

Review Article

Emerging Herbal Bio-actives as Reformed Dipeptidyl Peptidase (DPP-IV) Inhibitors Used for the Management of Diabetes Mellitus: A Brief Review

Sakshi Sharma^{1*}; Sonia Chauhan²¹Swift School of Pharmacy, Ghaggar Sarai, Tehsil Rajpura, District Patiala, Punjab – 140401, India²Noida Institute of Engineering and Technology (Pharmacy Institute), Greater Noida, UP, India***Corresponding author: Sakshi Sharma**

Swift School of Pharmacy, Ghaggar Sarai, Tehsil Rajpura, District Patiala, Punjab – 140401, India.

Email: sakshi.sharma1964@gmail.com

Received: April 06, 2024**Accepted:** May 08, 2024**Published:** May 15, 2024**Abstract**

Background: Diabetes mellitus is a severe metabolic disorder affecting nearly half of the population worldwide and increase patient risk by its other related complication. DPP-IV is a serine aminopeptidase, which plays a crucial part in the glucose metabolic process and causes incretin degradation in GLP-1. It is one of the validated targets for the treatment of Type 2 Diabetes because of its effect of incretin hormone. Nearly seven DPP-4 inhibitors are in the market for treating Type -II diabetes (i.e., Sitagliptin, Vildagliptin, Saxagliptin, Linagliptin, and Alogliptin). They all are synthesized chemically, having good therapeutic efficacy, but their long-term safety use is unknown.

Methods: Moreover, plant-derived products are available in the market and found to be safe and effective, and nearly half of the population consumes these natural or bio-active compounds. Therefore, for developing novel anti-diabetic drugs, these natural products or herbal medicine seems to have good utility.

Result: The present study deals with the progress of new herbal medicine, their crude extract with the mechanism of action having DPP-IV inhibitory activity. Therefore, these could be helpful evidence for developing the next generation of anti-diabetes medicines via inhibiting DPP-4 activity.

Conclusion: The researchers investigating the novel lead for managing diabetes (type II) can also search these natural bioactive for future use. We have summarised the reported herbal Bio-actives/Plants for the management of Diabetes mellitus from the year 2001- 2022.

Keywords: Diabetes mellitus; Bioactives; DPP-IV; Extract; Incretin; Medicinal plants

Abbreviation: WHO: World Health Organization; DM: Diabetes Mellitus; GIP: Glucose-dependent insulin-tropic hormone; GLP: Glucogen like peptide; DPP-IV: Dipeptidyl peptidase

Introduction

Diabetes Mellitus (DM) is a common metabolic disorder that is increasing rapidly in the growing era, and according to WHO data, it is considered a primary cause of death worldwide [1]. Amongst all types of diabetes, Type 2 is the most common disorder having 90% of cases. Patient with Type 2 diabetes mellitus shows no apparent symptoms, but still, it is a life-threatening condition. According to the International Diabetes Federation, the cases of diabetes will increase from 415 million (in 2015) and might be increased 642 million (in 2040) [2]. Numerous

synthetic drugs are introduced to lower the blood glucose level (i.e., biguanides, incretin mimetics, α glucosidase inhibitor, and DPP-IV inhibitor) [3]. The hormone incretin plays a significant role after ingesting food; GIP and GLP-1 are the two essential enzymes secreted by Incretin that stimulate the insulin release from the β pancreatic cells [4]. Incretin shows the insulinotropic effect; it decreases food intake, inhibits glucagon and gastric emptying time, and slows down glucagon secretion [5]. The action of Incretin (GLP- ad GIP-1) mainly depends on the two

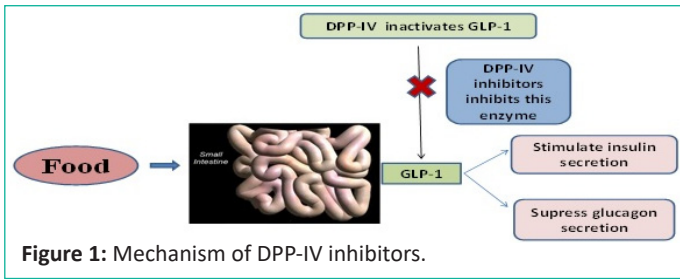


Figure 1: Mechanism of DPP-IV inhibitors.

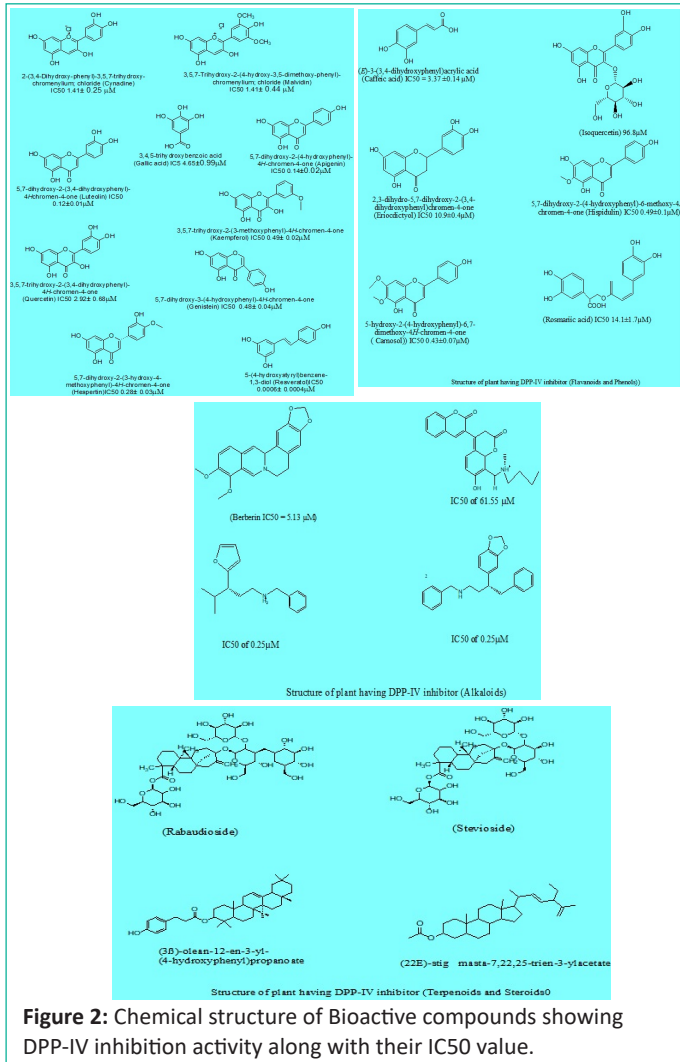
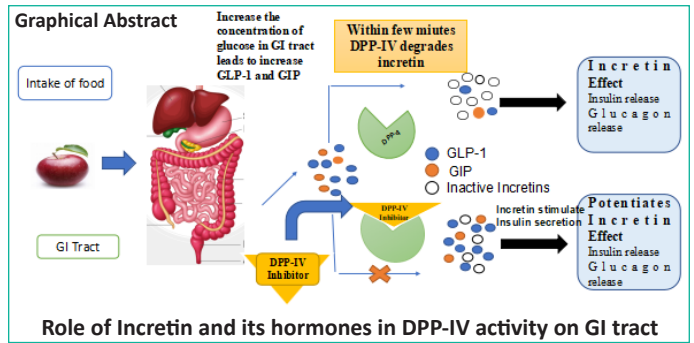


Figure 2: Chemical structure of Bioactive compounds showing DPP-IV inhibition activity along with their IC50 value.

