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Review Article

Immunological Phenotypes of Premalignant Oral Lesions and the Immune Shifts with the Development of Head and Neck Cancer

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Abstract

Compared to other cancer types, squamous cell carcinomas of the head and neck (HNSCC) have been understudied.Studies in other cancer types have demonstrated immunological changes that occur with the development of cancer.However, few studies have looked at the immune changes that occur in premalignant lesions.HNSCC is often preceded by the appearance of premalignant oral lesions.However, information about the immune shifts that occur as a result of the premalignant oral lesions and following progression to HNSCC is sparse.This review aims to pull together studies with patients and animal models to summarize the multiplicities of immune activities that are associated with premalignant oral lesions and HSNCC.In general, the immune phenotype in premalignant oral lesions and regional lymph nodes is inflammatory and activated.However, development of HNSCC results in a shift to an immune inhibitory environment.

Keywords: Cytokines, Head and neck cancer, HNSCC, Immune, Inflammation, Oral cancer, Premalignant oral lesions

Overview of the Immune Influences on Premalignant Oral Lesions and Their Progression to Cancer

Despite advances in diagnosis and treatment, the 5-year survival rates for patients with head and neck squamous cell carcinoma (HNSCC) have not improved over 60% [1]. HNSCC is a highly aggressive disease that develops from the oral epithelium and its appearance is often preceded by premalignant oral lesions, most commonly leukoplakias [2]. Even with advanced treatments for premalignant oral lesions, about 30% progress to cancer [3]. Persistent infection with high-risk human Papillomaviruses (HPV) is associated with increased risk of oral cancer. Also associated with risk for HNSCC are alcohol consumption and tobacco.Each of these contributors to the development of epithelial dysplasia and progression to cancer are immune modulatory and complicate the immunological phenotype of oral tissues that are progressing to cancer [4-6].

The multifaceted aspects of the immune system have placed it into opposing roles in the development and progression of cancer (Table 1). On one hand, immune activation, and in particular inflammation, is considered to be among the contributors of cancer onset.Examples of this include associations between cancer and inflammation associated with obesity, inflammatory bowel disease and even oral irritation[7-11]. On the other hand, immune activation can also repress tumor development, as demonstrated through clinical immunotherapeutic trials using various iterations of cancer vaccines or other immune modulatory approaches.Immune protection against cancer is also demonstrated by the increased cancer development in immune compromised individuals such as the increased incidence of non-AIDS-defining cancers in patients with HIV infection [12-14]. These opposing capabilities of the immune system to promote or protect against cancer raise the question of its role in the development of oral lesions and progression to cancer. The answer to this question

is highly fragmented, although there are more fragments that can be assembled to understand the immune status in overt HNSCC, as compared to the absence of information pertaining to the impact of the immune milieu on premalignant oral lesions.

The Immunological Environment of Premalignant Oral Lesions

Discussion of the immunological environment of premalignant oral lesions needs to focus not only on the epithelium undergoing transformation, but the myriad of associated cells that comprise the premalignant oral lesion. This includes immune modulatory cells such as endothelial cells of the expanding vasculature, the immune infiltrate consisting of a spectrum of cells further described below, smooth muscle cells and fibroblasts[2,15-17]. A prominent immunological impact of the development of the premalignant oral lesion is seen not only within the premalignant lesions, but throughout the oral cavity, within regional lymph nodes and, in some instances, in the circulation (Table 2). For example, compared to levels in the peripheral blood of healthy control patients, patients with premalignant oral lesions have an increased proportion of NK cells and activated B-cells [18]. Levels of the inflammatory cytokines interleukin (IL)-17 and IL-6 are increased in the peripheral blood of patients with premalignant oral lesions [19]. Other circulating inflammatory mediators whose levels are increased in patients with premalignant lesions are transforming growth factor- β (TGF- β) and C-reactive protein [20]. Also, levels of the inflammatory mediators IL-6 and tumor necrosis factor- α (TNF- α) have been shown to be elevated in the saliva of patients with premalignant oral lesions, although a separate study showed that salivary levels of the inflammatory mediator IL-8 not to be elevated [21,22]. Such studies raise the possibility of using salivary levels of select cytokines as biomarkers for oral carcinogenesis.Such immune reactivities to premalignant oral lesions can be triggered by the increased expression of some of

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Table 1: Opposing roles of immune system in developed	ment and progression of
cancer.	

	Immune contribution to cancer	Immune protection against
	cancer	
	Inflammation	Inflammation
	Increased cancer in inflammation associated	Indication of failed immune
	with obesity, oral irritation	anti-tumor attempt?
	Immune inhibitory cell populations	Immunotherapy
	Suppress immune anti-tumor defenses	Show potential of immune anti- tumor reactivity

Table 2: Inflammatory and immune stimulatory phenotype associated with premalignant oral lesions.

