

## Review Article

# Relationship between Mast Cells and Autoimmune Diseases

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**Abstract**

Mast Cells (MCs) are no longer considered as only effectors cell in allergic disorders, but also as important modulators of innate and adaptive immune responses. Increased numbers of MCs together with signs of their degranulation at sites of inflammation and tissue injury have been shown in studies of both human disease and animal models of autoimmune disorders. Despite this substantial evidence, MCs role in autoimmunity is still on debate. MCs can regulate the recruitment, survival and function of many immune cells. Therefore, they are able to enhance or suppress the initiation, magnitude and/or continuance of immune responses in autoimmune conditions. Further studies are essential to gain a more detailed understanding of MCs participation in autoimmunity. Targeting MC may be of value in future prevention and treatment of autoimmune diseases.

**Keywords:** Mast cells; Autoimmune diseases

## Autoimmune Diseases

Autoimmune diseases could be defined by inadequate or exacerbate excessive immune response against self-antigens. In this condition chronic diseases arise due the breakdown of host immune tolerance. The etiology of autoimmune disease is not clearly understood, but both genetic and environmental factors are implicated in autoimmune disease development [1,2]. Within many environmental factors described as a trigger of autoimmunity, we can detach chemical agents, drugs, UV radiation, however infectious agents could be considered as one of the main risk factors to autoimmunity pathology development [3]. A possible mechanism whereby infectious agents trigger autoimmunity is known as molecular mimicry. According to this concept microorganisms have molecular structure that cross-reacted with endogenous molecules [4]. Despite being considered a “rare” disease, the worldwide prevalence of autoimmune disease is around 5% and, curiously, of this total more than 70% are women. This elevated prevalence between women is attributed, at least in part, to hormonal changes throughout woman life [5]. Although each autoimmune disease shows specific characteristics depending on the affected tissue, all of them share a common pathologic mechanism, indicating a similar origin. The same environment and genetic inductors and also the same immune mediators of pathogenesis such as autoantibodies, leukocyte infiltrate and inflammatory pathway response are present in different autoimmune diseases context [6]. Activation of T and B cells plays an important role both in physiologic states and in response to pathogens but sometimes could trigger diseases, such as allergy and autoimmune diseases [3,7]. The organism has mechanisms that regulate all steps of lymphocytes activation and reduces deleterious effects possibility even in utero [8]. Tolerance is the major mechanism by which immune system control pathologic effects of T and B cells activation. Basically, tolerance acts by two ways: denominated central and peripheral tolerance. In the central tolerance, auto-reactive lymphocyte clones are deleted in the thymus.

In the case of peripheral tolerance, mature reactive clones cells are regulated by suppression or anergy mechanisms such as cytokines and different type of suppressive immune cells, wherein Treg cells have a pivotal role [9-11]. T-cell Receptors (TCRs) and B-cell Receptors (BCRs) recognize peptides presented by major histocompatibility complex molecules (MHC or HLA) to discriminate between self and non- self antigens. The V (D) J segments rearrange is a process that allows a wide variety of antigens to be recognized by the TCR and BCR. However, in some circumstances checkpoints in lymphocyte development fail and TCRs and BCRs self-reactive cells escape from tolerance [12,13]. MHC class I and II alleles also have been associated with increased susceptibility to autoimmune disorder, such as celiac disease, type 1 diabetes and rheumatoid arthritis [14,15].

In recent years, there has been demonstrated that innate immunity components also contribute for autoimmune diseases development. Toll-Like Receptors (TLRs), NOD-like receptors pathway and other Pattern Recognition Receptors (PRRs) are involved directly in the promotion of inflammatory milieu associate to autoimmune pathology, such as recruitment and activation of dendritic cells, macrophages, neutrophils and MCs that produce several amplifiers of autoimmunity [16]. Overall, both innate and adaptive immunity are mediators of complex mechanisms of autoimmune disease.

## Mast cell biology

MCs arise from CD34-positive hematopoietic progenitors in the bone marrow and are released into the circulation, from where they migrate to vascularized tissues or serosae cavities. MCs undergo terminal stages of their differentiation and/or maturation under the influence of microenvironmental conditions and through the action of Stem Cell Factor (SCF). In addition, cytokines such as interleukin IL-3, IL-4, IL-9, IL-10, Nerve Growth Factor (NGF), some chemokines and retinoid acid can regulate MCs differentiation [17]. MCs are widely distributed throughout vascularized tissues. On the basis of their location and protease content, MCs are subdivided

in two major subtypes in rodents: Mucosal MCs (MMC) that are associated with the epithelium of the lung and gastrointestinal tract, and which express chymases mMCP-1 and mMCP-2. The other subtype comprises the Connective Tissue MCs (CTMCs) that are found in the intestinal submucosa, peritoneum and skin. CTMCs are characterized by the expression of the chymase mMCP-4, an elastolytic enzyme (mMCP-5) and two tryptases (mMCP-6 and mMCP-7) as well as carboxypeptidase 3 (CPA3) [18]. In humans, MCs are classified as either MC<sub>T</sub>, which express tryptase only, or MC<sub>TC</sub>, which express tryptase, chymase and CPA3 [19]. The major mechanism of MCs activation is through antigen- and IgE-dependent aggregation of the high affinity IgE receptor, FcεRI, that triggers MCs degranulation [20]. However, MCs can also be differentially activated by complement system fractions (anaphylatoxins), immunoglobulin free light chains, hormones, neuropeptides and Toll Like Receptors (TLR) [21]. Thus, MCs can respond to different stimuli, and thereby participate in a wide variety of physiological and pathological processes. MCs activation can induce the release of various biologically active products. These include pre-formed molecules such as histamine, serotonin, TNF-α, kinins and proteases stored in secretory granules. Leukotrienes (LT), prostaglandins and Platelet Activated Factor (PAF) are synthesized during MCs activation from arachidonic acid. In addition, a number of cytokines (IL-1, 2, 5, 6, 8, 9, 13, 17, TNF and TGF-β1) chemokines (CCL1, CCL2, CCL3, CCL3L1, CCL4, CCL5, CCL7, CCL8, CCL11 and CXCL2) and growth factors (VEGF, PDGF, bFGF, EGF, IGF-1 and NGF) are synthesized *de novo* and released several hours after their stimulation [22]. MCs are no longer recognized only as eliciting allergy, but also as having many homeostatic functions, such as blood flow and coagulation, smooth-muscle contraction and peristalsis of the intestine, mucosal secretion, wound healing, as well as, regulation of innate and adaptive immune responses, peripheral tolerance and autoimmunity. MCs can regulate many aspects of the biology (recruitment, survival, development, phenotype or function) of immune cells, including granulocytes, monocytes/macrophages, dendritic cells, T cells, B cells, NKT cells and natural killer cells. Therefore, MCs are able to enhance or suppress the initiation, magnitude and/or maintenance of immune responses, inclusively autoimmunity [23].

### Mast cells crosstalk with other immune cells

Dendritic Cells (DCs) are likely to be one of the earliest target cells of MCs influence. These cells co-localize in most tissues, particularly at sites of Antigen (Ag) entry. Many lines of evidence have linked MCs or their products to the regulation of DC migration, maturation and function. MCs promote DCs migration by the release of TNF-α, IL-6 and IL-1β [24,25]. Indeed, MCs can induce the selective mobilization of specific DCs subsets [26]. By regulating DCs maturation, MCs are able to influence the ability of these cells to direct the quality of Th cell differentiation. Co-culture of activated MCs with DCs results in their maturation, demonstrated by increased expression of CD80 and CD86 [27]. Histamine has profound effects on DCs, which express all four of its receptors (H1-H4) [28,29]. *In vitro* data showed that histamine induces CD86 expression and chemokine production by immature DCs [27,28]. Besides preformed cytokines, metabolites from arachidonic acid and proteases, MCs granules also contain membrane vesicles termed exosomes [30]. MCs derived-exosomes were shown to induce immature DC to up-regulate MHC

class II, CD80, CD86, and CD40 molecules and to acquire a potent Ag presenting capacity to T cells [31]. MC-primed DCs stimulate CD4+ T cells to release high levels of IFN-γ and IL-17, indicating that MCs may promote Th1 and Th17 responses [32]. However, MCs can also induce the downregulation of IL-12 and stimulate the production of IL-10 by DC, resulting in a decrease in expression of the Th1 cytokine, IFN-γ, and an increase in the Th2 cytokine, IL-4, by T cells primed *in vitro* by these cells [33,34]. MCs can also directly influence T cell differentiation and function in a number of ways. T cell-MC co-culture experiments demonstrate that MCs significantly enhance T cell proliferation and cytokine production. MCs are infrequently found in lymph nodes and spleen, but they can migrate to these tissues during an immune response, where they mediated T lymphocyte activation [32]. MCs may also affect T-cell recruitment to sites of inflammation by direct release of chemotactic molecules [35,36], regulation of expression of adhesion molecules and induction of cytokine release by endothelial cells [37]. MCs can activate CD4+ or CD8+ T cells through Ag presentation by either MHC class II- or class I-context implying that they can actually serve as resident APCs [38-41]. Moreover, MCs expression of co-stimulatory molecules including members of the B7 family (ICOSL, PD-L1, and PD-L2) and the TNF/TNFR families (OX40L, CD153, Fas, 4-1BB, and glucocorticoid-induced TNFR) are important in regulating T cell activation and ultimate response [42,43]. Additionally, MCs produce the major cytokines involved in T-cells activation and polarization towards Th1, Th2, Th17 or Treg subsets [44]. The histamine released by MCs can promote Th1 cell activation through H1 receptors and suppress both Th1 and Th2 cell activation through H2 receptors [45]. MCs, in addition to cell-to-cell contact and cytokine release, can also regulate T-cell function by the secretion of exosomes [46]. MCs activation by adenosine receptors triggers IL-4 and IL-13 production which, in turn, induces the synthesis of immunoglobulin E (IgE) by B lymphocytes. Moreover, MCs protease I can enhance the production of IgG1 and IgE by B cells in the presence of IL-4 or LPS [47] in rats. These cells can also regulate CD40 surface expression on unstimulated B cells and the interaction between CD40 with CD40L on MC, together with MCs-derived IL-6, induce the differentiation of B cells into CD138+ plasma cells with selective secretion of IgA [48]. It was also demonstrated that MCs upon stimulation with IL-4 secrete exosomes that induce B cells proliferation and cytokine production [46]. Finally, some MCs-derived cytokines can influence B cell development, such as IL-4, IL-5, IL-6 and IL-13.