Table 1: List of synthetic DPP-IV inhibitors available in the market [14].

Marketed drugs	Vildagliptin	Saxagliptin	Sitagliptin	Linagliptin	Alogliptin
DPP-IV inhibitory potential	More than 95%	70-85%	More than 85-95%	More than 75%	More than 85%
Effectivity	Highly effective	Moderately effective	Very highly effective	It shows dose-dependent inhibition and highly effective	Shows high-affinity ad inhibition in a dose-dependent manner
Hypoglycemia risk	No risk detected	No risk detected	No risk detected	No risk detected	No risk detected
Half-life	1.4-1.5hr	2.4hr	7-14hr	11-12hr	12.5 to 24hr
Metabolism	Metabolized through kidney	Metabolized through kidney	Only 16% metabolised through kidney	Primary exertion through bile and secondary through kidney	Metabolized by cytochrome enzymes.
Approved countries	Europe	Europe and the United States	United States and Europe	Europe	Under Investigation
Brand name	Galvus	Onglyza	Januvia	Tradjenta	Nesina
Bioavailability	±85%	±67%	±100%	±30%	±100%
Structures					



N-terminal amino acids that DPP-IV removes. Within 1-2 min, the Incretin is metabolized into inactive form by enzyme DPP-IV and only (10 to 15%) circulating stimulates the pancreas [6]. DPP-IV is an enzyme belonging to the family of serine proteases which involves the degradation of peptide hormones (i.e., glucagon-like peptide and cytokinin) [7]. DPP-IV inhibitors are now widely used in the market. These inhibitors inhibit the enzyme and promote the glucose homestasis [8]. But these synthetic drugs have good therapeutic activity, but they are not safe for the long term, so there is an increase in attention towards plant-derived products; they're bioactive and used to treat type 2 diabetes. Therefore, this review aims to compile the various herbal plants, their extract, mechanism of action, IC50 value, and structure of chemical compounds during 2000-2022, depicted in Table 1 and Table 2.

Mechanism of DPP-IV Action

DPP-IV is an amino peptidase enzyme with 760 amino acids. It contains two subunits having two domains (N terminal domain and C terminal domain). It is already known that GIP (Glucose-dependent insulin-tropic hormone) and GLP-1 (Glucagon like peptide hormone) are incretin hormones that stimulate the release of insulin from pancreatic islets [10]. After food ingestion, these two incretin hormones release and contribute to insulin secretion [11]. DPP-IV enzymes mainly metabolize the Incretin (GLP and GIP-1), which release in response to food ingestion. DPP-IV inhibitors come under the class of oral hypoglycemic drugs, which competitively inhibit the enzyme DPP-IV and prevent the degradation of incretin hormone (GIP and GLP-1), ultimately increasing insulin secretion [12].

Table 2: List of reported medicinal plants having DPP-IV inhibitory activity.

S.No	Plant/Family	Common name	Dose mg/kg & ug/ml	Part used	Extract	Mechanism	IC50 value/% inhibition	Reference
1	<i>Pueraria tuberosa</i> (Fabaceae)	Kudzu	50 &100	Roots	Water	Inhibit DPP-IV enzyme	-	[15]
2	<i>Berberis arista</i> (Berberidaceae)	Indian barberry	500	Bark	Methanolic	Affect DPP-IV and endocrine system activity	14.46 µg/ml	[16]
3	<i>Camellia Sinensis</i>	White tea	200 µM	leaves	Ethanolic	Increase insulin secretion	227µg/ml	[17]
4	<i>Castanospermum australe</i> Cunn.	Blackbean	100 &150	Seeds	-	Reduced blood glucose level, HbA1c, and insulin	13.96 µg/ml	[18]
5	<i>Fagonia critica</i> (Zygophyllaceae) & <i>Hedera nepalensis</i> K. (Araliaceae)	Dhamasa Albumbar	-	Aerial	Ethyl acetate	Inhibit DPP_IV enzyme	38.1 µg/ml 17.2 µg/ml	[19]
6	<i>Desmodium gagicum</i> (Legumioiseae)	Sarivan	1000µg/ml	Aerial	Aqueous	Decrease the inactivation of GLP-1 and thereby increase its concentration and its action	255.5µg/ml	[20]
7	<i>Mangifera indica</i> (Anacardiaceae)	Mango	-	Leaves	Methanolic	Increase the level of GLP-1, improve glucose tolerance, and enhance insulin secretion	182.7µg/ml	[21]
8	<i>Withania Somifera</i> (Solanaceae)	Ashwagandha	5,25,125µg/ml	Roots, leaves, Fruit	Methanolic	It decreases the blood sugar level by increasing the GLP-1 in the body	8.76µg/ml	[22]
9	<i>Enicostemma lit-torale</i> (Gentianeae)	-	100µl	Whole plant	Ethyl acetate	It increases insulin secretion by increasing GLP -1 half-life	165.64µg/ml	[23]
10	<i>Berberis aristata</i> (Berberideaceae)	Oregon grapes	500µg/ml	Bark	Methanolic	It increases insulin secretion	14.4µg/ml	[24]
11	<i>Corylus avellana</i> L. (Betulaceae)	Hazelnut	50µl	nuts	Aqueous	-	24.7µg/ml	[25]
12	<i>Commiphora Mukul</i> (Burseraceae)	Guggul	200mg/kg	Gum	Hydroalcoholic	Decrease in blood glucose level and HA1c	16.45µg/ml	[26]
13	<i>Tinospora crispa</i> L. (Menispermaceae)	Guduchi	-	Stem	-	Increase the incretin and GLP-1 levels, which increases the insulin secretion in the body	65.86±1.02	[27]
14	<i>Curculigo latifolia</i> (Hypoxideaeae)	Lamba	30µl	Root, Fruit	Aqueous	It increases insulin secretion and glucose uptake	66.15 ± 4.09% 42.79 ± 1.47%	[28]
15	<i>Eucalyptus citriodora</i> (Mytraceae)	Lemon scented gum	250 mg/5 mL/kg	Leaf	Ethanolic	It increases insulin secretion and decreases glucose absorption	9-25%	[29]
16	<i>Urena Lobata</i> (Malvaceae)	Caserweed	625, 1 250, 2 500, 5 000 and 10 000 mg/mL	Leaf	Ethanolic	Prevents GLP degradation and increases insulin secretion	6 489.88 mg/mL	[30]
17	<i>Abelmoschus manihot</i> (L.) (Malvaceae)	Hibiscus	312.5; 625; 1250; 2500; 5000; and 10000 µg/mL	Leaves	Ethanolic	-	860.67 µg/mL	[31]
18	<i>Senna (Cassia) nigricans</i> (Fabaceae)	Sannai	17.3 µU/µl)	Whole plant	Methanolic	It breaks the incretin level and increases the GLP level	63.1±4.67%	[32]
19	<i>Ferula Assafoetida</i> (Umbelliferae)	Fennel	-	Seeds	Methanolic	It breaks the incretin level and increases the GLP level	24.5%	[33]