Premalignant lesion patient sample	4NQO mouse	e premalignant lesion model
Lesion tissue & blood	Lesions and regional lymph nodes	
↑NK cells ↑CD8⁺ granzyme b⁺T-cells ↑dendritic cells	↑Tc1 and Th1 T-cells	
↑Inflammatory mediators	↑Inflammatory mediators	
	IL-6	IL-17
IL-6	IL-23	TGF-β
IL-17	TNF-α	C-reactive protein
TGF-β	CCL2	CCL3
TNF-α	CCL4	CCL5
	PGE ₂	
↑Immune stimulatory cytokines	↑Immune stimulatory cytokines	
IFN-γ	IFN-γ	
IL-2	IL-2	

the same tumor antigens on lesions as are prominent in HNSCC [23]. In fact, this tumor antigen expression on premalignant oral lesions can be used to stimulate immune reactivity by autologous blood lymphocytes. Among these tumor antigens are the mucinMUC-1 and epidermal growth factor receptor (EGFR).

Studies using a carcinogen-induced mouse premalignant oral lesion model that progresses to oral cancer have allowed more detailed assessments of the immunological changes that occur in the course of the development of lesions than would be allowed with patient specimens. Mice treated with the carcinogen 4-nitroquinoline 1-oxide (4NQO) in drinking water develop premalignant oral lesions predominantly on the tongue. These premalignant oral lesions progress to HNSCC.Studies with this model have shown prominent immune stimulation and increased cytokine production by the premalignant lesions as well as within regional lymph nodes [15,24]. Increased immune reactivity and, in particular, inflammation occur within both regional lymph nodes and premalignant oral lesion tissues of 4NQO-treated mice. The lymph nodes of these mice with premalignant oral lesions are enlarged, and the level and composition of the immune infiltrate within the lesions is increased. Also seen is an increase in conventional CD4⁺ and CD8⁺ T-cells that express markers for activation compared to what is seen in either healthy controls or HNSCC-bearing mice[15]. These mice with premalignant oral lesions have increased levels of Th1/Tc1 cells expressing interferon-y (IFN-γ) and IL-2, and CD4⁺Th17 cells producing the inflammatory cytokine IL-17.Heightened levels of cytokines and chemokines are also secreted by lymph node cells from 4NQO-treated mice with premalignant lesions. These include IFN-y, IL-2, and members of the chemokine (C-C motif) ligand family CCL 5 (RANTES), CCL3 (macrophage inflammatory protein-1, MIP-1 α) and CCL4 (MIP-1 β) [15].

While few studies have examined the immunological milieu

within the premalignant oral lesion tissues of patients, there have been reports indicating that, like that seen in the mouse models of premalignant oral lesions, inflammation and immune activation also occur in patients with lesions.Similar to the increases seen in levels of NK cells in the peripheral blood, NK cells are also increased within premalignant oral lesions [25]. An analysis of the immune infiltrate in leukoplakias with or without dysplasia showed an increased in numbers of CD8⁺ T-cells within the epithelium of lesions with dysplasia as compared to levels in lesions without dysplasia or in HNSCC [26]. Also increased in these leukoplakias with dysplasia is the number of dendritic Langerhans cells, with prominent localization with infiltrating T-cells [25,26]. There have been demonstrations of increased levels of Th1 cytokines such as IL-2 and IFN-y, as well as inflammatory cytokines IL-6 and IL-17within premalignant oral lesions [19]. A separate study comparing cytokine mRNA levels among premalignant oral lesions and healthy gingiva showed no statisticalincrease in TNF-a expression, but significant increases in expression of TGF- β and in select growth factors [27]. In contrast, $TNF-\alpha$ was shown to be expressed by immunohistochemistry throughout the epithelia of premalignant oral lesions, but only rarely in normal oral tissues [28]. Levels of CD8+ T-cells expressing perforin or granzyme B are also increased in premalignant oral lesions compared to control tissue [29]. Half of the premalignant oral lesions examined expressed the inflammatory and pro-angiogenic chemokine C-X-C motif chemokine 12 (CXCL12, stromal cell-derived factor-1) and over a third expressed its C-X-C receptor, CXCR4 [30]. This CXCL12/CXCR4 axis could contribute not only to the inflammatory cell influx, but can also promote he angiogenesis that is needed by the progressing lesion and subsequent cancer [31]. Expression levels of Toll-like receptors TLR4 and TLR9, which have an essential role in innate immunity activation, is increased in premalignant oral lesions compared to that in control normal mucosa, and the level of expression coincides with the magnitude of dysplasia [32].

Of interest is that shifts in cytokine phenotypes in the patients' oral lesions do not always coincide with what's seen in the peripheral blood. While IL-17 and IL-6 levels are increased in both premalignant lesions and blood, levels of the Th1 cytokines IL-2 and IFN- γ are increased only in the premalignant lesions, but not in the blood of patients with lesions [19]. However, the cytokine shifts in the regional lymph nodes coincide more closely with what is seen in the premalignant oral lesions.Thus, the cytokine phenotype within regional lymph nodes may be more representative of that within the premalignant oral lesions than the peripheral blood cytokine phenotype.Consistent with the immune activation seen in premalignant oral lesions, is a diminished level of the non-classicmajor histocompatibility class Ib antigen, which lessens its capacity to subvert immune responses [33]. The decline in this immune moderator could contribute to the increased immune reactivity seen in premalignant oral lesions.