### Mast Cells in Rheumatoid arthritis

Rheumatoid Arthritis (RA) is the most common autoimmune disease in the world. RA is a chronic inflammatory disease that affects the joints and is characterized by autoantibody production and destruction of cartilage and bone [49,50]. Despite of established adaptive immunity participation in RA, different cells of innate immunity, including MCs, has been implicated in RA progress [50-52]. Patients with RA have a higher number of MCs at synovial cavity than health individuals [53,54]. Besides, animal experimental models studies (K/B×N model of arthritis e.g.) showed that MCs produce several mediators that are implicated in RA pathogenesis [55,56]. Histamine deficient mice developed a moderate form of arthritis when compared to the wild-type controls, and the use of pharmacological antagonists of the histamine receptors showed

that H4 is the most important for development of arthritis in autoantibody-induced arthritis [57-59]. The levels of tryptase in synovial fluid are increased in RA patients and was demonstrated that tryptase has an antiapoptotic effect on RA synovial fibroblasts through the activation of Rho, inducing hyperplasia of synovial tissue [60]. Tryptase can also activate synovial cells expressing PAR-2, enhancing tissue inflammation [60,61]. A tryptase member family, hTryptase- $\beta$ , was able to activate zymogen forms of MMP-3 and MMP-13, which are constitutively present in articular cartilage [62]. MCs produce many relevant cytokines in RA context, such as TNF- $\alpha$ , IL-6, IL-1 $\beta$ , IL-17 and IL-33. The TNF- $\alpha$  importance in RA development is demonstrated by RA treatment with anti-TNF- $\alpha$  antibody. The activated MCs have an enhanced production of TNF- $\alpha$  that may activate other innate effectors cells [63-65]. IL-33, a member of IL-1 family, exacerbates arthritis by activating MCs [64,66,67]. Interesting, MCs are the major producing of IL-17 in RA synovia, opposite to what is observed in others sites where Th17 are the main IL-17 source [68-71]. The presence of autoantibodies such as rheumatoid factor and anti-citrullinated protein antibody (ACPA) is related to RA progression, since these antibodies may trigger MCs activation through Fc $\gamma$ R and Fc $\epsilon$ R that are expressed in synovial MCs [49,72,73].

### Mast cells in neurological autoimmunity

Multiple sclerosis (MS) is the most prevalent autoimmune disease of those affecting Central Nervous System (CNS), featured by inflammatory, demyelinating lesions localized in the brain and axonal loss [74,75]. In the brain, MCs reside on the brain side of the blood-brain barrier (BBB), and interact with different cells of CNS [76]. A considerable amount of literature has demonstrated that MCs are implicated in MS disease, wherein MCs are found within the demyelinated plaques, in normal white matter, and tryptase was significantly elevated in cerebrospinal fluid of MS patients [77,78]. Mouse model of Experimental Autoimmune Encephalomyelitis (EAE) have contributed to the understanding of MS development [79]. However, the observed MS functions in EAE models are very confusing. Previous studies using MCs-deficient W/W<sup>v</sup> mice showed that MCs are important for early onset and severe disease in MS. The phenomenon observed in W/W<sup>v</sup> mice model was correlated with activation of CD4 and CD8 T cell, IFN- $\gamma$  production as well as IL-4 expression and neutrophil recruitment by MCs. In this mice model all these mechanisms described are decreased when compared with wild type animals developing EAE [79-82]. In contrast to the data with W/W<sup>v</sup> mice model, Kit W-sh/W-sh mice have an increased EAE development due to absence of immune suppression and there is evidence that chymase protects from post-traumatic brain inflammation. These studies suggest that MCs play a regulator role in EAE [83-86]. There is still some works showing that MCs could not contribute to EAE, this conclusion comes from studies using W/W<sup>v</sup>, W-sh and Cre-Lox system [87,88]. More studies will be needed to assess these divergent data.

### Mast cells in inflammatory bowel disease

Inflammatory bowel diseases (IBD), Crohn's disease and ulcerative colitis are chronic noninfectious inflammations manifested in the gastrointestinal tract [89]. The IBD etiology is unknown, however we know that interaction of several factors like heredity,

environment, intestinal microbiota and immune system cells can lead to IBD development [89]. The intestinal immune system has a dual function at the same time maintain the food antigens and commensal microorganism tolerance, but also protecting against pathogenic organisms. Thus when an unbalance in the homeostasis mechanism of gastrointestinal tract arises, the host became more susceptible to IBD pathology [90,91]. A large body of literature has investigated the participation of adaptive immune system in IBD, there is an strong association between exacerbated activation of T cells and IBD progress. In this context a mix of Th1/Th17 response is increased and is deleterious for intestinal mucosa Crohn's disease and ulcerative colitis [90,92]. Nowadays, the innate immune cells also has been implicated in IBD pathology, once that macrophage, DC and other innate immune cells have been implicated in IBD [93]. MCs are present in different regions of gastrointestinal tissue and together with other cells assist in this homeostatic maintenance [94,95]. Several studies have suggested that IBD patients present an increase in MC mediators, such as tryptase, chymase and histamine [94,96,97]. Besides that, *in vitro* and *in vivo* experimental models data also shown that MCs are important inflammation sources in IBD, whereas release of proteases, PAR-2 activation pathway and TNF- $\alpha$  contribute to deleterious effects of disease [98-101]. Interesting, recent works have examined the extracellular ATP's role, a known danger signal, in mediating MCs activation in the context of intestinal inflammation. This mechanism occurs by activation of P2 purine receptors (P2X7), and this ATP-induced activation of P2X7 exacerbates MCs inflammation induced [102].

### Mast cells in Type 1 diabetes

Type 1 Diabetes (T1D) is caused by the immune-mediated destruction of insulin-producing  $\beta$  cells in the pancreas. The selective immune response to  $\beta$ -cells develops by the emergence of islet antigen-reactive T cells concomitantly to an impairment of immune regulation. Susceptibility to T1D is conferred by a combination of genetic background and environmental factors [103]. Studies in NOD mice and biobreeding (BB) rats, which develop T1D with common characteristics to the human disease [103, 104], have identified roles for several different immune cell types in  $\beta$ -cell destruction. More important, it seems that the pathogenesis of T1D involves a considerable crosstalk between these cells. CD4+ and CD8+ T cells are pivotal for T1D onset and are considered to be the final executors of  $\beta$ -cell destruction. T1D can only be transferred from diabetic NOD mice to immune compromised syngeneic recipients by a combination of both T cell subsets [105,106]. Although being the source of the autoantigens that drive T1D,  $\beta$ -cells are unable to directly prime diabetogenic CD4+ or CD8+ T cell responses, making necessary the involvement of cross-presenting APCs (B lymphocyte, macrophage and DC) for the disease to develop. NOD mice deficient in B lymphocytes or depleted from DC and macrophages are protected from the onset of T1D [107-110]. MCs could be another innate immune cell involved in T1D. MCs reside in the peri-acinar space, pancreatic interstitium and mesentery and have already being implicated to pancreas inflammation and cancer [111-114]. The existent evidence that indicate a role of MCs in T1D is somewhat indirect. Transcripts associated with MCs were differentially expressed in the Pancreatic Lymph Nodes (PLN) of prediabetic BB rats [115]. In a subsequently study, the same group showed that MCs

found in the PLN from diabetic rats presented a gene expression profile indicative of activation and that treatment with cromolyn, a MC degranulation inhibitor, delayed the onset of T1D [116]. MCs could be involved in the development of T1D by directly eliciting  $\beta$ -cell destruction or by modulating the proinflammatory response against  $\beta$ -cell antigens. Cytokines immediately released upon MCs degranulation or newly synthesized may be important mediators of  $\beta$ -cell damage. MCs produce the three major cytokines involved in T1D development, TNF- $\alpha$ , INF- $\gamma$  and IL-1 $\beta$ . Treatment of neonatal NOD mice with recombinant TNF- $\alpha$  accelerated diabetes [117]. TNF- $\alpha$  stored in MCs granules may induce  $\beta$ -cell cytotoxicity via generation of nitric oxide (NO) [118] or by increasing the ability of APCs to activate  $\beta$ -cell-specific T cells [119,120]. There are data that support a synergistically effect of both IFN- $\gamma$  and TNF- $\alpha$  in the induction of  $\beta$  cell apoptosis [121]. NOD mice bearing a null mutation of the INF- $\gamma$  receptor  $\alpha$  chain showed a marked inhibition of insulinitis and no signs of diabetes [122]. It was shown that MCs can be a source of IFN- $\gamma$ , which secretion does not depend on IgE-mediated activation [123]. *In vitro* experiments demonstrated that IL-1 $\beta$  induction of NO production [124] and ER stress response [125] activates cell death programs and modifies the expression of several genes that may affect both function, viability and  $\beta$ -cell recognition by the immune system [126]. Moreover, IL-1R deficiency slows development of diabetes in NOD mice by blocking IL-1 $\beta$ -mediated induction of iNOS by TNF- $\alpha$  and IFN- $\gamma$  [127]. MCs production of IL-1 $\beta$  may be mediated by an inflammasome-dependent mechanism. IL-1 $\beta$  produced by these cells is primarily responsible for the cutaneous inflammation in the cryopyrin-associated periodic syndromes (CAPS), caused by mutations in the NLRP3 gene that lead to overactivation of inflammasome [128]. IL-1 $\beta$  is secreted as an inactive precursor, and processing of pro-IL-1 $\beta$  depends on cleavage by proteases. Interestingly, human MCs chymase is able to rapidly and specifically convert IL-1 $\beta$  precursor in its active form [129]. In contrast to the data that indicate a pathogenic function of MCs in T1D, recently, it was documented that repeated administration of anti-Fc $\epsilon$ R1 antibodies resulted in protection against diabetes in NOD mice by a mechanism that was partially dependent on IL-4 [130]. Authors concluded that this effect was due to activation of both basophils and MCs, although they have not evaluated exactly which cells were involved in that process. Thus, the exact role of MCs in T1D development still remains to be ascertained.