20	<i>Pueraria tuberosa</i> (Fabaceae)	Kudzu	95µl	Roots	Aqueous	It decreases glucose production and increases GLP-1 circulation in the body.	-	[34]
21	<i>Lens culinaris</i> (Fabaceae)	Lentil	-	Seeds	Ethanollic	-	51.69 ± 4.83µm	[35]
22	<i>Aloe vera</i> (Liliaceae)		30µl	Leaves	Ethanollic	Increase the plasma insulin ad GLP-1 level	8.59 ± 2.61 mM,	[36]
23	<i>Trigonella foenum graecum</i>	Fenugreek	320µg/ml	Seed	Methanollic	Increase the plasma insulin ad GLP-1 level	80.15%	[37]
24	<i>Annona squamosa</i> (Annonacin)	-	250mg/5ml	Leaves	Ethanollic	It suppresses the level of the Dpp-iv enzyme and increases the incretin level	33%	[38]
25	<i>Spirulina platensis</i> (Cyanophycean)		200µm	Whole plant	Butanollic	It suppresses the level of DPP-IV enzyme and increases the incretin level	70%	[39]
26	<i>Boesenbergia pandurata</i> Roxb (Zingiberaceae)	Fingerroot	-	Whole Plant	Ethanollic	It increases the level of GLP-1 and Incretin	-	[40]
27	<i>Pergularia extensa</i> Chiov (Asclepiadaceae)	Daemia extensa	10-30µm	Whole plant	Methanollic	It suppresses the level of DPP-IV enzyme and increases the incretin level	-	[41]
28	<i>Ephedra foeminea</i> (Ephedraceae)	Jordan	0.1ml	Aerial	Methanollic	It increases insulin release and decreases the glucose uptake	50%	[42]
29	<i>Picrorhiza kurroa</i> (Plantaginaceae)	Kutki	25µm	Whole plant	-	Increase the plasma insulin ad GLP-1 level	52.51 ± 5.71 nM	[43]
30	<i>Castanospermum australe</i>	-	6.4µg/ml	Seed	Ethanollic	It increases glucose utilization and increases the glycogen stores in the liver.	13.96 g/ml	[44]
31	<i>Coreopsis Lanceolata</i> (Compositae)	lance-leaved coreopsis	100µg	Flower	Methanollic	It increases insulin secretion and glucose uptake	87.2%	[45]
32	<i>Nauclea latifolia</i> (Rubiaceae)	Pin cushion tree	-	Leaf	Aqueous Ethanollic	-	89.6%	[46]
33	<i>Trillium govanianum</i> (Melanthiaceae)	Naag Chattri	181.3 ± 30.2 mg/g	Rhizome	-	It helps in managing the glucose level in the body.	17.68 ± 1.32 µM	[47]
34	<i>Angelica keiskei</i> Kodzumi (Apiaceae)	-	-	Stem Leaves	Ethanollic	It helps in reducing the plasma glucose level in the body	10.49 µM	[48]
35	<i>Phaseolus vulgaris</i> L. (Fabaceae)	Black Bean	1mg/ml	Bean	-	It helps in decreasing glucose uptake and increases the level of Incretin.	96.7%	[49]
36	<i>Quercus variabilis</i> Blume (Fagaceae)	Acorn	25µl	-	Ethanollic	It inhibits the digestive enzymes and glucose transporters.	5.25 mg/mL)	[50]
37	<i>Prunus amygdalus</i> (Rosaceae)	-	15ml	Seed	Methanollic	It inhibits the enzyme Incretin and DPP-IV	162.9(Ig/mL)	[51]
38	<i>Lagerstroemia speciosa</i> (Lythraceae)	Banaba	-	Leaves	Methanollic extract	It inhibits the enzyme Incretin	60.22±2.01	[52]
39	<i>Rhinacanthus nasutus</i> (L.) (Acanthaceae)	Snake jasmine	-	Leaves	Methanollic	It inhibits the digestive enzymes and glucose transporters	34.4 ig/mL	[53]
40	<i>Allium Sativum</i> (Alliaceae)	Garlic	10 µL	Bulb	Methanollic	It decreases the blood glucose level, increases the Incretin, and increases insulin secretion.	70.88 µg/mL	[54]
41	<i>Artemisia Judaica</i> (Compositae)	Common Mugwort	-	Aerial	Hydro-methanollic	It inhibits the enzyme DPP-IV	85.89 µg/mL	[55]

