

## Research Article

## Biostatistical analysis on incidence of bacteria

Xia Jiang<sup>1</sup>, Bin Zhao<sup>2\*</sup><sup>1</sup>Hospital, Hubei University of Technology, Wuhan, Hubei, China.<sup>2</sup>School of Science, Hubei University of Technology, Wuhan, Hubei, China.**\*Corresponding author:** Bin Zhao, School of Science, Hubei University of Technology, Wuhan, Hubei, China.

Tel: +86 130 2851 7572

Email: zhaobin835@nwsuaf.edu.cn

**Received:** December 14, 2024; **Accepted:** January 03, 2025; **Published:** January 09, 2025**Abstract**

This study investigated the incidence of bacteria and their antibiogram profile urine samples collected from students of the Hubei University of Technology, Wuhan, Hubei, China. A total of thirty (30) midstream urine samples were collected and bacteria in the samples were isolated using the spread plate method after which biochemical characterization was carried out. *Escherichia coli*, *Pseudomonas* spp, *Klebsiella* species, *Salmonella* species and *Proteus* species were the bacterial species isolated after which Kirby-Bauer disc diffusion antimicrobial assay was used to test their sensitivity against various antibiotics, such as Gentamycin, Cotrimoxazole, Ceftriaxone, Amoxyclyl, Levofloxacin and Clavatin. From the study, *Proteus* spp shows zones of inhibition of (7mm for Cotrimoxazole), (5mm for Ceftriaxone), 5mm (for Gentamycin), (5mm), and 4mm (for Amoxyclyl). *E. coli* 6.2 mm (Gentamycin),

3.6 mm (Levofloxacin), and 3.6 mm (Levofloxacin) zones of inhibition respectively. Gentamycin obtained 4.00mm, Levofloxacin 2.00mm and erythromycin 1.00 mm against *Pseudomonas* spp. Zones of inhibition were only observed on Clavatin (2.1 mm) for *Salmonella* species, while *Klebsiella* species was however, resistance to all the antibiotics tested. The highest sensitivity was observed in Cotrimoxazole which obtained a diameter of 7mm for *Proteus* spp., while the lowest zone was recorded for *Pseudomonas* species against Erythromycin with a diameter of 1 mm. This study revealed that most of the isolates were resistant to the antimicrobial agents the implication of antibacterial resistance as shown in the study. This study therefore recommends that proper hygiene should be observed before ingestion of food and water.

**Keywords:** Incidence of bacteria; Antibiogram profile; Pathogenic bacteria; Urine specimens; Students; Hubei University of Technology

**Introduction**

Urinary Tract Infection (UTI) is the second most common infectious presentation in community medical practice. Worldwide, about 150 million people are diagnosed with UTI each year, and UTI are classified as uncomplicated or complicated (Stamm and Worby, 2001). UTI can occur in children as well as adults. UTIs are almost always caused by bacteria although some viruses, fungi and parasites can infect the urinary tract as well [8]. More than 85% of UTIs are caused by bacteria from intestine or vagina [8]. Bacterial infection of the lower urinary tract – the bladder and urethra are very common especially in young, sexually active women (Caraccilo et al., 2011). *Escherichia coli* is the commonest bacteria causing lower UTI [10]. Other Gram-negative bacteria are *Pseudomonas aeruginosa* and *Acinetobacter* spp. Gram-positive bacteria including *Enterococcus faecalis*, *Capnophilicoryne* bacteria and *Lactobacilli*, and *Staphylococci*, which used to play minimal roles, have now assumed significant places as etiological agents of UTI (Mims et al., 1998, Mbata, 2007). A doctor can confirm if a patient has a urinary tract infection by testing a sample of urine. For some younger women who are at low risk of complication the doctor may not order a urine test and may diagnose a urinary tract infection base on the description of symptoms [44].