### **Mast cells in Systemic lupus erythematosus (SLE)-mediated glomerulonephritis**

SLE is an autoimmune disease with manifestations derived from the involvement of multiple organs including kidneys, joints, nervous system and hematopoietic organs. The combination of genetic, hormonal and environmental factors is involved in the activation of both innate and adaptive immune responses, resulting in loss of tolerance to ubiquitous self-antigens, particularly anti-double-stranded DNA [131]. Defective clearance of cell debris results in secondary necrosis and an overload of self-antigens. In SLE patients, nuclear particles, specially double-stranded DNA, are taken as viral particles and elicit a (pseudo) antiviral immune response that involves all antigen-presenting cells, particularly DC and B cells [132]. Activation of the innate immune system results in enhanced antigen presentation to T cells and the release of proinflammatory

cytokines, including type I IFNs. The expansion of T and B cell clones with specificities for predominantly nuclear autoantigens account for the production of antinuclear antibodies, immune complexes and T cell-dependent tissue damage. Antigen-antibody complexes mediate the injury to organs and tissues by binding to complement and attracting macrophages and neutrophils [133]. Immune complexes may also bind to receptors expressed by tissue-specific cells and alter their function [132]. Lupus Nephritis (LN) occurs in approximately 50% of adult and 80% of pediatric patients with SLE [134]. In the kidneys, Immune Complexes (IC) can deposit in the mesangial area, subendothelial and subepithelial spaces or in peritubular capillaries. IC deposition induce complement- and FcR-mediated inflammatory cascades that cause activation or injury of renal resident cells, which in turn release inflammatory factors, leading to the recruitment of inflammatory cells [135]. Cytotoxic T cells, Th17 T cells, as well as B cells infiltrate the kidney in LN. Long term renal damage is caused by continuous inflammation, vascular injury by systemic and local mediators, hypoxia and fibrosis [136]. MCs numbers were shown to be increased in kidney biopsies of patients with LN [137,138]. However, it was not evaluated if MCs accumulation was involved in the development of LN or if it was merely a response to subsequent autoimmune-mediated tissue injury. It was demonstrated that MCs-deficient mice subjected to the pristane-induced model of LN developed renal disease comparable to their WT counterparts, including glomerulonephritis, immune deposits, and proteinuria [139]. Thereafter, MCs contribution to LN needs to be evaluated.

### **Mast cells in psoriasis**

Psoriasis is one of the most common chronic inflammatory skin disorders and is characterized by scaly, reddened patches, papules, and plaques derived from excessive growth of skin epithelial cells [140]. Many stressful physiologic and psychological events and environmental factors are associated with onset and disease worsening. Carriage of HLA-Cw6 [141] and environmental triggers, such as  $\beta$ -haemolytic streptococcal infections [142] are major determinants of disease expression. Psoriasis is primarily a T cell-mediated autoimmune disease, and arises by the interaction of epidermal keratinocytes and mononuclear leukocytes [141]. At the psoriasis lesion it is observed a marked infiltration of mononuclear leukocytes (T cells and DC) into the dermis and elongated/hyperplastic blood vessels in the papillary dermal region [143]. The majority of the T cells infiltrating the dermis are of the CD4+ T cells subtype, whereas CD8+ T cells predominate in the injured epidermis. CD8+ T cells are likely to be the ultimate effectors cells that recognize autoepitopes presented by the binding pockets of HLA-Cw6 or other HLA class I molecules on the keratinocyte surface [142]. There are compelling evidences of the involvement of MCs in the pathogenesis of psoriasis. MCs, particularly the MC<sub>TC</sub> subset, are enriched in the papillary dermis of psoriasis lesions in humans [144-146] and show signals of degranulation, denoted by increased interstitial levels of histamine [147], in early eruptive and recurring lesions [146]. Increased expression of SCF by keratinocytes, endothelial cells and fibroblasts may account for the accumulation and activation of MCs in plaques lesions [148]. MCs may contribute to psoriasis development by the production of cytokines. For instance, MCs in psoriatic skin are strongly positive for interferon INF- $\gamma$  [149], believed to be one of the most important mediators in the cytokine

cascade of psoriasis [150]. Moreover, most of IL-17 producing cells in normal and psoriatic skin are MCs [151], not T cells. IL-17 induces the synthesis of antimicrobial peptides (i.e., defensins, lipocalin, and S100 proteins) [152] and also regulates a group of neutrophil-attracting CXCL chemokines [153] expression by keratinocytes. This cytokine is believed to be an important effector factor in psoriasis pathogenesis [154] and blockage of its pathway has been shown to be therapeutic on psoriasis [155]. Furthermore, tryptase released upon degranulation is believed to activate keratinocytes and endothelial cell by means of PAR-2 activation [156].

### Mast cells in bullous pemphigoid

Bullous pemphigoid (BP) is an acquired autoimmune skin disease associated with an IgG autoimmune response and characterized by detachment of the epidermis from the dermis and an intense inflammatory cell infiltrate in the upper dermis. Autoantibodies from BP patients react with two hemidesmosomal components: transmembrane collagen XVII (BP180 or BPAG2) and plakin family protein BP230 (BPAG1) [157]. BP pathogenesis include complement activation, recruitment of inflammatory cells and liberation of proteolytic enzymes that cleave and degrade type XVII collagen [158]. The activation of complement seems to play a central role in this disorder since mice depleted of complement did not develop BP following injection with pathogenic rabbit anti-BP180 antibodies [159]. Autoreactive T and B cell responses to BP180 have been found in patients with BP [160] although it was shown that these lymphocytes subtypes were not necessary for the emergence of subepidermal blistering in an experimental model [161]. Besides complement activation, lesion formation in BP depends upon MCs degranulation [162] and accumulation of neutrophils [163] and eosinophils. Mice deficient in neutrophils or MCs are resistant to experimental BP [161]. MC degranulation is a common feature of BP [164,165] and precedes neutrophil infiltration and subsequent dermal-epidermal separation [162]. MCs degranulation in BP was dependent on complement activation [162]. Moreover, MCs may also be activated by FcεRI cross-linking. In addition to IgG autoantibodies, most BP patients also produce IgE class autoantibodies that also react with BP180 and total IgE levels are often elevated in the disease [166]. IgE autoantibodies were found at the basement membrane zone and coating MCs in injured skin [167]. Upon degranulation, MCs release a variety of mediators such as leukotrienes, PAF and cytokines that contribute directly or indirectly to neutrophils recruitment [162,168,169]. Activated neutrophils release proteolytic enzymes that cause the splitting of epidermis from the dermis [157]. Elastase and gelatinase B (MMP-9) were found to be crucial for neutrophils induction of blisters [170]. Interestingly, MCs chymase MCP-4 can activate MMP-9 and also directly degrade BP180 *in vitro* [171]. Mice lacking specifically this protease were resistant to experimental BP [171].

### Mast cells in vasculitis

Vasculitis comprises a diverse group of conditions characterized by inflammation and necrosis of blood vessels that leads to vessel occlusion and ischemia of tissues. Vessels of any kind in any organ can be affected, which accounts for the heterogeneity of signs and symptoms of the disease. Vasculitis may occur as a primary condition or as a component of other diseases [172] and are categorized on the basis of the predominant type of vessel (large, medium or

small) affected [173]. Small-vessel vasculitis may be caused by IC-mediated inflammation as in the case of anti-glomerular basement membrane disease (Goodpasture's syndrome), IgA vasculitis (Henoch-Schönlein purpura) and vasculitides secondary to systemic immune complex diseases such as SLE, dysproteinemias, cryoglobulinaemias, and chronic infections. Vasculitis can also be induced by antibodies directed to autoantigens like in Anti-Neutrophil Cytoplasmic Antibody (ANCA)-associated small vessel vasculitis, which includes granulomatosis with polyangiitis (originally Wegener's granulomatosis), microscopic polyangiitis, and eosinophilic granulomatosis with polyangiitis (formerly Churg-Strauss syndrome) [172]. The Anti-Neutrophil Cytoplasmic Autoantibodies encountered in ANCA-associated vasculitis are mostly directed against the neutrophil azurophilic granule proteins proteinase 3 (PR3) and Myeloperoxidase (MPO). The inoculation of mice with hybridomas producing monoclonal IgG rheumatoid factors induces the development of a leukocytoclastic skin vasculitis caused by the deposition of cryoglobulins. The binding of IC by FcR positive cells stimulates the secretion of inflammatory mediators that recruit Polymorphonuclear (PMN) cells, which damage blood vessels. In this model of hypersensitivity angiitis, vascular inflammation was dependent on MC recognition of IC by FcγRIII and release of pre-formed TNF-α stored within its granules [174]. Administration of mercuric chloride (HgCl<sub>2</sub>) to Brown Norway rats causes a Th2-induced autoimmunity characterized by high IgE concentrations and production of multiple IgG autoantibodies to MPO. Animals develop polyarthritis and a leukocytoclastic vasculitis which predominantly affects the intestine. Vasculitis in this model has some similarities to human Churg-Strauss syndrome. It has been suggested that MC play a role in the pathogenesis of early disease [175,176], which is αβ T-cell and neutrophil independent [177,178]. Nontoxic concentrations of HgCl<sub>2</sub> can induce CTMC mediator release and cause up-regulation of IL-4 mRNA expression *in vitro* [172]. Besides, HgCl<sub>2</sub> administration induced an increase of serum rat MC protease II (RMCP II) [175] and reduction of toluidine blue positively stained cells (interpreted as MC degranulation), indicating *in vivo* MC activation [176]. Moreover, the use of the MC-stabilizing agents G63 and doxantrazole resulted in amelioration of early cecal vasculitis caused by HgCl<sub>2</sub> [175]. On the other hand, MC depicted a protective role in the development of focal necrotizing glomerulonephritis (GN) in ANCA-associated vasculitis. MC-deficient Kit W-sh mice developed augmented anti-MPO GN, characterized by enhanced renal injury compared to MC-intact C57BL/6 mice. Moreover, mice reconstitution with bone marrow derived-MC attenuated glomerular injury indicated by fewer abnormal glomeruli and less fibrin deposition. It was suggested that MC migrate from sites of MPO presentation to draining LN where they could modulate T regulatory cells function through secretion of IL-10, and these cells, in turn, would decrease MPO T effector cells capacity to produce proinflammatory cytokines [179].