42	<i>Terminalia Arjua</i> (Comitaceae)	Arjun	-	Bark	-	It helps in reducing the plasma glucose level in the body	28%	[56]
43	<i>Eucalyptus citriodora</i> (Myrtaceae)	lemon- or citron-scented gum	8 mU/mL	Leaves	Ethanollic	It increases the incretin level and decreases glucagon secretion	52%	[57]
44	<i>Spirulina platensis</i> (Cyanophyceae)	-	200 µM	Leaves	Ethanollic	It increases the icreti level y increasing the GIP-1 ad GLP	-	[58]
45	<i>Melicope Gabra</i> (Rutaceae)	Tenggek burung	50µL	Leaves Stem	Chloroform	It helps in reducing the plasma glucose level in the body	619.31±9.21µg/ml	[59]
46	<i>Tenebrio molitor</i>	yellow Mealworms	20µL	-	Hydroalcoholic	It increases the incretin level in the body	(57.56 ± 2.59%)	[60]
47	<i>Pisum Sativum</i> (Fabaceae)	Pea	100µm	seed	-	It formed the hydrophobic bond with the S1 pocket in DPP-IV and inhibited the DPP-IV enzyme.	11.04µm	[61]
48	<i>Brassica oleracea</i> (Brassicaceae)	Broccoli	-	Stem Leaves	Hydrolysis	It increases the incretin level in the body	99.68µm	[62]
49	<i>Chenopodium quinoa Willd</i> (Chenopodiaceae)	Quinoa	25µl	-	N-hexane	It stimulates gastrointestinal digestion and inhibit the DPP-IV enzyme.	3.40 ± 0.20 mg/mL	[63]
50	<i>Heritiera fomes</i>	Sundari	10µl	Bark	Ethanollic	It increases the insulin release	-	[64]
51	<i>Commiphora Mukul</i> (Combiteacea) and <i>Phyllanthus emblica</i>	Guggul Amla		Gum Fruit	Hydroalcoholic	Increase the incretin and GLP-1 levels, which increases the insulin secretion in body	0.36 µM 0.8 µg/mL	[65]
52	<i>Ocimum Sativum</i> (Labiataeae) <i>Momordica Charantia</i> (Cucurbitaceae)	Tulsi Karela	0.1 to 0.5mg/ml	Leaves	Methanollic	It activates the GLP-1 ad GIP ad increase the incretin level	66.81±0.05% 53.25±0.04%	[66]
53	<i>Palmaria palmata</i> (Palmariaceae)	Dulse	-	Species	Aqueous	It helps in reducing the plasma glucose level in body	1.47 ± 0.09mg/ml	[67]
54	<i>Lippia graveolens</i> (Verbenaceae)	Wild oregano	50 µL	Seeds	Methanollic	It inhibits the DPP-IV enzyme and decrease glucose secretion in the body	(3.9 ± 0.6 µM)	[68]
55	<i>Rosa Gallica</i> (Rosaceae)	Gallic source	100µM	Flower buds	Aqueous	It decreases glucose and increases insulin secretion, and increases the GIP and GLP-1 levels.	25.8 µM and 70%	[69]
56	<i>Antidesma madagascariense Lam.</i> (Euphorbiaceae)	Bois bigaignon bâtard	-	Leaves	Ethyl acetate	Increase the incretin and GLP-1 levels, which increases the insulin secretion in the body.	79.2 ± 2.8 µg/ml	[70]
57	<i>Aronia arbutifolia</i> (L.) (Rosaceae)	Red Chokeberry	-	Juice	Fractioned	It decreases the plasma glucose level in the body and inhibits the DPP-IV enzyme	81%	[71-72]
58	<i>Allophylus Cominia</i> (Sapindaceae)	Panigera	-	Leaves	Aqueous	It helps in reducing the plasma glucose level	344.3	[73]
59	<i>Calocybe Indica</i>	Mushroom	100µl	-	Ethanollic	It controls the glucose level and increases GLP, GIP-1.	60.91µg/mL	[74]
60	<i>Senna Nigricans</i> (Fabaceae)	-	1 µg/µl	Powder	Methanollic	It increases the incretin level	56.43%	[75]
61	<i>Helichrysum arearium</i> (Asteraceae)	Dwarf everlasting	-	Flower	Methanollic Ethanollic	It inhibits the increase in the blood glucose level	41.2 µg/ml 16.0µg/ml	[76]
62	<i>Mytilus edulis</i> (Mytillidae)	Blue mussel	-	-	Protein	It inhibits the enzyme incretin and increases the half-life of GIP and GLP-1.	0.66±0.17 mg/ml	[77]
63	<i>Abelmoschus manihot</i> L. Medic (Malvaceae)	Gedi	20 µL	Leaves	Ethanollic	It inhibits the DPP-IV enzyme and increases the incretin level	860.87 µg/mL	[78]

Table 3: List of herbal phytoconstituents having DPP-IV inhibitory activity.

S.No	Phytoconstituents	Biological plants/ compound	Description	IC50 value/% inhibition	Researchers	Reference
1	Alkaloids	<i>Coptis chinensis</i> (Berberine)	From this plant, 9 bioactive were selected for <i>in vitro</i> bioassay, of which 7 had DPP-IV activity (IC50<1.00 mM).	13.3 µM	Guasch et al	[79] [80]
2	Glycosides	<i>Lens culinaris</i> (kaempferol) <i>Lens culinaris</i> (Robinin)	Four compounds were isolated from the seeds of this plant and tested for their DPP-IV inhibitory activity. Robinin was isolated from the seeds of this plant and tested their DPP-IV inhibitory activity.	51.9±4.83 µM 37.01 µM	Hyo Young Kim <i>et al.</i> Kim et al. 1998	[81]
3	Flavanoids	<i>Lippia graveolens</i> & <i>Rosmarinus officinalis</i> L (Naringeni) <i>Camellia Sinesis</i> (Theaceae)	In this, it was reported that six bioactive from these plants showed potent DPP-IV inhibitory activity. Epigallocatechin gallate	2.5±0.3 µM 10.21 µM	Fan J <i>et al.</i>	[82]
4	Terpeoids and Steroids	<i>Stevia rebaudiana</i> (Rebaudioside A) (Stevioside)	Molecular docking identified two main compounds from this plant for DPP-IV inhibitory activity.	-	Ayachi et al.	[83]
5	Phenols and Stilbenoid	<i>Vitis thunbergii</i> var. ((+)-Hopeaphenol, (+)-vitamin A, and (-)-vitisin B	The ethanolic extract from the stems and leaves of plants processes DPP-IV inhibitory activity.	401 90.75 15.3 µM	Lin et al.	[84]

Marketed Available Drugs Having DPP-IV Inhibitor Activity

DPP-IV is a new class of oral anti-diabetic drugs. These drugs are active orally, safer, tolerable, and with a low risk of hypoglycemia. Various drugs are available in the market to inhibit the DPP-IV enzyme, out of which three DPP-IV inhibitors are under investigation. First, reversible drug analogs, including (pyrrolidines and thiazolidines, secondly covalently modifying analogs, including Cyanopyrrolidines and lastly, non-peptidase reversible analogs, including xanthines and aminomethyl pyrimidines [13]. These drugs have specific adverse effects and increase the risk of cardiovascular-related complications and tumor incidence. Table 1 summarizes the marketed drugs used in the management of diabetes mellitus which are approved by FDA.

Medicinal Plants Having DPP-IV Inhibitory Activity

In today's era, the use of herbal medicine is increasing very rapidly. This herbal medicine showed a promising effect in managing diabetes and its related complication with a lesser risk of side effects and greater acceptance. In this, we have compiled **Table 4:** List of clinical trial data on DPP-IV inhibitors⁸⁵.