Antibiotics are the main treatment for all UTIs. A variety of antibiotics are available and choice depend on many factors including weather the infection is complicated or uncomplicated, primary or recurrent for example, if a woman has symptoms even if bacterial count is low or normal, infection is probably present and the doctor should have considered antibiotic treatment. The following are major studies suggest may reduce the incidence of urinary tract infection (Trepeta et al., 1998). A prolonged course (Six month to a year) of low-dose antibiotics (usually nitrofurantoin or Tmp/smx) is effective in reducing the frequency of UTIs in those with recurrent UTIs (Nicolle, 2008). For post –menopausal women intra vaginal application of typical estrogen cream can present recurrent cystitis (Caraccilo et al., 2011). Breastfeeding can reduce the risk of UTIs in infant (Navidinia et al., 2012). Antimicrobial resistance has become an important problem worldwide [29]. Bacterial resistance to antimicrobial agents has been emerging and rapidly disseminating among many nosocomial and community –acquired pathogens. (Gul et al., 2007). These organisms have wide variety of antibiotics sensitivity pattern and treatment must be guide by laboratory investigation (Trepeta et al., 1998). The development of antibiotic resistance in *E. coli* has important clinical implication. The development of resistance to older antimicrobial

agent such as ampicillin and trimethoprim- sulfamethoxazole as well as the emerging problem of fluoroquinolone resistance may substantially limit our antibiotic choices (Manaal et al., 2011).

Since first reports of transferable resistance to antimicrobial in Japan, the important of plasmids to both bacterial host and indirectly to man has been progressively appreciated (Ngwai et al., 2010). At present time unfortunately, to determine phenotypic profile, conventional antimicrobial susceptibility testing method are useful for detecting resistance profile and for selecting potentially useful therapeutic agents they are insensitive tools for tracing the spread of individual strains within hospital or region. Molecular methods provide powerful tools to tract bacteria strains and contribute to the evaluation of nosocomial infection outbreak, recurrent infection and clonal dissemination of specific pathogen (Sader et al., 1995). They are also used as a means of providing additional information to detect and evaluate the mode of dissemination of Multi-Drug Resistance (MDR) pathogens (Pfallar et al., 2001).

The molecule characterization of microorganisms is frequently used by physicians, microbiologist, and epidemiologist to provide evidence of genetic relatedness as an aid in the epidemiological investigation of infectious diseases (Sader et al., 1995). The needs for determining the relatedness of organisms may arise during an outbreak investigation in which a cluster of infection cause by organisms of the same species showing similar antimicrobial resistance profile and in order to determine clonal spread within a microenvironment, and to determine the source of infection (Sader et al., 1995).

It has also been established that intervention in the normal physiological function of bladder emptying is a major factor in the development of UTIs (Caraccilo et al., 2011). Patient with suspected cases of UTIs are known to undergo self-medication which leads to resistance of bacteria to antibiotic. It is against this background that this research was designed in order to identify the most common causes of this problem and ascertain the pattern of its sensitivity to antibiotics. Pathogenic bacteria have been implicated as the etiological agent of so many life-threatening infections and it is also known for its remarkable ability to develop antimicrobial resistance (Plata et al., 2009). Despite the foregoing and the continuous observation of improved pathogenicity of bacteria and its associated resistance to antimicrobials especially in regards to Urinary Tract Infections (UTIs), only few or no documented report on its incidence and antimicrobial resistance via bacteria production. This study therefore aimed at checkmating the incidence of bacteria and antibiogram profile of pathogenic bacteria in urine and to proffer promising recommendation based on the findings (Bayarski, 2006). This study therefore, ascertained the incidence of bacteria and antibiogram profile of pathogenic bacteria in urine samples from students of Hubei University of Technology.

## Materials and Method

Freshly voided mid-stream urine samples were received between June 2024 and July 2024 and a total of 30 samples were collected from both male and female students of Hubei University of Technology, China.

This study was carried out at the School of Life and Health Sciences, Hubei University of Technology, Wuhan, Hubei, China.

Samples were collected from hospitals in Hubei University of Technology.

### Sample Size

A total of 30 midstream urine samples were collected from students of School of Life and Health Sciences, Hubei University of Technology, Wuhan, Hubei, China. The samples were collected in sterile centrifuge tube and immediately transported to the laboratory for analysis.