### Mast cells in other autoimmune conditions

MCs have been implicated in other autoimmune conditions in which were observed their accumulation in the vicinity of affected tissues. In Thyroid-Associated Ophthalmopathy (TAO), an extra thyroidal manifestation of Graves' disease, MC were among the cells that infiltrate the orbit [180-182]. It was shown a significant correlation between the numbers of MCs in minor salivary glands

**Table 1:** Overview of autoimmune disease in whose development Mast cells may be involved.

Disease	Animal model	Evidence of MC participation	Ref.
Type I Diabetes (T1D)	Spontaneous development of autoimmune insulinitis in Biobreeding rats.	Increased expression of MC transcripts in pancreatic lymph nodes from rats with T1D.	[115]
		Treatment with MC stabilizer delayed disease onset.	[116]
Systemic lupus erythematosus (SLE)		MC increased numbers in kidney biopsies of patients with lupus nephritis.	[137,138]
Psoriasis		MC enriched in the papillary dermis of psoriatic lesions in humans.	[144-146]
Bullous pemphigoid (BP)	Injection with pathogenic anti-BP180 IgG.	Degranulated MC in cutaneous lesions from patients with BP.	[164]
		Presence of histamine in the blister fluid of BP lesions.	[165]
		Mice deficient in MC or lacking mMCP-4 were resistant to experimental BP.	[161, 162, 171]
Vasculitis	Inoculation of mice with hybridomas producing monoclonal IgG rheumatoid factors.	Disease development was dependent of MC recognition of IC by FcγRIII and release of pre-formed TNF-α	[174]
	Administration of HgCl <sub>2</sub> to Brown Norway rats.	MC stabilization resulted in amelioration of early caecal vasculitis.	[175]
		Direct correlation between MC degranulation and early caecal vasculitis.	[176]
Thyroid-associated ophthalmopathy (TAO)	Genetic immunization of outbred NMRI mice with cDNA encoding the human thyrotropin receptor (TSHr)	MC were among the cells that infiltrate the orbit in patients with TAO.	[181]
		MC infiltrated the extraocular muscles from hyperthyroid animals.	[182]
Sjogren's syndrome (SS)		Association of MC presence with fibrosis and cell acid infiltration in the minor salivary glands of patients with primary SS.	[183]
Pemphigus vulgaris (PV)		Increased numbers of MC in skin lesions from patients with PV.	[184]
Scleroderma		Increased MC density in the skin of patients with scleroderma.	[185]
Rheumatoid arthritis (RA)	K/BxN mice	Higher number of mast cells at synovial cavity of RA patients.	[54,56]
		In MCs deficient animal models, mice were resistant to joint inflammation development.	[78]
Multiple sclerosis (MS)	EAE	MCs presence in demyelinated plaques. Animal models showing conflicting data. It has been demonstrated both deleterous as a regulatory effect or even none mast cells involvement.	[80,82,84,88]
Inflammatory bowel disease (IBD)	Dextran sodium sulfate (DSS) and trinitrobenzene sulfonic acid (TNBS) induced colitis	MCs mediators presence in patients.	[96,97]
		Treatment with MCs inhibitors mediators reduces disease progression.	[98,102]

and the degrees of fibrosis and lymphoid infiltration in Sjogren's syndrome [183]. MCs infiltration was also found in skin lesions from patients suffering from pemphigus vulgaris [184]. These cells were also present in increased numbers or in an activated state in the dermis of patients with scleroderma [185].

#### Alternative function of mast cells in autoimmune diseases

The presence MCs or of their products at the site of inflammation were readily considered as an indicative of MCs participation in disease onset. However, this evidence is supportive, but not definitive of a direct role of MCs in the development of autoimmunity. Instead of contributing to the initiation of autoimmunity, MCs could actually be recruited and activated in response to the inflammatory milieu, participating solely in the subsequent tissue damage associated with disease progression. In agreement to this, MCs induce human

orbital fibroblasts to produce increased levels of prostaglandin E<sub>2</sub> and hyaluronan, indicating that these cells are involved in tissue remodeling that occurs in TAO [186]. In scleroderma, it is likely that MCs play a major role in the clinical progression of skin changes, through regulation of fibroblast production of extracellular matrix [187,188].

#### Autoimmune disease study in MC-deficient mice

Kit mutant mice have been considered for decades as a powerful tool to evaluate *in vivo* MCs functions [189]. However, Kit is also involved in the development and function of many stem and mature cells inside and outside of the immune system [190]. Thereafter, some of the phenotype outcomes from Kit mutation are unrelated to MCs deficiency. Recently, it was generated a mice strain deficient in MCs (Cre-Lox system) which is independent of alterations in Kit thus

permitting direct analyses of the functions of MCs. Interestingly, some key data obtained earlier with Kit mutant mice were not confirmed with this new strain, inclusively data regarding the pathogenic role of MCs in experimental models of autoimmune arthritis and encephalomyelitis [191]. Of course, these results do not discredit all of the available MCs literature obtained with Kit mutant mice. But certainly, a re-evaluation of MCs immunological functions attributed based on the results obtained with this mice strain is necessary.

## Conclusion

Due to the widespread expression of inhibitory and activation cell-surface receptors on MCs [21], they will respond variably depending on the physiologic setting [192]. Additionally, MCs can release an impressive array of mediators, many of which can mediate proinflammatory, anti-inflammatory and/or immune regulatory functions [22]. Together, these morphological features of MCs translate to distinct effects they may exert in autoimmune diseases. Despite the substantial data from studies of both human disease and animal models that indicated a role of MCs in autoimmunity, there still is a debate regarding the actual function of MCs in those conditions [193]. Further studies are essential to gain a more detailed understanding of MC role in autoimmunity. Targeting MCs may be of value in future prevention and treatment of disease. The use of MCs-stabilizing agents would offer a possibility for the prevention of autoimmune conditions in whose development MCs may be involved (Table 1).

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## References

1. Root-Bernstein R, Fairweather D. Complexities in the relationship between infection and autoimmunity. *Curr Allergy Asthma Rep.* 2014; 14: 407.
2. Davidson A, Diamond B. Autoimmune diseases. *N Engl J Med.* 2001; 345: 340-350.
3. Wahren-Herlenius M, Dörner T. Immunopathogenic mechanisms of systemic autoimmune disease. *Lancet.* 2013; 382: 819-831.
4. Alam J, Kim YC, Choi Y. Potential role of bacterial infection in autoimmune diseases: a new aspect of molecular mimicry. *Immune Netw.* 2014; 14: 7-13.
5. Tiniakou E, Costenbader KH, Kriegel MA. Sex-specific environmental influences on the development of autoimmune diseases. *Clin Immunol.* 2013; 149: 182-191.
6. Anaya JM. Common mechanisms of autoimmune diseases (the autoimmune tautology). *Autoimmun Rev.* 2012; 11: 781-784.
7. Yin L, Dai S, Clayton G, Gao W, Wang Y, Kappler J, et al. Recognition of self and altered self by T cells in autoimmunity and allergy. *Protein Cell.* 2013; 4: 8-16.
8. Burt TD. Fetal regulatory T cells and peripheral immune tolerance in utero: implications for development and disease. *Am J Reprod Immunol.* 2013; 69: 346-358.
9. Sakaguchi S, Yamaguchi T, Nomura T, Ono M. Regulatory T cells and immune tolerance. *Cell.* 2008; 133: 775-787.
10. Singer BD, King LS, D'Alessio FR. Regulatory T Cells as Immunotherapy. *Front Immunol.* 2014; 5: 46.
11. Soyer OU, Akdis M, Ring J, Behrendt H, Cramer R, Lauener R, et al. Mechanisms of peripheral tolerance to allergens. *Allergy.* 2013; 68: 161-170.
12. Yin Y, Li Y, Mariuzza RA. Structural basis for self-recognition by autoimmune T-cell receptors. *Immunol Rev.* 2012; 250: 32-48.
13. von Boehmer H, Melchers F. Checkpoints in lymphocyte development and autoimmune disease. *Nat Immunol.* 2010; 11: 14-20.
14. Tobón GJ, Izquierdo JH, Cañas CA. B lymphocytes: development, tolerance, and their role in autoimmunity-focus on systemic lupus erythematosus. *Autoimmune Dis.* 2013; 2013: 827254.
15. Trowsdale J, Knight JC. Major histocompatibility complex genomics and human disease. *Annu Rev Genomics Hum Genet.* 2013; 14: 301-323.
16. Pollard KM, Kono DH. Requirements for innate immune pathways in environmentally induced autoimmunity. *BMC Med.* 2013; 11: 100.
17. Collington SJ, Williams TJ, Weller CL. Mechanisms underlying the localisation of mast cells in tissues. *Trends Immunol.* 2011; 32: 478-485.
18. Irani AM, Schwartz LB. Human mast cell heterogeneity. *Allergy Proc.* 1994; 15: 303-308.
19. Krishnaswamy G, Ajitawi O, Chi DS. The human mast cell: an overview. *Methods Mol Biol.* 2006; 315: 13-34.
20. Rivera J, Fierro NA, Olivera A, Suzuki R. New insights on mast cell activation via the high affinity receptor for IgE. *Adv Immunol.* 2008; 98: 85-120.
21. Gilfillan AM, Tkaczyk C. Integrated signalling pathways for mast-cell activation. *Nat Rev Immunol.* 2006; 6: 218-230.
22. Galli SJ, Nakae S, Tsai M. Mast cells in the development of adaptive immune responses. *Nat Immunol.* 2005; 6: 135-142.
23. Bischoff SC. Role of mast cells in allergic and non-allergic immune responses: comparison of human and murine data. *Nat Rev Immunol.* 2007; 7: 93-104.
24. Suto H, Nakae S, Kakurai M, Sedgwick JD, Tsai M, Galli SJ. Mast cell-associated TNF promotes dendritic cell migration. *J Immunol.* 2006; 176: 4102-4112.
25. Heib V, Becker M, Warger T, Rechtsteiner G, Tertilt C, Klein M, et al. Mast cells are crucial for early inflammation, migration of Langerhans cells, and CTL responses following topical application of TLR7 ligand in mice. *Blood.* 2007; 110: 946-953.
26. Dawicki W, Jawdat DW, Xu N, Marshall JS. Mast cells, histamine, and IL-6 regulate the selective influx of dendritic cell subsets into an inflamed lymph node. *J Immunol.* 2010; 184: 2116-2123.
27. Mazzoni A, Young HA, Spitzer JH, Visintin A, Segal DM. Histamine regulates cytokine production in maturing dendritic cells, resulting in altered T cell polarization. *J Clin Invest.* 2001; 108: 1865-1873.
28. Idzko M, la Sala A, Ferrari D, Panther E, Herouy Y, Dichmann S, et al. Expression and function of histamine receptors in human monocyte-derived dendritic cells. *J Allergy Clin Immunol.* 2002; 109: 839-846.
29. Gutzmer R, Langer K, Lisewski M, Mommert S, Rieckborn D, Kapp A, et al. Expression and function of histamine receptors 1 and 2 on human monocyte-derived dendritic cells. *J Allergy Clin Immunol.* 2002; 109: 524-531.
30. Skokos D, Goubran-Botros H, Roa M, Mécheri S. Immunoregulatory properties of mast cell-derived exosomes. *Mol Immunol.* 2002; 38: 1359-1362.
31. Skokos D, Botros HG, Demeure C, Morin J, Peronet R, Birkenmeier G, et al. Mast cell-derived exosomes induce phenotypic and functional maturation of dendritic cells and elicit specific immune responses in vivo. *J Immunol.* 2003; 170: 3037-3045.
32. Dudeck A, Suender CA, Kostka SL, von Stebut E, Maurer M. Mast cells promote Th1 and Th17 responses by modulating dendritic cell maturation and function. *Eur J Immunol.* 2011; 41: 1883-1893.
33. Mazzoni A, Siraganian RP, Leifer CA, Segal DM. Dendritic cell modulation by mast cells controls the Th1/Th2 balance in responding T cells. *J Immunol.* 2006; 177: 3577-3581.