NCT No.	Condition	Intervention	Characteristics	Age group	Sex	No. of patient
NCT01588587	Diabetes Mellitus (Type II)	1 st Drug: Sitagliptin 2 nd Drug: Vildagliptin	<u>Observational study type</u> <u>Model: Cohort studies</u> <u>Measures outcomes:</u> Cancer frequency, AGE concentration	25-95years	All type	500 patients
NCT00411411	Diabetes (Type II)	1 st Drug: Januvia 2 nd Drug: Placebo	Interventional study type Model: Randomized studies, Parallel assessment	18-88 years	All type	49 patients
NCT01545024	Diabetes Mellitus (Type II)	1 st Drug: Sitagliptin (50mg one time per day os)	Model: Observational	20-95years	All type	60 patients
NCT00111631	Diabetes Mellitus (Type II)	1 st Drug: DPP-IV inhibitor 2 nd Drug: Metformin 3 rd : Placebo	Model: Interventional (phase 2)	18-75 years	All type	218 patients
NCT03602638	Diabetes Mellitus (Type II) DPP-IV inhibitor GLP-1	1 st Drug: Sitagliptin 2 nd Drug: Acarbose	Model: Interventional (Phase 4)	18-80 years	All type	300 patients
NCT01937598	Diabetes Mellitus (Type II)	1 st Drug: Placebo 2 nd Drug: Sitagliptin 3 rd Drug: Liraglutide	Model: Interventional (Phase 3)	25-75years	All type	16 patients

the data of a list of medicinal plants from natural sources, active parts of plants, their extract, and mechanism of action, along with their IC50 value from 2001-2022, depicted in (Table 2). In this we have summarized the medicinal plants like *Pueraria tuberosa*, *Berberis arista* (14.46 µg/ml), *Mangifera indica* (182.7µg/ml), *Desmodium gangeticum* (255.5µg/ml), *Withania Somifera* (8.76µg/ml).

Natural Phytoconstituents Showing DPP-IV Inhibition Activity

The herbal plants, their extracts, and their phytoconstituents have been encouraging throughout the globe since ancient times. Various herbal-based potent leads have been manufactured through integrated methodologies. Most of the herbal leads showed a potent inhibitory effect against DPP-IV. The numerous medicinal plants are recognized and categorized according to their chemical structure, as depicted in Table 3. The various categories of drugs used for DPP-IV, their characteristics, age group of the patient and number of patients involved in the studies, are tabulated in Table 4 and ongoing clinical trials in Table 4.

Conclusion

Over the past few decades, the enzyme DPP-IV considers a promising target for the management of Diabetes mellitus. Moreover, some synthetically manufactured compounds are commercially available in the market, but they have been associated with several side effects, and their long-term use is still unknown. DPP-IV inhibitors work by inhibiting the incretin hormone or increasing the GLP-1 and GIP levels via stimulating insulin secretion and inhibiting glucagon secretion. Newer drugs like molecular docking can potentially manage diabetes using some natural products. *In-vivo* and *In-vitro* experiments studies are recommended for targeting the DPP-IV enzymes. Traditionally, many natural plants and bioactive extracts are used to treat diabetes. Therefore, the current data gives researchers, the industrial sector, and others a new idea for developing novel formulations but still needs more advanced studies in preclinical and clinical to prove their efficacy and safety in human beings.

Author Statements

Conflict of Interest

The authors declare that there is no conflict of interest.

Acknowledgment

The authors thank NIET (Pharmacy Institute) greater noida for supporting and guiding them.

References

- Wild S, Roglic G, Green A, Sicree R, King H. Global prevalence of diabetes: estimates for the year 2000 and projections for 2030. *Diabetes care*. 2004; 27: 1047-53.
- Barnett A. DPP-4 inhibitors and their potential role in managing type 2 diabetes. *International journal of clinical practice*. 2006; 60: 1454-70.
- Alberti KG, Zimmet PZ. Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: Diagnosis and classification of diabetes mellitus. Provisional report of a WHO consultation. *Diabetic medicine*. 1998; 15: 539-53.
- Kim W, Egan JM. The role of incretins in glucose homeostasis and diabetes treatment. *Pharmacological reviews*. 2008; 60: 470-512.
- Pigeon RL, Quddusi S, Paty B, D'Alessio DA. Suppression of glucose production by GLP-1 independent of islet hormones: a novel extrapancreatic effect. *American Journal of Physiology-Endocrinology and Metabolism*. 2003; 285: E701-7.