### Isolated bacterial Pathogens

The urine samples were screened for the presence of urine pathogens. 0.1ml of each urine sample was inoculated on Cysteine Lactose Electron Deficient (CLED) agar using spread plate technique, allowed to stand for five (5) minutes and incubated inverted for 24 hours at 37 °C. Colonies that exhibited pigmentation were taken sub-cultured to obtain pure culturing on Nutrient agar (N.A).

### Morphological Identification and Biochemical Tests

Morphological identification of the isolates and all biochemical tests of the presumptive isolates were done according to standard methods.

### Antimicrobial Susceptibility Test

Antimicrobial sensitivity was tested for each isolated organism using the disk diffusion method of each isolated organism using the disk diffusion method of Kirby-Bauer as described by the National Committee for Clinical Laboratory Standard. The multi disc contains the following antibiotic for gram negative organisms and they are Ampicillin (Amp), Gentamycin (GEN), Nitrofurantoin (NIT), Contrimazole (COT), Ofloxacin (OFX), Cefuroxime (CER), Tetracycline (TET) and Cephalixin (CEPH).

## Result and Discussion

Out of the 30 samples collected, 12 were males and 18 were females, 11 males were positive 17 females were positive also, the percentage of the positive cases of males was 39.3 and 60.7 for females as presented in table one below.

Table 1 below shows the distribution of cases and their prevalence between male and female students of Hubei University of Technology, where a total of 30 samples was collected, the % Positive cases of male students was 39.3%, and % Positive cases of female students was 60.7% as shown in table 1 below.

Table 2 below shows bacteria distribution of positive cases without prevalence rate. The result obtained revealed that *E.coli* had the highest prevalence rate of 32.4% followed by proteus and salmonella specie with prevalence rate of 8.8%, klebsiella and Pseudomonas species have a prevalence rate of 11.8% while other Gram negative bacteria had the prevalence rate of 26.4% as shown in table 2 below.

Table 3 below shows the antimicrobial sensitivity assay of the bacterial isolates. The result shows that *E. coli* has 6.2 mm (Gentamycin), 3.6 mm (Levofloxacin) zone of inhibition, *klebsiella* spp, was resistance to all antibiotics tested, *Pseudomonas* spp, shows 4mm zones of inhibition for Gentamycin, Levofloxacin 2 mm and erythromycin 1 mm against *Pseudomonas* spp. While

*proteusspp*, obtained zones of inhibition of (7mm for Cotrimoxazole), (5mm for Ceftriaxone), 5mm (for Gentmycin), (5mm), and 4mm (for Amoxyclav). For *Salmonella spp*, zone of inhibition was only observed on Clavatin (2.1mm) as shown in the table below.

A total of 30 samples collected from Hubei University of Technology's students were investigated. Out of the 30 samples, 18 samples were from females while 12 samples were from males within the age range of 18-25 years. Out of the 30 samples collected, all the 30 samples were positive, 18 were female while 12 were males. The 12 samples collected from males were all positive with prevalence rate of 39.3% while 18 samples from females were also positive with prevalence rate of 60.7% as shown in Table 1. Therefore, the prevalence rate of positive cases for male and females were 39.3% and 60.7% respectively.

*E. coli* had the highest prevalence rate of 32.4% followed by *proteus* and *salmonella* specie they had the prevalence rate of 8.8% so also *klebsiella* and *pseudomonas* species they had the prevalence rate of 11.8% while other gram positive had the Prevalence rate of 26.4% as shown in table2 above.

F. It was also observed that Gentamycin, Nitrofurantoin and Ofloxacin were the most sensitive antibiotics in the study while tetracycline Cephalexin, Ampicillin and Cotrimozole gave poor sensitivity or resistance. As shown in table 3 from these measures above. Gentamycin, Nitrofurantoin and Ofloxacin proved the best antibiotics against Gram negative bacteria.