34. Theiner G, Gessner A, Lutz MB. The mast cell mediator PGD2 suppresses IL-12 release by dendritic cells leading to Th2 polarized immune responses in vivo. *Immunobiology*. 2006; 211: 463-472.
35. Askenase PW. Mast cells and the mediation of T-cell recruitment in arthritis. *N Engl J Med*. 2003; 349: 1294.
36. Orinska Z, Bulanova E, Budagian V, Metz M, Maurer M, Bulfone-Paus S. TLR3-induced activation of mast cells modulates CD8+ T-cell recruitment. *Blood*. 2005; 106: 978-987.
37. Kunder CA, St John AL, Abraham SN. Mast cell modulation of the vascular and lymphatic endothelium. *Blood*. 2011; 118: 5383-5393.
38. Poncet P, Arock M, David B. MHC class II-dependent activation of CD4+ T cell hybridomas by human mast cells through superantigen presentation. *J Leukoc Biol*. 1999; 66: 105-112.
39. Malaviya R, Twesten NJ, Ross EA, Abraham SN, Pfeifer JD. Mast cells process bacterial Ags through a phagocytic route for class I MHC presentation to T cells. *J Immunol*. 1996; 156: 1490-1496.
40. Gaudenzio N, Espagnolle N, Mars LT, Liblaur R, Valitutti S, Espinosa E. Cell-cell cooperation at the T helper cell/mast cell immunological synapse. *Blood*. 2009; 114: 4979-4988.
41. Stelekati E, Bahri R, D'Orlando O, Orinska Z, Mittrücker HW, Langenhau R, et al. Mast cell-mediated antigen presentation regulates CD8+ T cell effector functions. *Immunity*. 2009; 31: 665-676.
42. Nakae S, Suto H, Iikura M, Kakurai M, Sedgwick JD, Tsai M, et al. Mast cells enhance T cell activation: importance of mast cell costimulatory molecules and secreted TNF. *J Immunol*. 2006; 176: 2238-2248.
43. Kashiwakura J, Yokoi H, Saito H, Okayama Y. T cell proliferation by direct cross-talk between OX40 ligand on human mast cells and OX40 on human T cells: comparison of gene expression profiles between human tonsillar and lung-cultured mast cells. *J Immunol*. 2004; 173: 5247-5257.
44. Sayed BA, Christy A, Quirion MR, Brown MA. The master switch: the role of mast cells in autoimmunity and tolerance. *Annu Rev Immunol*. 2008; 26: 705-739.
45. Jutel M, Watanabe T, Klunker S, Akdis M, Thomet OA, Malolepszy J, et al. Histamine regulates T-cell and antibody responses by differential expression of H1 and H2 receptors. *Nature*. 2001; 413: 420-425.
46. Skokos D, Le Panse S, Villa I, Rousselle JC, Peronet R, David B, et al. Mast cell-dependent B and T lymphocyte activation is mediated by the secretion of immunologically active exosomes. *J Immunol*. 2001; 166: 868-876.
47. Yoshikawa T, Imada T, Nakakubo H, Nakamura N, Naito K. Rat mast cell protease-1 enhances immunoglobulin E production by mouse B cells stimulated with interleukin-4. *Immunology*. 2001; 104: 333-340.
48. Merluzzi S, Frossi B, Gri G, Parusso S, Tripodo C, Pucillo C. Mast cells enhance proliferation of B lymphocytes and drive their differentiation toward IgA-secreting plasma cells. *Blood*. 2010; 115: 2810-2817.
49. McInnes IB, Schett G. The pathogenesis of rheumatoid arthritis. *N Engl J Med*. 2011; 365: 2205-2219.
50. Boissier MC, Semerano L, Challal S, Saldenber-Kermanac'h N, Falgarone G. Rheumatoid arthritis: from autoimmunity to synovitis and joint destruction. *J Autoimmun*. 2012; 39: 222-228.
51. Davignon JL, Hayder M, Baron M, Boyer JF, Constantin A, Apparailly F, et al. Targeting monocytes/macrophages in the treatment of rheumatoid arthritis. *Rheumatology (Oxford)*. 2013; 52: 590-598.
52. Cornish AL, Campbell IK, McKenzie BS, Chatfield S, Wicks IP. G-CSF and GM-CSF as therapeutic targets in rheumatoid arthritis. *Nat Rev Rheumatol*. 2009; 5: 554-559.
53. Crisp AJ, Chapman CM, Kirkham SE, Schiller AL, Krane SM. Articular mastocytosis in rheumatoid arthritis. *Arthritis Rheum*. 1984; 27: 845-851.
54. Malone DG, Wilder RL, Saavedra-Delgado AM, Metcalfe DD. Mast cell numbers in rheumatoid synovial tissues. Correlations with quantitative measures of lymphocytic infiltration and modulation by anti-inflammatory therapy. *Arthritis Rheum*. 1987; 30: 130-137.
55. Eklund KK. Mast cells in the pathogenesis of rheumatic diseases and as potential targets for anti-rheumatic therapy. *Immunol Rev*. 2007; 217: 38-52.
56. Lee DM, Friend DS, Gurish MF, Benoist C, Mathis D, Brenner MB. Mast cells: a cellular link between autoantibodies and inflammatory arthritis. *Science*. 2002; 297: 1689-1692.
57. Rajasekaran N, Solomon S, Watanabe T, Ohtsu H, Gajda M, Bräuer R, et al. Histidine decarboxylase but not histamine receptor 1 or 2 deficiency protects from K/BxN serum-induced arthritis. *Int Immunol*. 2009; 21: 1263-1268.
58. Nent E, Frommholz D, Gajda M, Bräuer R, Illges H. Histamine 4 receptor plays an important role in auto-antibody-induced arthritis. *Int Immunol*. 2013; 25: 437-443.
59. Abd-Allah AR, Ahmad SF2, Alrashidi I2, Abdel-Hamied HE3, Zoheir KM4, Ashour AE2, Bakheet SA2. Involvement of histamine 4 receptor in the pathogenesis and progression of rheumatoid arthritis. *Int Immunol*. 2014; 26: 325-340.
60. Sawamukai N, Yukawa S, Saito K, Nakayama S, Kambayashi T, Tanaka Y. Mast cell-derived tryptase inhibits apoptosis of human rheumatoid synovial fibroblasts via rho-mediated signaling. *Arthritis Rheum*. 2010; 62: 952-959.
61. Palmer HS, Kelso EB, Lockhart JC, Sommerhoff CP, Plevin R, Goh FG, et al. Protease-activated receptor 2 mediates the proinflammatory effects of synovial mast cells. *Arthritis Rheum*. 2007; 56: 3532-3540.
62. Magarinos NJ, Bryant KJ, Fosang AJ, Adachi R, Stevens RL, McNeil HP. Mast cell-restricted, tetramer-forming tryptases induce aggrecanolytic activity in articular cartilage by activating matrix metalloproteinase-3 and -13 zymogens. *J Immunol*. 2013; 191: 1404-1412.
63. Markatseli TE, Papagoras C, Nikoli A, Voulgari PV, Drosos AA. Certolizumab for rheumatoid arthritis. *Clin Exp Rheumatol*. 2014; 32: 415-423.
64. Kashiwakura J, Yanagisawa M, Lee H, Okamura Y, Sasaki-Sakamoto T, Saito S, et al. Interleukin-33 synergistically enhances immune complex-induced tumor necrosis factor alpha and interleukin-8 production in cultured human synovium-derived mast cells. *Int Arch Allergy Immunol*. 2013; 161: 32-36.
65. Sandler C, Lindstedt KA, Joutsiniemi S, Lappalainen J, Juutilainen T, Kolah J, et al. Selective activation of mast cells in rheumatoid synovial tissue results in production of TNF-alpha, IL-1beta and IL-1Ra. *Inflamm Res*. 2007; 56: 230-239.
66. Xu D, Jiang HR, Li Y, Pushparaj PN, Kurowska-Stolarska M, Leung BP, et al. IL-33 exacerbates autoantibody-induced arthritis. *J Immunol*. 2010; 184: 2620-2626.
67. Xu D, Jiang HR, Kewin P, Li Y, Mu R, Fraser AR, Pitman N. IL-33 exacerbates antigen-induced arthritis by activating mast cells. *Proc Natl Acad Sci U S A*. 2008; 105: 10913-10918.
68. Suurmond J, Dorjée AL, Boon MR, Knol EF, Huizinga TW, Toes RE, et al. Mast cells are the main interleukin 17-positive cells in anticitrullinated protein antibody-positive and -negative rheumatoid arthritis and osteoarthritis synovium. *Arthritis Res Ther*. 2011; 13: R150.
69. Appel H, Maier R, Wu P, Scheer R, Hempfing A, Kayser R, et al. Analysis of IL-17(+) cells in facet joints of patients with spondyloarthritis suggests that the innate immune pathway might be of greater relevance than the Th17-mediated adaptive immune response. *Arthritis Res Ther*. 2011; 13: R95.
70. Hueber AJ, Asquith DL, Miller AM, Reilly J, Kerr S, Leipe J, et al. Mast cells express IL-17A in rheumatoid arthritis synovium. *J Immunol*. 2010; 184: 3336-3340.
71. Noordenbos T, Yerenenko N, Gofita I, van de Sande M, Tak PP, Cañete JD, et al. Interleukin-17-positive mast cells contribute to synovial inflammation in spondylarthritis. *Arthritis Rheum*. 2012; 64: 99-109.
72. Lee H, Kashiwakura J, Matsuda A, Watanabe Y, Sakamoto-Sasaki T, Matsumoto K, et al. Activation of human synovial mast cells from rheumatoid arthritis or osteoarthritis patients in response to aggregated IgG through Fcγ3