- Mentlein R, Dahms P, Grandt D, Krüger R. Proteolytic processing of neuropeptide Y and peptide YY by dipeptidyl peptidase IV. *Regulatory peptides*. 1993; 49: 133-44.
- Richter B, Bandeira EE, Bergerhoff K, Lerch C. Dipeptidyl peptidase-4 (DPP-4) inhibitors for type 2 diabetes mellitus. *Cochrane Database of Systematic Reviews*. 2008.
- Kieffer TJ, Francis Habener J. The glucagon-like peptides. *Endocrine Reviews*. 1999; 20: 876-913.
- Langley AK, Suffoletta TJ, Jennings HR. Dipeptidyl peptidase IV inhibitors and the incretin system in type 2 diabetes mellitus. *Pharmacotherapy: The Journal of Human Pharmacology and Drug Therapy*. 2007; 27: 1163-80.
- McKennon SA, Campbell RK. The physiology of incretin hormones and the basis for DPP-4 inhibitors. *The Diabetes Educator*. 2007; 33: 55-66.
- Neumiller JJ, Wood L, Campbell RK. Dipeptidyl peptidase-4 inhibitors for treating type 2 diabetes mellitus. *Pharmacotherapy: The Journal of Human Pharmacology and Drug Therapy*. 2010; 30: 463-84.
- McIntosh CH. Incretin-based therapies for type 2 diabetes. *Canadian Journal of Diabetes*. 2008; 32: 131-9.
- Bhavya K, Purohit MN. Research and Reviews: *Journal of Chemistry*. 2013; 2: 1-6.
- Gupta R, Walunj SS, Tokala RK, Parsa KV, Singh SK, Pal M. Emerging drug candidates of dipeptidyl peptidase IV (DPP IV) inhibitor class for treating type 2 diabetes. *Current drug targets*. 2009; 10: 71-87.
- Martins BD. Effects of incretin-based therapies on the gastrointestinal motility of an animal model of Multiple Sclerosis (Doctoral dissertation, Universidade de Coimbra). 2018.
- Srivastava S, Shree P, Tripathi YB. Active phytochemicals of *Pueraria tuberosa* for DPP-IV inhibition: in silico and experimental approach. *Journal of Diabetes & Metabolic Disorders*. 2017; 16: 1-9.
- Chakrabarti R, Bhavtaran S, Narendra P, Varghese N, Vanchhawng L, Mohamed Sham SH, et al. Dipeptidyl peptidase-IV inhibitory activity of *Berberis aristata*. *J Nat Prod*. 2011; 4: 158-63.
- Bharti SK, Krishnan S, Kumar A, Rajak KK, Murari K, Bharti BK, et al. Antihyperglycemic activity with DPP-IV inhibition of alkaloids from seed extract of *Castanospermum australe*: Investigation by experimental validation and molecular docking. *Phytomedicine*. 2012; 15: 20: 24-31.
- Saleem S, Jafri L, Haq I, Chang LC, Calderwood D, Green BD, Mirza B. Plants *Fagonia cretica* L., and *Hedera nepalensis* K. Koch contain natural compounds with potent dipeptidyl peptidase-4 (DPP-4) inhibitory activity. *Journal of Ethnopharmacology*. 2014; 28: 156:26-32.
- Bisht R, Bhattacharya S, Jaliwala YA. Evaluating the use of *Desmodium gangeticum* as Alpha Glucosidase and DPP-IV Inhibitor for Type-II Diabetes. *Am J Phytomed Clin Ther*. 2014; 2: 530-9.
- Kempegowda PK, Zameer F, Narasimashetty CK, Kollur SP, Murari SK. Inhibitory potency of *Withania somnifera* extract against DPP-4: an in vitro evaluation. *African Journal of Traditional, Complementary and Alternative Medicines*. 2018; 15: 11-25.
- Haque MA, Konatham TK, Gawai N, Jyothi A, Vidhya B, Saboo S, et al. In-vitro Antioxidant and DPP-IV Enzyme Assay of Ethyl Acetate Extract of *Enicostemma littorale*. *Journal of Pharmaceutical Research International*. 2022; 34: 50-8.
- Yogisha S, Raveesha KA. Dipeptidyl Peptidase IV inhibitory activity of *Mangifera indica*. *J Nat Prod*. 2010; 3: 9.
- Chakrabarti R, Bhattarai S, Narendra P, Varghese N, Vanchhawng L, Mohamed Sham Shihabudeen H, et al. Dipeptidyl peptidase-IV inhibitory activity of *Berberis aristata*. *J Nat Prod*. 2011; 4: 158-63.
- Çağlar AF, Göksu AG, Çakır B, Gülseren İ. *Tombul hazelnut (Corylus avellana L.)* peptides with DPP-IV inhibitory activity: In vitro and silico studies. *Food chemistry*. 2021; 12: 100151.
- Borde MK, Mohanty IR, Maheshwari U, Suman RK, Deshmukh YA. DPP-4 inhibitory activity and myocardial salvaging effects of *Commiphora mukul* in experimental diabetes. *Int J Basic Clin Pharmacol*. 2019; 8: 575-83.
- Riyanti S, Suganda AG, Sukandar EY. Dipeptidyl peptidase-IV inhibitory activity of some Indonesian medicinal plants. *Asian Journal of Pharmaceutical and Clinical Research*. 2016; 9: 375-7.
- Zabidi NA, Ishak NA, Hamid M, Ashari SE, Mohammad MA. Inhibitory evaluation of *Curculigo latifolia* on α -glucosidase, DPP (IV) and in vitro studies in antidiabetic with molecular docking relevance to type 2 diabetes mellitus. *Journal of Enzyme Inhibition and Medicinal Chemistry*. 2021; 36: 109-21.