The sources of the immune mediators that are produced within the premalignant lesions are multiple, including the lesion cells themselves.Immune mediators that are produced directly by the epithelium of premalignant lesions have been defined by establishing primary premalignant lesion cell cultures from tongue tissue of mice with carcinogen-induced lesions.These cultures produce high levels of pro-inflammatory and Th1-associated cytokines including granulocyte colony-stimulating factor (G-CSF), CCL5, and CCL2 (monocyte chemotactic protein 1, MCP-1), with the levels produced being greater than those produced by primary HNSCC cultures established from mice whose lesions progressed to HNSCC [15,24]. Consistent with the increased production of inflammatory mediators was an increased production of prostaglandin E_2 (PGE₂) by premalignant lesion cells as compared to levels produced by HNSCC.IL-23, which is critical to the maintenance of IL-17 cells, is also produced in increased levels by premalignant oral lesion cells, suggesting this to be a route by which lesion cells may be stimulating the Th17 cell content [19]. These studies suggest that Th1/Tc1 and inflammatory immune reactivities are stimulated early prior to the appearance of HNSCC, during the premalignant lesion stages, and are more prominent than in HNSCC.

How Can A Small Premalignant Oral Lesion Cause Distant Immune Alterations?

One might ask- how can premalignant oral lesions that may be only a few millimeters in diameter cause significant immune alterations at distant sites?While premalignant lesion cells produce mediators that can alter immune reactivity, it is likely that much of the immunological skewing associated with premalignant oral lesions is the result of amplifications in cytokine cascades.For example, mediators from premalignant lesion cells can stimulate normal resting spleen cells to produce increased levels of Th1 and inflammatory cytokines, thus triggering greater reactivity than would be possible by the lesion cells alone [24]. This capacity is accentuated by the lesion production of chemokines that, in turn, stimulate an influx of immune cells that are then also exposed to the cytokinetriggering mediators produced by premalignant lesions [24,30]. As indicated above, these infiltrating cells include, but are not limited to, T-cells expressing markers of activations, skewed macrophages and neutrophils that are also recruited by chemokines, and activated endothelial cells associated with the premalignant lesions [15,17].

What is not often appreciated is the role of endothelial cells as immune regulatory cells. The development of premalignant oral lesions and their progression to cancer requires a healthy and increasing vasculature. The pro-inflammatory potential of endothelial cells, especially following stimulation, has been reasonably well described [34-36]. Because of their location, vascular endothelial cells have an intimate relationship with immune cells, which facilitates their recruitment of the immune infiltrate. Endothelial cells can secrete numerous immune suppressive as well as inflammatory products including vascular endothelial cell growth factor (VEGF), PGE, TGF-B, IL-6 and IL-10 [37,38]. In addition, endothelial cell-derived mediators can stimulate spleen cell production of pro-inflammatory cytokines such as IL-6, IL-9, IL-17, and TNF- $\!\alpha\!;$ production of the predominantly T-cell derived chemokine CCL5; and production of several predominantly monocyte-derived chemokines to include CXCL9 (monokine induced by y-interferon, MIG) and CCL2 [39].

While premalignant oral lesions may be small in relative size, their immunological impact can be highly prominent. This prominent impact is orchestrated by the dysplastic epithelium, and accentuated by soldiers in the cascade including the endothelium and infiltrating immune cells. Whether this escalation of immune reactivity is an attempt to mount an immune response to the developing malignancy or whether it contributed to the progression toward cancer is not certain, although these two possibilities are not necessarily mutually exclusive.What is certain is that if it is an attempt to eliminate the dysplastic epithelial cells, the attempt often times fails with the resulting emergence of HNSCC.

The Immunological Environment of HNSCC

While the immune phenotype of premalignant oral lesions is stimulated and inflammatory in nature, the immune phenotype of HNSCC is different as immune inhibitory mediators and suppressor cells become more prominent (Table 3). As for premalignant oral lesions, levels of CD4⁺ and CD8⁺ cells in patient HNSCC tissue specimens are increased compared to that seen in normal healthy oral tissue [40,41]. Also increased are levels of dendritic cells in HNSCC, with the dendritic cells co-localizing with the T-cell infiltrate [26]. There are, however, some region-specific differences.For example, levels of CD8⁺ cells expressing perforin and granzyme B are increased in squamous carcinoma of the lip versus oral cavity [29]. Increases in the immune infiltrate and cytokines are also seen in HNSCC tissue and lymph nodes from the 4NQO HNSCC model compared to that seen in healthy control epithelium [15,19].

The increases in immune infiltration may give the perception of anti-tumor immune reactivity, but a prevailing inflammatory and immune inhibitory environment merges with the appearance of HNSCC.Inflammatory mediators whose levels are increased in the peripheral blood of HNSCC patients include IL-6, TGF- β and C-reactive protein, and increased levels of these mediators are associated with increased cancer aggressiveness and recurrence [20]. Some of these mediators, such as TGF- β and IL-6, have dual roles of being able to be both inflammatory and immune inhibitory. Levels of the pro-inflammatory mediator IL-8 have also been shown to be increased in the saliva of patients with HNSCC, although no association was seen between the levels of IL-8 and the cancer TNM stage [22].