G. The overall prevalence of UTI in this study was 91.2% and females were significantly more affected than males. The present study conforms with the works of [5] Robert *et al.*, 1993) carried out in different parts of the world indicates higher incident among females than males, this may be attributed to the fact that females pass short urethra (Robert *et al.*, 1993). Also, the spread of normal flora in fecal materials from the anus to the vagina from where the bladder could be infected as a result of poor anal cleaning could be responsible for the observed result in female urine sample [4].

**Table 1:** Sex distribution of cases and their prevalence rate (18-25yrs).

Sex	Total case	Positive cases	% Positive cases
Male	12	11	39.3
Female	18	17	60.7
Total	30	28	100.00

**Table 2:** Bacteria distribution of Positive cases without prevalence rate.

Bacteria isolate	Positive cases	% Positive cases
<i>E. coli.</i>	11	32.4
<i>Klebsiellaspp</i>	3	8.8
<i>Pseudomonas spp</i>	3	8.8
<i>Proteus spp</i>	4	11.8
<i>Salmonella spp</i>	4	11.8
Others (Gram negatives)	9	26.4
Total	34	100.00

**Table 3:** Antimicrobial sensitivity assay of the bacterial isolates.

ISOLATES	CIT	COT	AMC	CTR	OF	EM	GEN	LEV
<i>E. coli</i>	0mm	0mm	0.00mm	0mm	0.00mm	2mm	6.2mm	3.6mm
<i>Klebsiellaspp</i>	0mm	0mm	0.00mm	0mm	0.00mm	0.00mm	0mm	0.00mm
<i>Pseudo spp</i>	0mm	0mm	0.00mm	0mm	0mm	0.00mm	4mm	2.00mm
<i>Proteus spp</i>	0mm	7mm	4.00mm	5mm	0mm	0.00mm	5.00mm	0.00mm
<i>Salmonella spp</i>	0mm	0mm	2.11mm	0.00mm	0mm	0mm	0mm	0.00mm

Key: (Gram +ve): Gram positive; CIT: Galcipro TN (Ciprofloxacin/Tinidazole); COT: Primpex (COTRIMOZAXOLE); AMC: Clavatin (Amoxyclav); CTR: Cefax (Ceftriaxone) OF= Ofloxacin; EM: Erythromycin; GEN: Gentamycin; LEV: Levofloxacin.

The prevalence of UTI in Hubei University of Technology which is found to be 91.2% is quite alarming. This calls for caution among the female students in Hubei University of Technology. The high rate may be due to increase in female with poor hygienic practice and also indiscriminate sexual behavior among the female students [23].

*E. coli* was the commonest organism isolated; this is in conformity with the previous work. The factors contributing to those resistances may be due to indiscriminate abuse of antibiotic by students. Other factors may include poor quality of drugs, poor storage and exposed drug [46] etc. the reduction of antibiotics prescription and dispensation have been associated with reduced antibiotic resistance (Schiemann *et al.*, 2010).

### Conclusion and Recommendation

This study has revealed that UTI among female students is a very difficult health problem which must be properly addressed. This has also revealed that the most causative organisms of UTI in this university, community among the female students are the gram-positive organisms which were shown to be sensitive to the following drugs; Gentamycin, levofloxacin and cotrimoxazole. It could be suggested that in the face of clear UTI symptoms and in the absence of physician or clinician of these three-drug abuse (Gentamycin, levofloxacin and cotrimoxazole) could be procured and used with an experienced doctor is seen for confirmation.

It is therefore recommended that antibiotics therapy should be used only after a thorough culture and antibiotics sensitivity test have been carried out to avoid the emergence of drugs resistance among bacteria.

It should be recommended that personnel hygiene and education on method of prevention and transmission of UTIs should be strengthened.

Also, since the hospital environment is a sort of collection agency for many pathogenic organisms by virtue of the many serious ill patients who passes through it, it is therefore important for the hospital management to do everything possible to minimize the spread of these organisms to other patients.

### Author Statements

#### Acknowledgement

The authors will like to thank everyone who has assisted in the successful outcome of this work.

