biomarkers able to distinguish AD from controls [111,112]. In addition, certain peptide fragments (such as, for example, Aβ 25-35) have the intrinsic capacity of peroxidizing membrane lipids and generate typical products such as 4-hidroxinonenal (HNE) [113] – aldehyde with proven ability to interfere with membrane ATPase and ion channels, including those involved in calcium homeostasis. Furthermore, HNE is involved in altering the metabolism of glucose via neuronal GLUT-3 transporter and the GLUT-1 astrocytic isoform. Impaired glucose metabolism is proven by studies FDG-PET (18F-2-fluoro-deoxy-D-glucose) and is a common denominator for all changes in cognition from MCI and vascular dementia up to AD. An interesting correlation can be made with the results of PET studies in MCI and patients with possible or probable AD, which showed a decrease in glucose transport into nerve tissue, with hippocampal deficit level as a predictor of evolution to AD [114]. In turn, hydroxyl radicals can react with Aβ, triggering enhanced oligomerization and aggregation [115].

Astrocytes

Astrocytes are the most numerous cells in the CNS, involved in maintenance of neuronal homeostasis. They have been proposed to be part of the proper synaptic signaling ( in the so called tripartite synapse [116]). In vitro, human astrocytes are highly sensitive to oxidative stress and trigger a senescence program when faced with multiple types of stress. One study showed that the frontal cortex of AD patients harbored a significantly greater burden of senescent astrocytes compared with non-AD age-matched controls. These astrocytes grown in vitro released a number of inflammatory cytokines and were, in turn, stimulated into senescence by Aβ (1-42) [117]. Aβ treatment is directly able to induce intracellular calcium transients and spontaneous intercellular calcium waves in isolated astrocytes in purified cultures, even at low concentrations [118], inducing alterations in astrocyte cooperation, which is known to occur through connexins-organizing gap junctions. In the brain of AD transgenic mice, an overexpression of connexins 30 and 43 were observed around amyloid plaques [119], supporting the abnormal activation of astrocytic signaling in AD. Conversely, astrocytes appear to be involved in the clearance of Aβ (1-42) [120].

Astrocytes, along with oligodendrocytes are also a source of APP [121] and inflammatory cytokines increase levels of endogenous BACE1, APP, and Aβ and stimulate amyloidogenic APP processing in astrocytes and neurons (Figure 2). The same effect was observed under oligomeric and fibrillar Aβ 42 treatment [122], creating a vicious circle in the AD brain.

**Endothelial cells and blood-brain barrier**

BBB impairment is the common denominator of many neurological disorders, whether purely neurological, or systemic. Alzheimer’s disease is not an exception and BBB alteration hypothesis is emerged from observing Congo Red positive amyloid deposits in cerebral microvasculature [123].

One hypothesis suggests that increased production of Alzheimer’s disease peptide exceeds the capabilities of clearance of nervous tissue, including transportation through the BBB. The CSF of healthy individuals contains picograms of Aβ peptides, which are increased in AD patients, especially the 42 amino acid form, which seems to confirm the existence of an alteration of the BBB or at least a lack of transport through the BBB. Studies in patients with AD showed an increase in Aβ peptides bound to lipoproteins and serum proteins, what might indicate a correlation between brain deposits and the circulating peptides distribution. The importance of the blood stream as a source of Aβ peptides is highlighted by studies of transgenic mice in which the treatment with a peptide chelating agent would reduce the plasma and cerebral amyloid load, by virtue of the equilibrium maintained between the two compartments by specific transport mechanisms [124].

Altered BBB permeability holds, from this perspective, a major role in disrupting the blood-brain circuit of the neurotoxic peptide Aβ42. Amyloid precursor protein exists as a membrane receptor in circulating blood cells and blood platelets membrane as well as in soluble circulating form. Opening BBB initiates pathological parenchymal amyloid deposition and neuro toxic mechanisms that would result in the destruction of neurons sensitive to ischemia, mostly hippocampus granular neurons. Is chemicejury, leading to BBB breakage, is associated with increased expression of β-secretase, which tilts the balance in favor of the amyloidogenic pathway. This result is associated with the demonstrated accumulation of the Ct of the
APP fragments in cell-free amyloid plaques, in animal models of focal cerebral infarction [125].