The inhibitory immune environment within HNSCC as well as systemically in the HNSCC bearer is viewed as a major mechanism of immune evasion by the HNSCC. By secreting cytokines such as TGF- β and IL-10, HNSCC tumor cells promote a Th2-skewed response, which is associated with decreased antitumor efficacy [42,43]. This Th2-skewing is evident systemically as peripheral blood

Table 3: Shift to inhibitory phenotype in HNSC	C.
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HNSCC patient sample	4NQO mouse HNSCC model
Lesion tissue & blood	Lesions and regional lymph nodes
↑Th2/Tc2 ↑MDSC and CD34⁺ cells ↑Treg ↑Th2-skewed dendritic cells	1Th2/Tc2 1MDSC and CD34* cells 1Treg 1Th2-skewed dendritic cells 1M2 macrophages
↑Inflammatory mediators	↑Inflammatory mediators
IL-1 IL-6 PGE ₂ TNF-α TGF-β	IL-6 PGE2 TNF-α TGF-β
↑Immune inhibitory mediators	↑Immune inhibitory cytokines
IL-6 IL-10 PGE ₂ TGF-β	IL-6 IL-10 PGE ₂ TGF-β

leukocytes isolated from HNSCC patients secrete abnormally high levels of Th2 cytokines [44,45]. Th2 cytokines include IL-10, whose levels are increased in HNSCC patients [46]. These studies also showed that select polymorphisms of plasma IL-10 are associated with more advanced disease.

Along with Th2-skewed cytokines, HNSCC tumor secretion of factors such as TGF- β function to directly inhibit cytotoxic T cellmediated immunity and to recruit to the tumor site additional immunosuppressive cells, including myeloid-derived suppressor cells (MDSCs),the less mature CD34⁺ progenitor cells, as well as M2-skewed macrophages [47,48]. Tumor-secreted granulocytemacrophage colony-stimulating factor (GM-CSF) promotes MDSC and CD34⁺ progenitor cells, and high levels of GM-CSF in HNSCC patients are associated with a poorer prognosis [49,50]. Once at the tumor site, these immunosuppressive cells facilitate tumor-promoting functions, resulting in increased tumor growth and angiogenesis.

HNSCC tumor secretion of factors that are typically associated with a pro-inflammatory response harnesses these immune modulators to favor growth, angiogenesis and, paradoxically, immune escape. GM-CSF, TNF-a, IL-1 and PGE,, which have traditional pro-inflammatory roles, are significantly increased in HNSCC tissues from the 4NQO carcinogenmouse model [51]. Many of these mediators are also produced by HNSCC cell lines established from patients with head and neck cancer [52]. Increased levels of PGE, are associated with invasion and angiogenesis in aggressive early-stage tumors [53]. Studies in other models have shown that PGE, also has inhibitory effects on granulocyte function and macrophage phagocytosis, IL-2 production by T cells, T-cell responsiveness to IL-2, and antigen-specific T cell responses [54-56]. Dendritic cells have also been shown to be impacted by PGE, as it inhibits their differentiation and alters the activity of Th1associated dendritic cells to result in decreased IL-12 production and their increased promotion of Th2-skewed mature T-cells [57-59]. Blocking PGE, signaling reduces accumulation of MDSC and delays tumor growth [60]. A retrospective review of HNSCC patients taking cyclooxygenase inhibitors showed an increased survival over those not taking cyclooxygenase inhibitors following cancer treatment [61].

Other factors secreted by HNSCC tumors, such as the chemokine CCL2 (MCP-1), have been shown to contribute to immunosuppression at the tumor site by recruiting a population of M2-skewed tumor-associated macrophages secreting IL-10 and TGF- β [62]. The cytokine macrophage inhibitory factor could also contribute to the accumulation of these macrophages as levels of this mediator are increased within the tumors as well as in the inflammatory infiltrate [63]. By secreting a host of immune modulators, HNSCC tumors thwart an effective immune response and become increasingly difficult to treat.Combinations of cytokines, such as PGE₂ together with IL-6, can heighten the induction of immune inhibitory populations including Th2-skewed T-cells, M2-skewed macrophages, Treg and MDSC [64-66].

The developing HNSCC has prominent impacts on immune reactivity distal from the HNSCC site, with the phenotypes of both the distant and intratumoral reactivities being associated with patient outcomes [67-69]. Most studies involving the impact of HNSCC on patient immune status have assessed immune capabilities and phenotypes of patient peripheral blood.Peripheral blood lymphocytes have a reduced reactive capability to either proliferate or to release cytokines that evoke anti-tumor reactivity [70,71]. Circulating Treg cells from patients with advanced HNSCC were inhibitorier to the proliferation of effector T-cells than Treg from healthy controls or from patients without nodal involvement [72]. The activation status of peripheral blood CD8⁺T-cells was reduced with increased cancer aggressiveness [73]. Thus, it is clear that the immune modulations in HNSCC patients can impact on clinical outcome [74,75].