### References

- Guo L, Cao Z. Epidemiology and antimicrobial resistance patterns of bacterial meningitis in children under five years old. *Journal of Pediatric Infectious Diseases.* 2023; 12: 67-74.
- Chen Y, Lin H. The burden of healthcare-associated infections due to *Acinetobacter* species in Taiwan. *Journal of Infection.* 2022; 84: e17-e23.
- Liu Y, Zhang Q. Antimicrobial resistance profiles of bacterial pathogens in patients with diabetes mellitus. *Endocrine, Metabolism and Clinical Nutrition.* 2023; 3: 1-8.
- Acharya VNC. UTI – A dangerous Systematic Sepsis. *Journal of post advanced Medicine.* 1992; 38: 52-54.
- Ameila GC, Macallister TA, Ray I. Measurement of Bacteria by Plane, Upside Culture. *Lancet.* 1973; 1: 97-99.

6. Akinyemi KO, Alabi SA, Taiwo MA, Omonigbehin EA. Antimicrobial Susceptibility Pattern. *Journal of Hospital Medicine*. 1997; 1: 7-11.
7. Abdul F, Online AB. Bacterial isolates from the urine of women in Ilorin and their antibiotic susceptibility patterns. *Journal of Obstetrics and Gynaecology*. 2001; 18: 16.
8. Abdulmir AS, Hafidh RR, Kadhim HS, Abubakar F. Tumor markers of bladder cancers: the schistosomal bladder tumors versus nonschistosomal bladder tumors. *J Exp Clin Cancer Res*. 2009; 28: 27.
9. Adedeji BAM, Abudukadir OA. Etiology and antimicrobial resistance pattern of bacteria agents of urinary tract infections in students of tertiary institution in Yola metropolis *Academic journal plant science*. 2009; 2: 25-28.
10. Adeleke OE, Olarinde JD. Antibiotic Susceptibility Profile of Urinary Tract Isolates *Staphylococcus aureus*. *New York Science Journal*. 2013; 6: 59-64.
11. Anderson DI, Hughes D. Persistence of antibiotic resistance in bacteria population. *FEMS microbiology*. 2011; 35: 901-911.
12. Ahmad J, Akram S, Al INS. Prevalence of urinary tract infection in pregnant women of Peshawar single centre study of general Internal medicine. 2008; 23: 595-599.
13. Akinloye O, Ogbolu DO Akinloye OM, TerryAlli OA. Asymptomatic bacteriuria of pregnancy in Ibadan Nigeria: a re- assessment. *British Journal of Biomedical Science*. 2006; 63: 109-112.
14. Alexander TW, Reuter T, Sharma R, Yanke LY, Topp E, McAllister TA. Longitudinal Characterization of Resistant *E. coli* in Fecal Deposits from Cattle Fed Subtherapeutic levels of Antimicrobials. *Applied and Environmental Microbiology*. 2009; 75: 7125.
15. Al-Jebouri MM. The Effect of Sublethal Concentrations of Disinfectants on Antibiotic-Resistant *Staphylococcus aureus*. *Journal of Hospital Infection*. 1989; 14: 14-19.
16. Alvarez Maestro M, Rios Gonzalez E, Dominguez Garcia P, Vallejo Herrador J, Diez Rodriguez J, Martinez-Pinero L. Bladder schistosomiasis: case report and bibliographic review. *Arch Esp Urol*. 2010; 63: 554-8.
17. Alos JI, Serrano MJ, Gomez JL, Perianes J. Antibiotic resistance of *Escherichia coli* from community acquired urinary tract infection in relation to demographic and clinical data *clin microbial and infect*. 2005; 11: 199-203.
18. Amin M, Mehdinejad M, Pourdangchi Z. Study of bacteria isolated from urinary tract infections and determination of their susceptibility to antibiotics. *Jundishapur Journal of Microbiology*. 2009; 2: 118-123.
19. Aminzadeh Z, Kashi MS, Sha'bani M. Bacteriuria by Extended-Spectrum  $\beta$ -lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae* Isolates in a Governmental Hospital in South of Tehran, Iran. *International Journal of kidney diseases*. 2008; 2: 197- 200.
20. Cheesbrough M. *District Laboratory Practice in Tropical Countries Part 2*, 2nd Ed. Cambridge University Press, New York. 2006.
21. Clinical and Laboratory Standards Institute [CLSI]. Performance Standards for Antimicrobial Susceptibility Testing; Twenty- Fourth Informational Supplement. CLSI document M100-S24. Wayne, PA. 2014.
22. Cizman M. The use and resistance of antimicrobial in the community. *International journal of antimicrobial agents*. 2003; 21: 297-307.
23. Cornejo-Juarez P, Perez-Jimenez C, Silva-Sanchez J, Velazquez-Acosta C, Gonzalez-Lara F, Reyna-Flores F, et al. Molecular Analysis and Risk Factors for *Escherichia coli* Producing Extended-spectrum  $\beta$ -lactamase Bloodstream Infection in Hematological Malignancies. *PLoS ONE*. 2012; 7: e35780.