- receptor I and Fc $\epsilon$ 1 receptor II. *Arthritis Rheum.* 2013; 65: 109-119.
73. Malbec O, Daéron M. The mast cell IgG receptors and their roles in tissue inflammation. *Immunol Rev.* 2007; 217: 206-221.
  74. Lassmann H, van Horssen J, Mahad D. Progressive multiple sclerosis: pathology and pathogenesis. *Nat Rev Neurol.* 2012; 8: 647-656.
  75. Hauser SL, Chan JR, Oksenberg JR. Multiple sclerosis: Prospects and promise. *Ann Neurol.* 2013; 74: 317-327.
  76. Silver R, Curley JP. Mast cells on the mind: new insights and opportunities. *Trends Neurosci.* 2013; 36: 513-521.
  77. Olsson Y. Mast cells in plaques of multiple sclerosis. *Acta Neurol Scand.* 1974; 50: 611-618.
  78. Rozniecki JJ, Hauser SL, Stein M, Lincoln R, Theoharides TC. Elevated mast cell tryptase in cerebrospinal fluid of multiple sclerosis patients. *Ann Neurol.* 1995; 37: 63-66.
  79. Secor VH, Secor WE, Gutekunst CA, Brown MA. Mast cells are essential for early onset and severe disease in a murine model of multiple sclerosis. *J Exp Med.* 2000; 191: 813-822.
  80. Gregory GD, Robbie-Ryan M, Secor VH, Sabatino JJ Jr, Brown MA. Mast cells are required for optimal autoreactive T cell responses in a murine model of multiple sclerosis. *Eur J Immunol.* 2005; 35: 3478-3486.
  81. Gregory GD, Raju SS, Winandy S, Brown MA. Mast cell IL-4 expression is regulated by Ikaros and influences encephalitogenic Th1 responses in EAE. *J Clin Invest.* 2006; 116: 1327-1336.
  82. Christy AL, Walker ME, Hessner MJ, Brown MA. Mast cell activation and neutrophil recruitment promotes early and robust inflammation in the meninges in EAE. *J Autoimmun.* 2013; 42: 50-61.
  83. Piconese S, Costanza M, Musio S, Tripodo C, Poliani PL, Gri G, et al. Exacerbated experimental autoimmune encephalomyelitis in mast-cell-deficient Kit W-sh/W-sh mice. *Lab Invest.* 2011; 91: 627-641.
  84. Li H, Nourbakhsh B, Safavi F, Li K, Xu H, Cullimore M, et al. Kit (W-sh) mice develop earlier and more severe experimental autoimmune encephalomyelitis due to absence of immune suppression. *J Immunol.* 2011; 187: 274-282.
  85. Hendrix S, Kramer P, Pehl D, Warnke K, Boato F, Nelissen S, et al. Mast cells protect from post-traumatic brain inflammation by the mast cell-specific chymase mouse mast cell protease-4. *FASEB J.* 2013; 27: 920-929.
  86. Nelissen S, Vanganswinkel T, Geurts N, Geboes L, Lemmens E, Vidal PM, et al. Mast cells protect from post-traumatic spinal cord damage in mice by degrading inflammation-associated cytokines via mouse mast cell protease 4. *Neurobiol Dis.* 2014; 62: 260-272.
  87. Bennett JL, Blanchet MR, Zhao L, Zbytniuk L, Antignano F, Gold M, et al. Bone marrow-derived mast cells accumulate in the central nervous system during inflammation but are dispensable for experimental autoimmune encephalomyelitis pathogenesis. *J Immunol.* 2009; 182: 5507-5514.
  88. Feyerabend TB, Weiser A, Tietz A, Stassen M, Harris N, Kopf M, et al. Cre-mediated cell ablation contests mast cell contribution in models of antibody- and T cell-mediated autoimmunity. *Immunity.* 2011; 35: 832-844.
  89. Zhang YZ, Li YY. Inflammatory bowel disease: pathogenesis. *World J Gastroenterol.* 2014; 20: 91-99.
  90. Cader MZ, Kaser A. Recent advances in inflammatory bowel disease: mucosal immune cells in intestinal inflammation. *Gut.* 2013; 62: 1653-1664.
  91. Reis BS, Mucida D. The role of the intestinal context in the generation of tolerance and inflammation. *Clin Dev Immunol.* 2012; 2012: 157948.
  92. Maloy KJ, Powrie F. Intestinal homeostasis and its breakdown in inflammatory bowel disease. *Nature.* 2011; 474: 298-306.
  93. Geremia A, Biancheri P, Allan P, Corazza GR, Di Sabatino A. Innate and adaptive immunity in inflammatory bowel disease. *Autoimmun Rev.* 2014; 13: 3-10.
  94. Kurashima Y, Goto Y, Kiyono H. Mucosal innate immune cells regulate both gut homeostasis and intestinal inflammation. *Eur J Immunol.* 2013; 43: 3108-3115.
  95. Dahlin JS, Hallgren J2. Mast cell progenitors: Origin, development and migration to tissues. *Mol Immunol.* 2014;.
  96. Stasikowska-Kanicka O, Danilewicz M, GÅowacka A, WÅ...gowska-Danilewicz M. Mast cells and eosinophils are involved in activation of ulcerative colitis. *Adv Med Sci.* 2012; 57: 230-236.
  97. He SH, Xie H, Fu YL. Inhibition of tryptase release from human colon mast cells by histamine receptor antagonists. *Asian Pac J Allergy Immunol.* 2005; 23: 35-39.
  98. Hamilton MJ, Sinnamon MJ, Lyng GD, Glickman JN, Wang X, Xing W, Kriliis SA. Essential role for mast cell tryptase in acute experimental colitis. *Proc Natl Acad Sci U S A.* 2011; 108: 290-295.
  99. Isozaki Y, Yoshida N, Kuroda M, Handa O, Takagi T, Kokura S, Ichikawa H. Anti-tryptase treatment using nafamostat mesilate has a therapeutic effect on experimental colitis. *Scand J Gastroenterol.* 2006; 41: 944-953.
  100. Rijnierse A, Koster AS, Nijkamp FP, Kraneveld AD. TNF-alpha is crucial for the development of mast cell-dependent colitis in mice. *Am J Physiol Gastrointest Liver Physiol.* 2006; 291: G969-976.
  101. Lohman RJ, Cotterell AJ, Suen J, Liu L, Do AT, Vesey DA, et al. Antagonism of protease-activated receptor 2 protects against experimental colitis. *J Pharmacol Exp Ther.* 2012; 340: 256-265.
  102. Kurashima Y, Amiya T, Nochi T, Fujisawa K, Haraguchi T, Iba H, et al. Extracellular ATP mediates mast cell-dependent intestinal inflammation through P2X7 purinoceptors. *Nat Commun.* 2012; 3: 1034.
  103. van Belle TL, Coppieters KT, von Herrath MG. Type 1 diabetes: etiology, immunology, and therapeutic strategies. *Physiol Rev.* 2011; 91: 79-118.
  104. Wallis RH, Wang K, Marandi L, Hsieh E, Ning T, Chao GY, Sarmiento J. Type 1 diabetes in the BB rat: a polygenic disease. *Diabetes.* 2009; 58: 1007-1017.
  105. Matsumoto M, Yagi H, Kunimoto K, Kawaguchi J, Makino S, Harada M. Transfer of autoimmune diabetes from diabetic NOD mice to NOD athymic nude mice: the roles of T cell subsets in the pathogenesis. *Cell Immunol.* 1993; 148: 189-197.
  106. Yagi H, Matsumoto M, Kunimoto K, Kawaguchi J, Makino S, Harada M. Analysis of the roles of CD4+ and CD8+ T cells in autoimmune diabetes of NOD mice using transfer to NOD athymic nude mice. *Eur J Immunol.* 1992; 22: 2387-2393.
  107. Hu CY, Rodriguez-Pinto D, Du W, Ahuja A, Henegariu O, Wong FS, et al. Treatment with CD20-specific antibody prevents and reverses autoimmune diabetes in mice. *J Clin Invest.* 2007; 117: 3857-3867.
  108. Serreze DV, Chapman HD, Varnum DS, Hanson MS, Reifsnyder PC, Richard SD, et al. B lymphocytes are essential for the initiation of T cell-mediated autoimmune diabetes: analysis of a new "speed congenic" stock of NOD.Ig mu null mice. *J Exp Med.* 1996; 184: 2049-2053.
  109. Saxena V, Ondr JK, Magnusen AF, Munn DH, Katz JD. The countervailing actions of myeloid and plasmacytoid dendritic cells control autoimmune diabetes in the nonobese diabetic mouse. *J Immunol.* 2007; 179: 5041-5053.
  110. Hutchings P, Rosen H, O'Reilly L, Simpson E, Gordon S, Cooke A. Transfer of diabetes in mice prevented by blockade of adhesion-promoting receptor on macrophages. *Nature.* 1990; 348: 639-642.
  111. Zimnoch L, Szynaka B, Puchalski Z. Mast cells and pancreatic stellate cells in chronic pancreatitis with differently intensified fibrosis. *Hepatogastroenterology.* 2002; 49: 1135-1138.
  112. Dib M, Zhao X, Wang XD, Andersson R. Role of mast cells in the development of pancreatitis-induced multiple organ dysfunction. *Br J Surg.* 2002; 89: 172-178.
  113. Dib M, Zhao X, Wang X, Andersson R. Mast cells contribute to early pancreatitis-induced systemic endothelial barrier dysfunction. *Pancreatol.* 2002; 2: 396-401.