Transition from Premalignant Lesions to HNSCC

While there have only been a few studies examining the immunological status of patients with premalignant oral lesions as well as those with HNSCC, they have shown a shift from an immune stimulatory or inflammatory environment to an immune suppressive phenotype.In most instances, however, these studies have not necessarily used the same immunological measurements and have certainly not tracked the same patients with premalignant oral lesions that subsequently develop HNSCC.Such analyses have more reliably been conducted in murine models where premalignant oral lesions progress to oral cancer.Nevertheless, the culmination of these studies has shown increased cellularity within regional lymph nodes, but differences in cellular and cytokines phenotypes, with a greater number of Treg, MDSC, Th2 cells, and M2 macrophages within HNSCC as compared to that seen in cervical lymph nodes or blood of those with premalignant lesions [15,19,24]. Contributing to the immune infiltrate within premalignant lesions and HNSCC is the increased expression of the chemokine CXCL12 in lesions and more so in HNSCC, which is capable of recruiting inflammatory and immune inhibitory cells such as macrophages, which can be skewed to become immune inhibitory M2 macrophages in the tumor environment [31,76]. The increased expression of select TLRs as premalignant lesions become more dysplastic and progress to HNSCC has led to suggestions that their role in activation of innate immunity could contribute to tumorigenesis [32]. An increased level of Th17 cells and Th1/Tc1 cells in mice with premalignant oral lesions compared to those with HNSCC suggests robust immune reactivity being attempted against the lesions, but which doesn't persist once HNSCCdevelops [15,19]. The simultaneous increase in expression of not only markers for activation, but also for exhaustion in mice with premalignant oral lesions could be an early indicator of a failing response against the lesions since they indicate that T-cells have been activated, but they are at the point their lifespan where, upon further stimulation, they will undergo programmed cell death rather than perform effector functions [15,77,78]. This suggests that the exhaustion is a reflection of a failed attempt to immunologically prevent the progression of premalignant oral lesions to cancer. This attempted, but failing, response is far less evident in HNSCC and, as HNSCC progresses, is instead replaced by pronounced levels of a broad myriad of immune inhibitory cells and their inhibitory cytokines [15,19,24]. Also, the levels and select polymorphisms of the Th2 cytokine IL-10 in patients with premalignant lesions were associated with increased risk of progression to HNSCC [46].

What then triggers the shift from robust immune reactivity in the premalignant lesion environment to the immune suppressive

environment in HNSCC?Possibilities include the contribution of the inflammatory state that is so prominent in the premalignant oral lesion stage. Among the inflammatory mediators are PGE, and IL--6. These mediators can be produced by a multitude of cell types in the lesion and developing HNSCC environment, including epithelial cells, fibroblasts, and infiltrating immune cells and cancer cells [79]. Furthermore, the role of PGE, seems to shift during tumor progression whereas in the early stages of inflammation, PGE, promotes the infiltration of neutrophils, macrophages and mast cells [80-82]. However, PGE, and IL-6 can also skew the immune infiltrate toward an immune inhibitory phenotype consisting of Th2 T-cells, M2 macrophages, Treg and MDSC, thereby contributing to the shift from an immune stimulatory to inhibitory environment [64-66,83,84]. Among the mediators whose levels are also increased in premalignant lesions and regional lymph nodes is TGF- β [19] Studies with a mouse skin carcinogenesis model showed that inhibition of TGF-ß signaling lessened the level of inflammation that is associated with carcinogenesis, but increased premalignant progression to squamous cell carcinoma, raising the question of the pro-tumorigenic or anti-tumorigenic roles of inflammation [85].

TGF- β and IL-6 levels could be contributors to the increase levels of Th17 cells in premalignant oral lesions, whose phenotype is stabilized by IL-23, which is also in increased levels in premalignant lesions. However, IL-23 levels decline and TGF- β levels further increase in the HNSCC environment[19]. Since changing the balance to increased TGF- β levels while also reducing IL-23 levels favors skewing of T-cells toward suppressive Th2 cells, shifts in the levels of these mediators could be a cause of the decline in the Th17 cells that are seen in premalignant lesions to an increase in Th2 cells seen in HSNCC. The role of Th17 cells in tumor progression or protection from tumor is also controversial [86-88]. Th17 cell presence has been associated with improved prognosis in early stage ovarian cancer and malignant pleural effusions, as they promote a Th1 cytokine environment through the combined effect of IL-17 plus IFN-y stimulation of CXCL9 and CXCL10 (interferon y-induced protein 10, IP-10) to recruit effector T-cells [87,89]. Th17 cells have also been shown to have direct anti-proliferative and apoptosis-inducing effects toward HNSCC [86]. In other cancer scenarios, the presence of Th17 cells has been associated with increased cancer development [90]. These studies with Th17 cells and their plasticity to become Th2 cells suggest it may not be the cytokines per se that direct an inflammatory versus inhibitory environment, but the ratios of the cytokines may be equally important.