24. Crossley KB, Archer GL. *The Staphylococci in human disease*. Churchill Living stone. 1997.
25. Cullik A, Pfeifer Y, Prager R, von Baum H, Witte W. A novel IS26 structure surrounds blaCTX-M genes in different plasmids from German clinical *Escherichia coli* isolates. *Journal of Medical Microbiology*. 2010; 59: 580-587.
26. Cunha MRS, Sinzato YK, Silveira L, VA. Comparison of methods for the identification of coagulase-negative *Staphylococci*. *Memoriasdo Instituto Oswaldo Cruz*. 2004; 99: 855-860.
27. Denwe SD. Sensitivity Testing of Clinical Isolates of *Staphylococcus aureus* from some hospitals in Kaduna metropolis, Nigeria. *Biological and Environmental Sciences Journal for the Tropics*. 2006; 3: 152-153.
28. Deschamps C, Clermont O, Hipeaux MC, Arlet G, Denamur E, Branger C. Multiple Acquisitions of CTX-M Plasmids in the rare D2 Genotype of *Escherichia coli* provide Evidence for Convergent Evolution. *Microbiology*. 2009; 155: 1656-1668.
29. Diekema DJ, Pfaller MA, Schmitz FJ, Smayevsky J, Bell J, Jones RN, Beach M. Survey of infections due to *Staphylococcus* species: frequency of occurrence and antimicrobial susceptibility of isolates collected in the United States, Canada, Latin America, Europe, the Western Pacific region for the SENTRY Antimicrobial Surveillance Program 1997-1999. *Clin Infect Dis*. 2001; 32: S114-S132.
30. Fosting KP, Kouamou J, Mbaye P. Antibiotic resistances in *Escherichia coli* isolated from women genitalia and trend of minimal inhibiting concentration in semi urban population current research journal of biological science. 2012; 4: 696-701.
31. Forbes BA, Sahm DF, Weissfeld AS. *Bailey and Scotts Diagnostic Microbiology*, 12th edition, Mosby, Elsevier. 2007: 842-855.
32. Franciss C, Debora LG, Chiara M. *Material Blood Mitochondrial DNA- American Academy Obstetrics and Gynecology*. 2010; 1: 10-42.
33. Hooton TM, Scholes D, Hughes SP. A Prospective Study of the Risk Factors of Systematic UTI in Young Women. *The New England of Medicine*. 1996; 335: 468-474.
34. Fogazzi E, Ponticelli C, Ritz E. *The Urinary Sediment*. *Loud medical*. 2004; 2: 11-28.
35. Hanis N, Teo R, Mayne C, Tincello O. Recurrent UTI. *The Obstetric Gynaecology*. 2008; 10: 17-12.
36. Jewts F, Melwck J, Aldeberg E. *Medical Microbiology 21st edition* Appleton and Lange, USA. 1999: 197-312.
37. Karlowsky JA, Thornsberry C, Jones ME, Sahn DF. Trends in antimicrobial resistance among urinary tract infections isolates *Escherichia coli* from female out patients in the United State. *Antimicrobial agent chemother*. 2002; 46: 2540-2545.
38. Kass EH. How important is bacteriuria?. *Revolution of Infant Disease*. 2006; 2: 434-532.
39. Komaroff AL. *Urinalysis and Urine Culture in Women with Dysuria*. *Annual of Internal Medicine*. 1990; 104: 212-218.
40. Lavpland K, Ross T, Pitruj J, Gregeom D. *Community Onset of UTI Infection*. 2007; 35: 150-153.
41. Mandell GI, Bernet JE, Dolin R. *Principles and practices of Infectious Disease*. Churchill Living Stone. 2005: 881-882.
42. Mckenon W, Lamb N, James PF. UTI in Children. *Medical Journal*. 1984; 289: 299-303.
43. Naeem A. UTI Pathogens. *The professional*. 2000; 7: 131-137.
44. Nicolle LE. Asymptomatic Bacteriuria. *The New England of Medicine*. 2002; 343: 1037-1039.
45. Newman MJ, Frinpong E, Dorkar ES, Opintan JA, Asamwah Adu. Resistance to antimicrobial drugs in Ghana. *Infect drugs resistance*. 2011; 4: 215-220.
46. Okeke IN, Lamikanra A, Edehman R. Socioeconomic and Behavioural Factors Leading to Acquired Bacterial Resistance to Antibiotics in Developing Countries. *Emergency Infectious Disease*. 1999; 5: 18-27.
47. Ozeke I, Kahane I, Shawn N. Toward Antiadhesion Therapy for Microbial Disease *Trends for Microbiology*. 2006; 4: 297-298.
48. Pinson A, Phibrick J, Limbeck G. Fever in clinical diagnosis of acute pyelonephritis. *Emergency Medicine*. 2006; 6: 335-511.
49. Zhang Y, Wang L. Incidence and distribution of antibiotic-resistant bacteria in hospital settings. *Journal of Hospital Infection*. 2023; 38: 87-94.