114. Strouch MJ, Cheon EC, Salabat MR, Krantz SB, Gounaris E, Melstrom LG, Dangi-Garimella S. Crosstalk between mast cells and pancreatic cancer cells contributes to pancreatic tumor progression. *Clin Cancer Res*. 2010; 16: 2257-2265.
115. Hessner MJ, Wang X, Meyer L, Geoffrey R, Jia S, Fuller J, et al. Involvement of eotaxin, eosinophils, and pancreatic predisposition in development of type 1 diabetes mellitus in the BioBreeding rat. *J Immunol*. 2004; 173: 6993-7002.
116. Geoffrey R, Jia S, Kwitek AE, Woodliff J, Ghosh S, Lernmark A, et al. Evidence of a functional role for mast cells in the development of type 1 diabetes mellitus in the BioBreeding rat. *J Immunol*. 2006; 177: 7275-7286.
117. Yang XD, Tisch R, Singer SM, Cao ZA, Liblau RS, Schreiber RD, et al. Effect of tumor necrosis factor alpha on insulin-dependent diabetes mellitus in NOD mice. I. The early development of autoimmunity and the diabetogenic process. *J Exp Med*. 1994; 180: 995-1004.
118. Cetkovic-Cvrlje M, Eizirik DL. TNF-alpha and IFN-gamma potentiate the deleterious effects of IL-1 beta on mouse pancreatic islets mainly via generation of nitric oxide. *Cytokine*. 1994; 6: 399-406.
119. Green EA, Eynon EE, Flavell RA. Local expression of TNFalpha in neonatal NOD mice promotes diabetes by enhancing presentation of islet antigens. *Immunity*. 1998; 9: 733-743.
120. Green EA, Flavell RA. Tumor necrosis factor-alpha and the progression of diabetes in non-obese diabetic mice. *Immunol Rev*. 1999; 169: 11-22.
121. Suk K, Kim S, Kim YH, Kim KA, Chang I, Yagita H, et al. IFN-gamma/TNF-alpha synergism as the final effector in autoimmune diabetes: a key role for STAT1/IFN regulatory factor-1 pathway in pancreatic beta cell death. *J Immunol*. 2001; 166: 4481-4489.
122. Wang B, André I, Gonzalez A, Katz JD, Aguet M, Benoist C, et al. Interferon-gamma impacts at multiple points during the progression of autoimmune diabetes. *Proc Natl Acad Sci U S A*. 1997; 94: 13844-13849.
123. Gupta AA, Leal-Berumen I, Croitoru K, Marshall JS. Rat peritoneal mast cells produce IFN-gamma following IL-12 treatment but not in response to IgE-mediated activation. *J Immunol*. 1996; 157: 2123-2128.
124. Ankarcrona M, Dypbukt JM, Brüne B, Nicotera P. Interleukin-1 beta-induced nitric oxide production activates apoptosis in pancreatic RINm5F cells. *Exp Cell Res*. 1994; 213: 172-177.
125. Wang Q, Zhang H, Zhao B, Fei H. IL-1beta caused pancreatic beta-cells apoptosis is mediated in part by endoplasmic reticulum stress via the induction of endoplasmic reticulum Ca<sup>2+</sup> release through the c-Jun N-terminal kinase pathway. *Mol Cell Biochem*. 2009; 324: 183-190.
126. Chen MC, Schuit F, Eizirik DL. Identification of IL-1beta-induced messenger RNAs in rat pancreatic beta cells by differential display of messenger RNA. *Diabetologia*. 1999; 42: 1199-1203.
127. Thomas HE, Irawaty W, Darwiche R, Brodnicki TC, Santamaria P, Allison J, et al. IL-1 receptor deficiency slows progression to diabetes in the NOD mouse. *Diabetes*. 2004; 53: 113-121.
128. Nakamura Y, Franchi L, Kambe N, Meng G, Strober W, Núñez G. Critical role for mast cells in interleukin-1 $\beta$ -driven skin inflammation associated with an activating mutation in the nlrp3 protein. *Immunity*. 2012; 37: 85-95.
129. Mizutani H, Schechter N, Lazarus G, Black RA, Kupper TS. Rapid and specific conversion of precursor interleukin 1 beta (IL-1 beta) to an active IL-1 species by human mast cell chymase. *J Exp Med*. 1991; 174: 821-825.
130. Gibbs BF, Haas H, Wolff HH, Grabbe J. Early IgE-dependent release of IL-4 and IL-13 from leukocytes is restricted to basophils: a comparison with other granulocytes and mononuclear cells. *Inflamm Res*. 2000; 49 Suppl 1: S9-10.
131. Mok CC, Lau CS. Pathogenesis of systemic lupus erythematosus. *J Clin Pathol*. 2003; 56: 481-490.
132. Crispín JC, Lioussis SN, Kis-Toth K, Lieberman LA, Kyttaris VC, Juang YT, Tsokos GC. Pathogenesis of human systemic lupus erythematosus: recent advances. *Trends Mol Med*. 2010; 16: 47-57.
133. Choi J, Kim ST, Craft J. The pathogenesis of systemic lupus erythematosus—an update. *Curr Opin Immunol*. 2012; 24: 651-657.
134. Liu Z, Davidson A. Taming lupus—a new understanding of pathogenesis leading to clinical advances. *Nat Med*. 2012; 18: 871-882.
135. Borchers AT, Leibushor N, Naguwa SM, Cheema GS, Shoenfeld Y, Gershwin ME. Lupus nephritis: a critical review. *Autoimmun Rev*. 2012; 12: 174-194.
136. Lech M, Anders HJ. The pathogenesis of lupus nephritis. *J Am Soc Nephrol*. 2013; 24: 1357-1366.
137. Danilewicz M, Wagrowska-Danilewicz M. Quantitative analysis of interstitial mast cells in lupus and non-lupus membranous glomerulopathy. *Pol J Pathol*. 2001; 52: 211-217.
138. Hiromura K, Kurosawa M, Yano S, Naruse T. Tubulointerstitial mast cell infiltration in glomerulonephritis. *Am J Kidney Dis*. 1998; 32: 593-599.
139. Lin L, Gerth AJ, Peng SL. Susceptibility of mast cell-deficient W/W<sup>v</sup> mice to pristane-induced experimental lupus nephritis. *Immunol Lett*. 2004; 91: 93-97.
140. Griffiths CE, Barker JN. Pathogenesis and clinical features of psoriasis. *Lancet*. 2007; 370: 263-271.
141. Krueger JG, Bowcock A. Psoriasis pathophysiology: current concepts of pathogenesis. *Ann Rheum Dis*. 2005; 64 Suppl 2: ii30-36.
142. Valdimarsson H, Thorleifsdottir RH, Sigurdardottir SL, Gudjonsson JE, Johnston A. Psoriasis—as an autoimmune disease caused by molecular mimicry. *Trends Immunol*. 2009; 30: 494-501.
143. Lowes MA, Bowcock AM, Krueger JG. Pathogenesis and therapy of psoriasis. *Nature*. 2007; 445: 866-873.
144. Harvima IT, Naukkarinen A, Harvima RJ, Aalto ML, Neittaanmäki H, Horsmanheimo M. Quantitative enzyme-histochemical analysis of tryptase- and chymase-containing mast cells in psoriatic skin. *Arch Dermatol Res*. 1990; 282: 428-433.
145. Iversen OJ, Lysvand H, Jacobsen T, Bergh K, Lie BA. The psoriasis-associated antigen, pso p27, is expressed by tryptase-positive cells in psoriatic lesions. *Arch Dermatol Res*. 1995; 287: 503-505.
146. Harvima IT, Naukkarinen A, Paukkonen K, Harvima RJ, Aalto ML, Schwartz LB, Horsmanheimo M. Mast cell tryptase and chymase in developing and mature psoriatic lesions. *Arch Dermatol Res*. 1993; 285: 184-192.
147. Krogstad AL, Lönnroth P, Larson G, Wallin BG. Increased interstitial histamine concentration in the psoriatic plaque. *J Invest Dermatol*. 1997; 109: 632-635.
148. Yamamoto T, Katayama I, Nishioka K. Possible contribution of stem cell factor in psoriasis vulgaris. *J Dermatol Sci*. 2000; 24: 171-176.
149. Ackermann L, Harvima IT, Pelkonen J, Ritamäki-Salo V, Naukkarinen A, Harvima RJ, Horsmanheimo M. Mast cells in psoriatic skin are strongly positive for interferon-gamma. *Br J Dermatol*. 1999; 140: 624-633.
150. Yao Y, Richman L, Morehouse C, de los Reyes M, Higgs BW, Boutrin A, et al. Type I interferon: potential therapeutic target for psoriasis? *PLoS One*. 2008; 3: e2737.
151. Lin AM, Rubin CJ, Khandpur R, Wang JY, Riblett M, Yalavarthi S, et al. Mast cells and neutrophils release IL-17 through extracellular trap formation in psoriasis. *J Immunol*. 2011; 187: 490-500.
152. Liang SC, Tan XY, Luxenberg DP, Karim R, Dunussi-Joannopoulos K, Collins M, et al. Interleukin (IL)-22 and IL-17 are coexpressed by Th17 cells and cooperatively enhance expression of antimicrobial peptides. *J Exp Med*. 2006; 203: 2271-2279.
153. Nograles KE, Zaba LC, Guttman-Yassky E, Fuentes-Duculan J, Suárez-Fariñas M, Cardinale I, Khatcherian A. Th17 cytokines interleukin (IL)-17 and IL-22 modulate distinct inflammatory and keratinocyte-response pathways. *Br J Dermatol*. 2008; 159: 1092-1102.
154. Chiricozzi A, Guttman-Yassky E, Suárez-Fariñas M, Nograles KE, Tian S, Cardinale I, Chimenti S. Integrative responses to IL-17 and TNF- $\alpha$  in human keratinocytes account for key inflammatory pathogenic circuits in psoriasis. *J Invest Dermatol*. 2011; 131: 677-687.