Overall, the diversity of the immune responses that are triggered by the premalignant oral lesion environment and the HNSCC environment is broad, reflecting attempts to eradicate the dysplastic cells, the failure of the immune eradiation, and the capture of immune reactivities by the emerging tumor to be subversive to anti-tumor immune defenses.Difficulties in deciphering, the aspects of the immunological components that are anti-tumorigenic versus pro-tumorigenic are compounded by the plasticity of immune cells and their phenotypic and functional shifts.A more thorough understanding of the roles of the immunological statesin the premalignant lesion versus HNSCC environments will allow targeted modulation of immune responsesto sustain the immunological attempt to eradicate the developing tumor.

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References

- 1. National Cancer Institute. Surveillance Epidemiology and End Results. 2011.
- Neville BW, Day TA. Oral cancer and precancerous lesions. CA Cancer J Clin. 2002; 52: 195-215.
- Sudbø J, Lippman SM, Lee JJ, Mao L, Kildal W, Sudbø A, et al. The influence of resection and aneuploidy on mortality in oral leukoplakia. N Engl J Med. 2004; 350: 1405-1413.
- Feller L, Wood NH, Khammissa RA, Chikte UM, Meyerov R, Lemmer J, et al. HPV modulation of host immune responses. SADJ. 2010; 65: 266-268.
- Arimilli S, Damratoski BE, Prasad GL. Combustible and non-combustible tobacco product preparations differentially regulate human peripheral blood mononuclear cell functions. Toxicol In Vitro. 2013; 27: 1992-2004.
- Asquith M, Pasala S, Engelmann F, Haberthur K, Meyer C, Park B, et al. Chronic ethanol consumption modulates growth factor release, mucosal cytokine production, and microRNA expression in nonhuman primates. Alcohol Clin Exp Res. 2014; 38: 980-993.
- Vielma SA, Klein RL, Levingston CA, Young MR. Premalignant lesions skew spleen cell responses to immune modulation by adipocytes. Anticancer Res. 2013; 33: 1809-1818.
- Achyut BR, Bader DA, Robles AI, Wangsa D, Harris CC, Ried T, et al. Inflammation-mediated genetic and epigenetic alterations drive cancer development in the neighboring epithelium upon stromal abrogation of TGF-b signaling. PLoS Genet. 2013; 9: e1003251.
- Meeker S, Seamons A, Paik J, Treuting PM, Brabb T, Grady WM, et al. Increased dietary vitamin d suppresses MAPK signaling, colitis, and colon cancer. Cancer Res. 2014; 74: 4398-4408.
- Beigel F, Steinborn A, Schnitzler F, Tillack C, Breiteneicher S, John JM, et al. Risk of malignancies in patients with inflammatory bowel disease treated with thiopurines or anti-TNF alpha antibodies. Pharmacoepidemiol Drug Saf. 2014; 23: 735-744.
- Fronie A, Dumitrescu D, VascÄf V, Fronie AS. Histopathological aspects of benign epithelial tumors located in areas of friction or chronic irritation of the tongue. Rom J Morphol Embryol. 2011; 52: 831-835.
- Marshall NA, Galvin KC, Corcoran AM, Boon L, Higgs R, Mills KH. Immunotherapy with PI3K inhibitor and toll-like receptor agonist induces IFN- g+IL-17+ polyfunctional T cells that mediate rejection of murine tumors. Cancer Res. 2012; 72: 581-591.
- Alfaro C, Perez-Gracia JL, Suarez N, Rodriguez J, Fernandez dS, Sangro B, et al. Pilot clinical trial of type 1 dendritic cells loaded with autologous tumor lysates combined with GM-CSF, pegylated IFN, and cyclophosphamide for metastatic cancer patients. J Immunol. 2011; 187: 6130-6142.
- Patel P, Hanson DL, Sullivan PS, Novak RM, Moorman AC, Tong TC, et al. Incidence of types of cancer among HIV-infected persons compared with the general population in the United States, 1992-2003. Ann Intern Med. 2008; 148: 728-736.
- De Costa AM, Schuyler CA, Walker DD, Young MR. Characterization of the evolution of immune phenotype during the development and progression of squamous cell carcinoma of the head and neck. Cancer Immunol Immunother. 2012; 61: 927-939.
- 16. Kapse SC, Rathod N, Baad R, Mandlik J, Sharma AS, Bommanavar S, et al. Quantitative assessment of myofibroblast in severe dysplasia, microinvasion and oral squamous cell carcinoma: an immunohistochemical study. J Contemp Dent Pract. 2013; 14: 34-38.
- 17. Lee LT, Chen PH, Chang CT, Wang J, Wong YK, Wang HW. Quantitative

physiology and immunohistochemistry of oral lesions. Biomed Opt Express. 2013; 4: 2696-2709.