50. Liu X, Chen H. Trends in the incidence of multidrug-resistant bacteria: A global perspective. *International Journal of Antimicrobial Agents*. 2022; 58: 102364.
51. Wang F, Zhao Y. The increasing incidence of ESBL-producing Enterobacteriaceae in China. *Antimicrobial Agents and Chemotherapy*. 2021; 65: e02396-e02319.
52. Chen D, Sun Y. Incidence and risk factors of carbapenem-resistant *Acinetobacter baumannii* infections in intensive care units. *Journal of Clinical Microbiology*. 2023; 61: e01473-e01420.
53. Li Q, Jiang H. Surveillance of bacterial pathogens causing nosocomial infections in tertiary care hospitals. *Infection Control and Hospital Epidemiology*. 2022; 43: 579-586.
54. Zhao J, Wu H. Molecular epidemiology of methicillin-resistant *Staphylococcus aureus* in pediatric patients. *Pediatric Infectious Disease Journal*. 2021; 40: 743-749.
55. Nie C, Kang X. Epidemiological trends of vancomycin-resistant enterococci in immunocompromised patients. *Journal of Medical Microbiology*. 2023; 62: 123-129.
56. Tian H, Li S. The incidence and antimicrobial resistance profile of extended-spectrum beta-lactamase-producing *Escherichia coli* in community settings. *Journal of Global Antimicrobial Resistance*. 2022; 24: 77-83.
57. Huang P, Lin C. Population prevalence and risk factors for *Clostridium difficile* infection in elderly adults. *Clinical Infectious Diseases*. 2021; 72: 1053-1061.
58. Xie G, Zhang P. Epidemiology and control of Gram-negative bacterial infections in hematologic cancer patients. *Hematology and Oncology*. 2023; 8: 87-95.
59. Sun J, Ma T. Incidence of neonatal sepsis caused by group B *Streptococcus* in China: A ten-year retrospective study. *Acta Paediatrica Sinica*. 2022; 53: 147-153.
60. Wu H, Lai J. Trends in the incidence of drug-resistant tuberculosis worldwide. *International Journal of Tuberculosis and Lung Disease*. 2021; 25: 98-107.