29. Ansari P, Choudhury ST, Abdel YH. Insulin secretory actions of ethanol extract of eucalyptus citriodora leaf, including plasma DPP-IV and GLP-1 levels in high-Fat-Fed rats, as well as characterization of biologically effective phytoconstituents. *Metabolites*. 2022; 12: 757.
30. Purnomo Y, Soeatmadji DW, Sumitro SB, Widodo MA. Anti-diabetic potential of Urena lobata leaf extract by inhibiting dipeptidyl peptidase IV activity. *Asian Pacific Journal of Tropical Biomedicine*. 2015; 5: 645-9.
31. Tangka J, Barung EN, Lyrawati D, Soeatmadji D, Nurdiana N. DPP-IV Inhibitory Activity of the Ethanolic Extract of Red Gedi Leaves *Abelmoschus manihot* L. *Medic. Macedonian Journal of Medical Sciences*. 2022; 10: 207-13.
32. Saidu Y, Muhammad SA, Bilbis LS, Sani BM. Inhibitory activity of fractions of *Senna nigricans* toward protein tyrosine phosphatase 1B and dipeptidyl peptidase IV. *Journal of Medicinal Plants Research*. 2016; 10: 242-7.
33. Yarzade A, Niazi AL. In vitro, antidiabetic effects of ferula asafoetida extract through dipeptidyl peptidase iv and α -glucosidase inhibitory activity. *In Vitro*. 2017; 10.
34. Srivastava S, Shree P, Tripathi YB. Active phytochemicals of *Pueraria tuberosa* for DPP-IV inhibition: in silico and experimental approach. *Journal of Diabetes & Metabolic Disorders*. 2017; 16: 1-9.
35. Kim BR, Kim HY, Choi I, Kim JB, Jin CH, Han ARJM. DPP-IV inhibitory potentials of flavonol glycosides isolated from the seeds of *Lens culinaris*: in vitro and molecular docking analyses. 2018; 23: 1998.
36. Prasannaraja C, Kamalanathan A, Vijayalakshmi M, Venkataraman K. Biotechnology, A dipyrrole derivative from *Aloe vera* inhibits an anti-diabetic drug target Dipeptidyl Peptidase (DPP)-IV in vitro. *Prep Biochem Biotechnol*. 2020; 50: 511-520.
37. Shrestha D, Sharma P, Adhikari A, Mandal AK, Verma A. A review on Nepalese medicinal plants used traditionally as alpha-amylase and alpha-glucosidase inhibitors against diabetes mellitus. *Current Traditional Medicine*. 2021; 7: 63-72.
38. Ansari P, Hannan J, Seidel V, Abdel-Wahab YHJM. Polyphenol-rich leaf of *Annona squamosa* stimulates insulin release from BRIN-BD11 cells and isolated mouse islets, reduces (CH₂O) n digestion and absorption, and improves glucose tolerance and GLP-1 (7-36) levels in high-fat-fed rats. 2022; 12: 995.
39. Hannan J, Ansari P, Azam S, Flatt PR, Wahab YHAA. Effects of *Spirulina platensis* on insulin secretion, dipeptidyl peptidase IV activity, and carbohydrate digestion and absorption indicate potential as an adjunctive therapy for diabetes. *Br J Nutr*. 2020; 124: 1021-1034.
40. Nidianti E, Rukman NK. An in silico study on Antidiabetic activity DPP-IV inhibitors and bioactive compounds *Boesenbergia pandurata* Roxb. *Surabaya International Health Conference*. 2019; 1: 121-126.
41. Elayaraja A, Muthupandi S, Radhakrishnan M, Rahaman SA. In vitro antioxidant and antibacterial activity of plant extracts of *Pergularia extensa* chiov. 2015; 7: 510-512.
42. Hajleh MNA, Khleifat KM, Alqaraleh M, Al-Hraishat E, Al-Limoun MO, Qaralleh H, et al. Antioxidant and Antihyperglycemic Effects of *Ephedra foeminea* Aqueous Extract in Streptozotocin-Induced Diabetic Rats. *Nutrients*. 2022; 14: 2338.
43. Thakur S, Chimwal J, Joshi R, Kumari M, Padwad Y, Kumar R. Evaluating Peptides of *Picrorhiza kurroa* and Their Inhibitory Potential against ACE, DPP-IV, and Oxidative Stress. *Journal of proteome research*. *J Proteome Res*. 2021; 20: 3798-3813.
44. Bharti SK, Krishnan S, Kumar A, Rajak KK, Murari K, Bharti BK, et al. Antihyperglycemic activity with DPP-IV inhibition of alkaloids from seed extract of *Castanospermum australe*: Investigation by experimental validation and molecular docking. *Phytomedicine: international journal of phytotherapy and phytopharmacology*; 2012; 20: 24-31.
45. Kim BR, Paudel SB, Nam JW, Jin CH, Lee IS, Han AR. Constituents of *Coreopsis lanceolata* flower and their dipeptidyl peptidase IV inhibitory effects. 2020; 25: 4370.
46. Iheagwam FN, Ogunlana O, Chinedu S. Model optimization and in silico analysis of potential dipeptidyl peptidase iv antagonists from GC-MS identified compounds in *Nauclea latifolia* leaf extracts. *Int J Mol Sci*. 2019; 20: 5913.
47. Suresh PS, Singh PP, Padwad YS, Sharma U. Pharmacology, Steroidal saponins from *Trillium govanianum* as α -amylase, α -glucosidase, and dipeptidyl peptidase IV inhibitory agents. 2021; 73: 487-495.
48. Aulifa DL, Adnyana IK, Sukrasno S, Levita J. Inhibitory activity of xanthoangelol isolated from *Ashitaba* (*Angelica keiskei* Koidzumi) towards α -glucosidase and dipeptidyl peptidase-IV: in silico and in vitro studies. *Heliyon*. 2022; 8: e09501.
49. Mojica L, De Mejía E. Function, Optimization of enzymatic production of anti-diabetic peptides from black bean (*Phaseolus vulgaris* L.) proteins, their characterization and biological potential. *Food Funct*. 2016; 7: 713-727.
50. Wu M, Yang Q, Wu Y, Ouyang J. Inhibitory effects of acorn (*Quercus variabilis* Blume) kernel-derived polyphenols on the activities of α -amylase, α -glucosidase, and dipeptidyl peptidase IV. *Food Bioscience*. 2021; 43: 101224.
51. Kumar V, Sachan R, Rahman M, Sharma K, Al-Abbasi FA, Anwar F. *Prunus amygdalus* extract exert antidiabetic effect via inhibition of DPP-IV: In-silico and in-vivo approaches. *Journal of Biomolecular Structure and Dynamics*. 2021; 39: 4160-4174.
52. Klein G, Kim J, Himmeldirk K, Cao Y, Chen X. Anti-diabetes and anti-obesity activity of *Lagerstroemia speciosa*. *Evidence-Based Complementary and Alternative Medicine*. 2007; 4: 401-407.
53. Rao PV, Naidu DM. *Rhinacanthus nasutus*: a plant with potential activity in radical scavenging capacity. *Current Trends in Biotechnology and Pharmacy*. 2010; 4: 791-794.
54. Kalhotra P, Chittepu VC, Osorio-Revilla G, Gallardo-Velazquez T. Phytochemicals in garlic extract inhibit therapeutic enzyme DPP-4 and induce skeletal muscle cell proliferation: A possible mechanism of action to benefit the treatment of diabetes mellitus. 2020; 10: 305.
55. Bhat SH, Ullah MF, Abu-Duhier FM. Bioactive extract of *Artemisia judaica* causes in vitro inhibition of dipeptidyl peptidase IV and pancreatic/intestinal enzymes of the carbohydrate absorption cascade: Implication for anti-diabetic new molecular entities (NMEs). *Oriental Pharmacy and Experimental Medicine*. 2019; 19: 71-80.
56. Mohanty IR, Borde M, Kumar CS, Maheshwari U. Dipeptidyl peptidase IV Inhibitory activity of *Terminalia arjuna* attributes to its cardioprotective effects in experimental diabetes: In silico, in vitro and in vivo analyses. *Phytomedicine: international journal of phytotherapy and phytopharmacology*. 2019; 57: 158-165.
57. Ansari P, Choudhury ST, Abdel-Wahab YHA. Insulin Secretory Actions of Ethanol Extract of *Eucalyptus citriodora* Leaf, including Plasma DPP-IV and GLP-1 Levels in High-Fat-Fed Rats, as well as Characterization of Biologically Effective Phytoconstituents. *Metabolites*. 2022; 12: 757.
58. Hannan JMA, Ansari P, Azam S, Flatt PR, Abdel Wahab YHA. Effects of *Spirulina platensis* on insulin secretion, dipeptidyl peptidase IV activity and both carbohydrate digestion and absorption indicate potential as an adjunctive therapy for diabetes. *The British journal of nutrition*. 2020; 124: 1021-1034.