- Lee JJ, Lin CL, Chen TH, Kok SH, Chang MC, Jeng JH, et al. Changes in peripheral blood lymphocyte phenotypes distribution in patients with oral cancer/oral leukoplakia in Taiwan. Int J Oral Maxillofac Surg. 2010; 39: 806-814.
- Woodford D, Johnson SD, De Costa A-MA, Young MRI. An inflammatory cytokine milieu is prominent in premalignant oral lesions, but subsides when lesions progress to squamous cell carcinoma. J Clin Cell Immunol. 2014 [in press].
- Chang PY, Kuo YB, Wu TL, Liao CT, Sun YC, Yen TC. Association and prognostic value of serum inflammation markers in patients with leukoplakia and oral cavity cancer. Clin Chem Lab Med. 2013; 51: 1291-1300.
- Juretić M, Cerović R, Belušić-Gobić M, Brekalo Pršo I, Kqiku L, Špalj S, et al. Salivary levels of TNF-α and IL-6 in patients with oral premalignant and malignant lesions. Folia Biol (Praha). 2013; 59: 99-102.
- Punyani SR, Sathawane RS. Salivary level of interleukin-8 in oral precancer and oral squamous cell carcinoma. Clin Oral Investig. 2013; 17: 517-524.
- Young MR, Neville BW, Chi AC, Lathers DM, Boyd Gillespie M, Day TA, et al. Oral premalignant lesions induce immune reactivity to both premalignant oral lesions and head and neck squamous cell carcinoma. Cancer Immunol Immunother. 2007; 56: 1077-1086.
- Johnson SD, De Costa AM, Young MR. Effect of the premalignant and tumor microenvironment on immune cell cytokine production in head and neck cancer. Cancers (Basel). 2014; 6: 756-770.
- Bondad-Palmario GG. Histological and immunochemical studies of oral leukoplakia: phenotype and distribution of immunocompetent cells. J Philipp Dent Assoc. 1995; 47: 3-18.
- Öhman J, Magnusson B, Telemo E, Jontell M, Hasséus B. Langerhans cells and T cells sense cell dysplasia in oral leukoplakias and oral squamous cell carcinomas--evidence for immunosurveillance. Scand J Immunol. 2012; 76: 39-48.
- Wenghoefer M, Pantelis A, Najafi T, Deschner J, Allam JP, Novak N, et al. Gene expression of oncogenes, antimicrobial peptides, and cytokines in the development of oral leukoplakia. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2010; 110: 351-356.
- Younes F, Quartey EL, Kiguwa S, Partridge M. Expression of TNF and the 55-kDa TNF receptor in epidermis, oral mucosa, lichen planus and squamous cell carcinoma. Oral Dis. 1996; 2: 25-31.
- Costa NL, Gonçalves AS, Souza-Lima NC, Jaime-Paiva LG, Junqueira-Kipnis AP, Silva TA, et al. Distinct expression of perforin and granzyme B in lip and oral cavity squamous cell carcinoma. J Oral Pathol Med. 2011; 40: 380-384.
- Xia J, Chen N, Hong Y, Chen X, Tao X, Cheng B, et al. Expressions of CXCL12/CXCR4 in oral premalignant and malignant lesions. Mediators Inflamm. 2012; 2012: 516395.
- 31. Li B, Bai W, Sun P, Zhou B, Hu B, Ying J. The Effect of CXCL12 on Endothelial Progenitor Cells: Potential Target for Angiogenesis in Intracerebral Hemorrhage. J Interferon Cytokine Res. 2014.
- 32. Kotrashetti VS, Nayak R, Bhat K, Hosmani J, Somannavar P. Immunohistochemical expression of TLR4 and TLR9 in various grades of oral epithelial dysplasia and squamous cell carcinoma, and their roles in tumor progression: a pilot study. Biotech Histochem. 2013; 88: 311-322.
- Fregonezi PA, Silva TG, Simões RT, Moreau P, Carosella ED, Kläy CP, et al. Expression of nonclassical molecule human leukocyte antigen-G in oral lesions. Am J Otolaryngol. 2012; 33: 193-198.
- Wang W, Deng M, Liu X, Ai W, Tang Q, Hu J, et al. TLR4 activation induces nontolerant inflammatory response in endothelial cells. Inflammation. 2011; 34: 509-518.
- 35. Johnson LA, Jackson DG. Inflammation-induced secretion of CCL21 in lymphatic endothelium is a key regulator of integrin-mediated dendritic cell transmigration. Int Immunol. 2010; 22: 839-849.