155. Ariza ME, Williams MV, Wong HK. Targeting IL-17 in psoriasis: from cutaneous immunobiology to clinical application. *Clin Immunol*. 2013; 146: 131-139.
156. Steinhoff M, Corvera CU, Thoma MS, Kong W, McAlpine BE, Caughey GH, et al. Proteinase-activated receptor-2 in human skin: tissue distribution and activation of keratinocytes by mast cell tryptase. *Exp Dermatol*. 1999; 8: 282-294.
157. Kasperkiewicz M, Zillikens D. The pathophysiology of bullous pemphigoid. *Clin Rev Allergy Immunol*. 2007; 33: 67-77.
158. Nishie W. Update on the pathogenesis of bullous pemphigoid: an autoantibody-mediated blistering disease targeting collagen XVII. *J Dermatol Sci*. 2014; 73: 179-186.
159. Liu Z, Giudice GJ, Swartz SJ, Fairley JA, Till GO, Troy JL, et al. The role of complement in experimental bullous pemphigoid. *J Clin Invest*. 1995; 95: 1539-1544.
160. Thoma-Uszynski S, Uter W, Schwietzke S, Schuler G, Borradori L, Hertl M. Autoreactive T and B cells from bullous pemphigoid (BP) patients recognize epitopes clustered in distinct regions of BP180 and BP230. *J Immunol*. 2006; 176: 2015-2023.
161. Chen R, Fairley JA, Zhao ML, Giudice GJ, Zillikens D, Diaz LA, et al. Macrophages, but not T and B lymphocytes, are critical for subepidermal blister formation in experimental bullous pemphigoid: macrophage-mediated neutrophil infiltration depends on mast cell activation. *J Immunol*. 2002; 169: 3987-3992.
162. Chen R, Ning G, Zhao ML, Fleming MG, Diaz LA, Werb Z, et al. Mast cells play a key role in neutrophil recruitment in experimental bullous pemphigoid. *J Clin Invest*. 2001; 108: 1151-1158.
163. Liu Z, Giudice GJ, Zhou X, Swartz SJ, Troy JL, Fairley JA, et al. A major role for neutrophils in experimental bullous pemphigoid. *J Clin Invest*. 1997; 100: 1256-1263.
164. Wintroub BU, Mihm MC Jr, Goetzl EJ, Soter NA, Austen KF. Morphologic and functional evidence for release of mast-cell products in bullous pemphigoid. *N Engl J Med*. 1978; 298: 417-421.
165. Katayama I, Doi T, Nishioka K. High histamine level in the blister fluid of bullous pemphigoid. *Arch Dermatol Res*. 1984; 276: 126-127.
166. Fairley JA, Burnett CT, Fu CL, Larson DL, Fleming MG, Giudice GJ. A pathogenic role for IgE in autoimmunity: bullous pemphigoid IgE reproduces the early phase of lesion development in human skin grafted to nu/nu mice. *J Invest Dermatol*. 2007; 127: 2605-2611.
167. Messingham KN, Pietras TA, Fairley JA. Role of IgE in bullous pemphigoid: a review and rationale for IgE directed therapies. *G Ital Dermatol Venereol*. 2012; 147: 251-257.
168. Biedermann T, Kneilling M, Mailhammer R, Maier K, Sander CA, Kollias G, et al. Mast cells control neutrophil recruitment during T cell-mediated delayed-type hypersensitivity reactions through tumor necrosis factor and macrophage inflammatory protein 2. *J Exp Med*. 2000; 192: 1441-1452.
169. De Filippo K, Dudeck A, Hasenberg M, Nye E, van Rooijen N, Hartmann K, et al. Mast cell and macrophage chemokines CXCL1/CXCL2 control the early stage of neutrophil recruitment during tissue inflammation. *Blood*. 2013; 121: 4930-4937.
170. Liu Z, Shipley JM, Vu TH, Zhou X, Diaz LA, Werb Z, et al. Gelatinase B-deficient mice are resistant to experimental bullous pemphigoid. *J Exp Med*. 1998; 188: 475-482.
171. Lin L, Bankaitis E, Heimbach L, Li N, Abrink M, Pejler G, et al. Dual targets for mouse mast cell protease-4 in mediating tissue damage in experimental bullous pemphigoid. *J Biol Chem*. 2011; 286: 37358-37367.
172. Warrington KJ, Matteson EL. A primer on vasculitis. *Minn Med*. 2013; 96: 36-39.
173. Waller R, Ahmed A, Patel I, Luqmani R. Update on the classification of vasculitis. *Best Pract Res Clin Rheumatol*. 2013; 27: 3-17.
174. Watanabe N, Akikusa B, Park SY, Ohno H, Fossati L, Vecchietti G, et al. Mast cells induce autoantibody-mediated vasculitis syndrome through tumor necrosis factor production upon triggering Fcγ receptors. *Blood*. 1999; 94: 3855-3863.
175. Kiely PD, Pecht I, Oliveira DB. Mercuric chloride-induced vasculitis in the Brown Norway rat: alpha beta T cell-dependent and -independent phases: role of the mast cell. *J Immunol*. 1997; 159: 5100-5106.
176. Vinen CS, Turner DR, Oliveira DB. A central role for the mast cell in early phase vasculitis in the Brown Norway rat model of vasculitis: a histological study. *Int J Exp Pathol*. 2004; 85: 165-174.
177. Harris FE, Turner DR, Oliveira DB. Early vasculitis in the mercuric chloride induced Brown Norway rat model is neutrophil independent. *Int J Exp Pathol*. 1999; 80: 133-142.
178. Qasim FJ, Thiru S, Mathieson PW, Oliveira DB. The time course and characterization of mercuric chloride-induced immunopathology in the brown Norway rat. *J Autoimmun*. 1995; 8: 193-208.
179. Gan PY, Summers SA, Ooi JD, O'Sullivan KM, Tan DS, Muljadi RC, et al. Mast cells contribute to peripheral tolerance and attenuate autoimmune vasculitis. *J Am Soc Nephrol*. 2012; 23: 1955-1966.
180. Ludgate M, Baker G. Unlocking the immunological mechanisms of orbital inflammation in thyroid eye disease. *Clin Exp Immunol*. 2002; 127: 193-198.
181. Hufnagel TJ, Hickey WF, Cobbs WH, Jakobiec FA, Iwamoto T, et al. Immunohistochemical and ultrastructural studies on the exenterated orbital tissues of a patient with Graves' disease. *Ophthalmology*. 1984; 91: 1411-1419.
182. Costagliola S, Many MC, Deneff JF, Pohlenz J, Refetoff S, Vassart G. Genetic immunization of outbred mice with thyrotropin receptor cDNA provides a model of Graves' disease. *J Clin Invest*. 2000; 105: 803-811.
183. Skopouli FN, Li L, Boumba D, Stefanaki S, Hanel K, Moutsopoulos HM, et al. Association of mast cells with fibrosis and fatty infiltration in the minor salivary glands of patients with Sjögren's syndrome. *Clin Exp Rheumatol*. 1998; 16: 63-65.
184. Levi-Schaffer F, Klapholz L, Kupietzky A, Weinrauch L, Shalit M, Okon E. Increased numbers of mast cells in pemphigus vulgaris skin lesions. A histochemical study. *Acta Derm Venereol*. 1991; 71: 269-271.
185. Akimoto S, Ishikawa O, Igarashi Y, Kurosawa M, Miyachi Y. Dermal mast cells in scleroderma: their skin density, tryptase/chymase phenotypes and degranulation. *Br J Dermatol*. 1998; 138: 399-406.
186. Smith TJ, Parikh SJ. HMC-1 mast cells activate human orbital fibroblasts in coculture: evidence for up-regulation of prostaglandin E2 and hyaluronan synthesis. *Endocrinology*. 1999; 140: 3518-3525.
187. Shiota N, Kakizoe E, Shimoura K, Tanaka T, Okunishi H. Effect of mast cell chymase inhibitor on the development of scleroderma in tight-skin mice. *Br J Pharmacol*. 2005; 145: 424-431.
188. Garbuzenko E, Nagler A, Pickholtz D, Gillery P, Reich R, Maquart FX, et al. Human mast cells stimulate fibroblast proliferation, collagen synthesis and lattice contraction: a direct role for mast cells in skin fibrosis. *Clin Exp Allergy*. 2002; 32: 237-246.
189. Grimbaldston MA, Chen CC, Piliiposky AM, Tsai M, Tam SY, Galli SJ. Mast cell-deficient W-sash c-kit mutant Kit W-sh/W-sh mice as a model for investigating mast cell biology in vivo. *Am J Pathol*. 2005; 167: 835-848.
190. Ashman LK. The biology of stem cell factor and its receptor C-kit. *Int J Biochem Cell Biol*. 1999; 31: 1037-1051.
191. Feyerabend TB, Weiser A, Tietz A, Stassen M, Harris N, Kopf M, et al. Cre-mediated cell ablation contests mast cell contribution in models of antibody- and T cell-mediated autoimmunity. *Immunity*. 2011; 35: 832-844.
192. Galli SJ, Kalesnikoff J, Grimbaldston MA, Piliiposky AM, Williams CM, et al. Mast cells as "tunable" effector and immunoregulatory cells: recent advances. *Annu Rev Immunol*. 2005; 23: 749-786.
193. Brown MA, Hatfield JK. Mast Cells are Important Modifiers of Autoimmune Disease: With so Much Evidence, Why is There Still Controversy? *Front Immunol*. 2012; 3: 147.