59. Quek A, Kassim NK, Lim PC, Tan DC, Mohammad Latif MA, Ismail A, et al. α -Amylase and dipeptidyl peptidase-4 (DPP-4) inhibitory effects of *Melicope latifolia* bark extracts and identification of bioactive constituents using in vitro and in silico approaches. *Pharmaceutical biology*. 2021; 59: 964-973.
60. Tan J, Yang J, Zhou X, Hamdy AM, Zhang X, Suo H, et al. Tenebrio Molitor Proteins-Derived DPP-4 Inhibitory Peptides: Preparation, Identification, and Molecular Binding Mechanism. *Foods (Basel, Switzerland)*. 2022; 11: 3626.
61. Zhang M, Zhu L, Wu G, Liu T, Qi X, Zhang H. Rapid Screening of Novel Dipeptidyl Peptidase-4 Inhibitory Peptides from Pea (*Pisum sativum* L.) Protein Using Peptidomics and Molecular Docking. *Journal of agricultural and food chemistry*. 2022; 70: 10221-10228.
62. Pei J, Liu Z, Pan D, Zhao Y, Dang Y, Gao X. Transport, Stability, and In Vivo Hypoglycemic Effect of a Broccoli-Derived DPP-IV Inhibitory Peptide VPLVM. *Journal of agricultural and food chemistry*. 2022; 70: 4934-4941.
63. You H, Wu T, Wang W, Li Y, Liu X, Ding L. Preparation and identification of dipeptidyl peptidase IV inhibitory peptides from quinoa protein. *Food Research International*. 2022; 156: 111176.
64. Ansari P, Hannan JA, Abdel-Wahab YH, Flatt PR. Antidiabetic and insulinotropic properties of the bark of *Heritiera fomes*: inhibit starch digestion, protein glycation, DPP-IV activity, and glucose absorption in the gut. *Planta Med*. 2021; 87: 1252.
65. Mohanty IR, Kumar CS, Borde M. Antidiabetic activity of *Commiphora mukul* and *Phyllanthus emblica* and Computational analysis for the identification of active principles with dipeptidyl peptidase IV inhibitory activity. *Indian Journal of Pharmacology*. 2021; 53: 384-387.
66. Singh AK, Joshi J, Jatwa R. Dipeptidyl peptidase IV (DPP-IV/CD26) inhibitory and free radical scavenging potential of *W. somnifera* and *T. foenum-graecum* extract. *International Journal of Phyto-medicine*. 2013; 5: 503.
67. Harnedy PA, O'Keeffe MB, FitzGerald RJ. Purification and identification of dipeptidyl peptidase (DPP) IV inhibitory peptides from the macroalga *Palmaria palmata*. *Food chemistry*. 2015; 172: 400-6.
68. Bower AM, Real Hernandez LM, Berhow MA, de Mejia EG. Bioactive compounds from culinary herbs inhibit a molecular target for type 2 diabetes management, dipeptidyl peptidase IV. *Journal of agricultural and food chemistry*. 2014; 62: 6147-58.
69. Kato E, Uenishi Y, Inagaki Y, Kurokawa M, Kawabata J. Isolation of rugosin A, B and related compounds as dipeptidyl peptidase-IV inhibitors from rose bud extract powder. *Bioscience, biotechnology, and biochemistry*. 2016; 80: 2087-2092.
70. Beidokhti M, Lobbens E, Rasoavaivo P, Staerk D, Jäger AK. Investigation of medicinal plants from Madagascar against DPP-IV linked to type 2 diabetes. *South African Journal of Botany*. 2018; 115: 113-119.
71. Valcheva-Kuzmanova S, Kuzmanov K, Tancheva S, Belcheva A. Hypoglycemic and hypolipidemic effects of *Aronia melanocarpa* fruit juice in streptozotocin-induced diabetic rats. *Methods and findings in experimental and clinical Pharmacology*. 2007; 29: 101-5.
72. Yamane T, Kozuka M, Wada-Yoneta M, Sakamoto T, Nakagaki T, Nakano Y, et al. *Aronia* juice suppresses the elevation of postprandial blood glucose levels in adult healthy Japanese. 2017; 12: 20-26.
73. Calero JS, Young LC, Faz EM, Harvey AL. Inhibitory effect of *Allophylus cominia* (L.) Sw leaves aqueous extract on tyrosine phosphatase 1B and dipeptidyl peptidase IV proteins. *Revista Cubana de Farmacia*. 2014; 48: 672-683.
74. Amit R, Pushpa P. Assessment of the mechanism of action of antidiabetic activity of *Calocybe indica* by enzyme inhibitory activity. *Biosciences Biotechnology Research Asia*. 2016; 13: 2117-2123.
75. Saidu Y, Muhammad SA, Bilbis LS, Sani BM. Inhibitory activity of fractions of *Senna nigricans* toward protein tyrosine phosphatase 1B and dipeptidyl peptidase IV. *Journal of Medicinal Plants Research*. 2016; 10: 242-247.
76. Morikawa T, Ninomiya K, Akaki J, Kakiyama N, Kuramoto H, Matsumoto Y, et al. Dipeptidyl peptidase-IV inhibitory activity of dimeric dihydrochalcone glycosides from flowers of *Helichrysum arenarium*. *Journal of natural medicines*. 2015; 69: 494-506.
77. CunhaNeves A, Harnedy-Rothwell PA, FitzGerald RJ. Technology, In vitro angiotensin-converting enzyme and dipeptidyl peptidase-IV inhibitory, and antioxidant activity of blue mussel (*Mytilus edulis*) byssus collagen hydrolysates. *European Food Research and Technology*. 2022; 248: 1721-1732.
78. Tangka J, Barung EN, Lyrawati D, Soeatmadji D, Nurdiana N. DPP-IV Inhibitory Activity of the Ethanol Extract of Red Gedi Leaves *Abelmoschus manihot* L. *Medic*. 2022; 10: 207-213.
79. Guasch L, Ojeda MJ, González-Abuín N, Sala E, Cereto-Massagué A, Mulero M, et al. Identification of novel human dipeptidyl peptidase-IV inhibitors of natural origin (part I): virtual screening and activity assays. *PloS one*. 2012; 7: e44971.
80. Al-Masri IM, Mohammad MK, Tahaa MO. Inhibition of dipeptidyl peptidase IV (DPP IV) is one of the mechanisms explaining the hypoglycemic effect of berberine. *Journal of enzyme inhibition and medicinal chemistry*. 2009; 24: 1061-6.
81. Kim BR, Kim HY, Choi I, Kim JB, Jin CH, Han AR. DPP-IV Inhibitory Potentials of Flavonol Glycosides Isolated from the Seeds of *Lens culinaris*: In Vitro and Molecular Docking Analyses. *Molecules (Basel, Switzerland)*. 2018; 23: 1998.
82. Fan J, Johnson MH, Lila MA, Yousef G, de Mejia EG. Berry and Citrus Phenolic Compounds Inhibit Dipeptidyl Peptidase IV: Implications in Diabetes Management. *Evidence-based complementary and alternative medicine: eCAM*. 2013; 2013: 479505.
83. Gao Y, Zhang Y, Zhu J, Li B, Li Z, Zhu W, et al. Recent progress in natural products as DPP-4 inhibitors. *Future medicinal chemistry*. 2015; 7: 1079-89.
84. Gao Y, Zhang Y, Zhu J, Li B, Li Z, Zhu W, et al. Recent progress in natural products as DPP-4 inhibitors. *Future Medicinal Chemistry*. 2015; 7: 1079-89.
85. Lin YS, Chen CR, Wu WH, Wen CL, Chang CI, Hou WC. Anti- α -glucosidase and Anti-dipeptidyl Peptidase-IV Activities of Extracts and Purified Compounds from *Vitis thunbergii* var. *taiwaniana*. *Journal of agricultural and food chemistry*. 2015; 63: 6393-401.
86. <https://clinicaltrials.gov/>