Aβ is cleared through the BBB using a pair of transporters: RAGE (receptor for advanced glycation end products) and the soluble LRP1 (LDL Receptor-related Protein1), which under normal conditions operates as a chaperon protein for the Aβ peptide in plasma and is responsible for its transport to the liver where it is metabolized. In patients with AD, LRP1 is found in oxidized form, which alters its binding ability. Transport rate is influenced by ApoE, that correlates with increased risk of AD in carriers of isoform ε4. It has already been shown that ApoE plays a role as Aβ peptide chaperone and carrier so fallele 2 and 3 have a rate of transport through the BBB significantly higher than the allele 4 carriers [124,126]. The transport can also be increased by neuroinflammatory cytokines TNF and IL-6, which contribute to the inflammatory micro-environment and participate at the same time to increased BBB permeability. Another possible modulator of transport by RAGE/LRP1 is the endothelial NO synthase, whose level is reported to be low in the brains of AD patients [127].

Another clearance mechanism at the BBB involves glycoprotein P (Fgg) [128], which seems to be decreased in amyloid angiopathy at endo the lial level. Although considered in conclusive, there were reports that the inverse relationship between Fgg expression and amyloid deposition is related particularly to Aβ40 [129].

Kinetic analysis of Aβ distribution across the mouse BBB showed that most of Aβ40 is cleared across BBB. Clearance rates differ, however, between mouse and human models, but the ratio between transport and local degradation is maintained in favour of BBB clearance [130].

AD endothelial dysfunction has been reported in vascular defects of organization at the microscopic level-increased amount of collagen IV, laminin and proteoglycans in the vascular basal lamina, reduced vascular bed [131]. Vascular changes in the AD brain are also reflected in aberrant angiogenesis. Vessels of neoformation, fenestrated, with more permeable tight junctions, morphological abnormalities and changes in endo the lial basal lamina were described in the brain of AD patients. The diameter of vascular branch in gand abnormal basal lamina abnormalities have been reported in animal models of AD brain by Nakajima [132] and Meyer [133]. Also demonstrated in the brains of AD patients, over production of angiogenic mediators (Angiopoietin-2 and VEGF) may contribute to vessel formation. VEGF is produced by astrocytes in response to hypoxia, which, as mentioned above, it has been proposed to link pathogenic and the decrement in NPC number was correlated with accumulation of Aβ, even in oligomeric, diffusible form [145]. Although Kolecki et al. confirmed the previous results, they reported that overexpressing APP and Aβ in transgenic mice do not interfere with the mitotic activity of NPC, as assessed by Ki-67 [146]. Other reports showed that fibrillar Aβ-42 seems to be involved in hindering neurogenesis either by generating an inappropriate environment for neuroblasts to mature, or inducing senescence of neuronal stem or progenitor cells [147]. Exposure of NSCs to amyloid-β oligomers lowers their dividing potential, favours gliogenesis and attenuates mobility [148].

Neural stem cells and progenitor cells can be cultured from rapid autopsy samples of SVZ from elderly human subjects, including patients with age-related neurologic disorders and maintained in vitro to generate neurospheres [149]. Interestingly, the percentages of β-tubulin (III+) cells differentiated from AD and control SVZ neurospheres using a neuron-promoting differentiation protocol were equivalent, promoting the idea that committing abilities of NSC are not modified by the course of the disease.

Brain mesenchymal stem cells

Although initially described as a distinct subpopulation in the haematopoietic bone marrow stroma, Mesenchymal Stem Cells (MSC) have been also identified in dermis, adipose tissue, skeletal muscle and tendons [150] and recently, in the brain, in relationship with brain vasculature. Brain mesenchymal stem cells express mesenchymal (CD105, CD13) and pericyte markers (PDGFR-β) along with, α-SMA, NG2, CD13, CD49d, CD73, CD90, CD106 and RGS5.
They are multipotent, showing adipocytic, chondrocytic and osteocytic activity, under appropriated stimuli, but also, when exposed to glial induction medium, were able to differentiate into oligodendrocytes or astrocytes. They are also able to generate an immature neuronal phenotype [151].

One MSC particular feature is autophagy regulatory digestion pathway for aggregated proteins and organelles that cause various neurodegenerative diseases, such as a synuclein in PD or tau deposits in tauopathies and AD. Unlike NSCs, MSCs possess immunomodulatory functions, are able to influence activation of microglia to an alternative phenotype, as part of the repair process and extracellular matrix reorganization, not accompanied by production of pro-inflammatory cytokines [152], increase microglial-related Aβ clearance, reduce tau hyperphosphorylation and improve spatial learning and memory impairments in MSC transplanted AD animal models [153].

Future perspectives

The intricacies of AD pathological links seem to be increasing, as the perspectives widens in attempt to a better understanding of the disease. The deeper one looks inside the cell, more correlations will be brought to surface, out of which the significant, disease-modifying ones have to be highlighted. Therapeutic pipelines still address spatial learning and memory impairments in MSC transplanted AD animal models [153].

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References


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