- Wilhelmsen K, Mesa KR, Prakash A, Xu F, Hellman J. Activation of endothelial TLR2 by bacterial lipoprotein upregulates proteins specific for the neutrophil response. Innate Immun. 2012; 18: 602-616.
- Dalrymple NA, Mackow ER. Endothelial cells elicit immune-enhancing responses to dengue virus infection. J Virol. 2012; 86: 6408-6415.
- Mulligan JK, Young MR. Tumors induce the formation of suppressor endothelial cells in vivo. Cancer Immunol Immunother. 2010; 59: 267-277.
- Vielma SA, Klein RL, Levingston CA, Young MR. Skewing of immune cell cytokine production by mediators from adipocytes and endothelial cells. Adipocyte. 2014; 3: 126-131.
- Gannot G, Gannot I, Vered H, Buchner A, Keisari Y. Increase in immune cell infiltration with progression of oral epithelium from hyperkeratosis to dysplasia and carcinoma. Br J Cancer. 2002; 86: 1444-1448.
- Zancope E, Costa NL, Junqueira-Kipnis AP, Valadares MC, Silva TA, Leles CR, et al. Differential infiltration of CD8+ and NK cells in lip and oral cavity squamous cell carcinoma. J Oral Pathol Med. 2010; 39: 162-167.
- 42. Jewett A, Head C, Cacalano NA. Emerging mechanisms of immunosuppression in oral cancers. J Dent Res. 2006; 85: 1061-1073.
- Young MR. Protective mechanisms of head and neck squamous cell carcinomas from immune assault. Head Neck. 2006; 28: 462-470.
- 44. Bose A, Chakraborty T, Chakraborty K, Pal S, Baral R. Dysregulation in immune functions is reflected in tumor cell cytotoxicity by peripheral blood mononuclear cells from head and neck squamous cell carcinoma patients. Cancer Immun. 2008; 8: 10.
- Lathers DM, Achille NJ, Young MR. Incomplete Th2 skewing of cytokines in plasma of patients with squamous cell carcinoma of the head and neck. Hum Immunol. 2003; 64: 1160-1166.
- 46. Zhou J, Zhang D, Chen B, Li Q, Zhou L, Liu F, et al. Association of interleukin-10 promoter polymorphisms and corresponding plasma levels with susceptibility to laryngeal squamous cell carcinoma. Oncol Lett. 2014; 7: 1721-1727.
- 47. Lu SL, Reh D, Li AG, Woods J, Corless CL, Kulesz-Martin M, et al. Overexpression of transforming growth factor b1 in head and neck epithelia results in inflammation, angiogenesis, and epithelial hyperproliferation. Cancer Res. 2004; 64: 4405-4410.
- Wahl SM, Wen J, Moutsopoulos N. TGF-beta: a mobile purveyor of immune privilege. Immunol Rev. 2006; 213: 213-227.
- Bronte V, Chappell DB, Apolloni E, Cabrelle A, Wang M, Hwu P, et al. Unopposed production of granulocyte-macrophage colony-stimulating factor by tumors inhibits CD8+ T cell responses by dysregulating antigen-presenting cell maturation. J Immunol. 1999; 162: 5728-5737.
- 50. Young MR, Wright MA, Lozano Y, Prechel MM, Benefield J, Leonetti JP, et al. Increased recurrence and metastasis in patients whose primary head and neck squamous cell carcinomas secreted granulocyte-macrophage colony-stimulating factor and contained CD34+ natural suppressor cells. Int J Cancer. 1997; 74: 69-74.
- Gannot G, Buchner A, Keisari Y. Interaction between the immune system and tongue squamous cell carcinoma induced by 4-nitroquinoline N-oxide in mice. Oral Oncol. 2004; 40: 287-297.
- 52. Chen Z, Malhotra PS, Thomas GR, Ondrey FG, Duffey DC, Smith CW, et al. Expression of proinflammatory and proangiogenic cytokines in patients with head and neck cancer. Clin Cancer Res. 1999; 5: 1369-1379.
- Hambek M, Baghi M, Wagenblast J, Schmitt J, Baumann H, Knecht R, et al. Inverse correlation between serum PGE2 and T classification in head and neck cancer. Head Neck. 2007; 29: 244-248.
- Srivastava MK, Andersson Å, Zhu L, Harris-White M, Lee JM, Dubinett S. Myeloid suppressor cells and immune modulation in lung cancer. Immunotherapy. 2012; 4: 291-304.
- 55. Bao YS, Zhang P, Xie RJ, Wang M, Wang ZY, Zhou Z, et al. The regulation of CD4+ T cell immune responses toward Th2 cell development by prostaglandin E2. Int Immunopharmacol. 2011; 11: 1599-1605.

- 56. Woolard MD, Wilson JE, Hensley LL, Jania LA, Kawula TH, Drake JR, et al. Francisella tularensis-infected macrophages release prostaglandin E2 that blocks T cell proliferation and promotes a Th2-like response. J Immunol. 2007; 178: 2065-2074.
- Bekeredjian-Ding I, Schäfer M, Hartmann E, Pries R, Parcina M, Schneider P, et al. Tumour-derived prostaglandin E and transforming growth factorbeta synergize to inhibit plasmacytoid dendritic cell-derived interferon-alpha. Immunology. 2009; 128: 439-450.
- Pockaj BA, Basu GD, Pathangey LB, Gray RJ, Hernandez JL, Gendler SJ, et al. Reduced T-cell and dendritic cell function is related to cyclooxygenase-2 overexpression and prostaglandin E2 secretion in patients with breast cancer. Ann Surg Oncol. 2004; 11: 328-339.
- KaliÅ_sski P, Hilkens CM, Wierenga EA, Kapsenberg ML. T-cell priming by type-1 and type-2 polarized dendritic cells: the concept of a third signal. Immunol Today. 1999; 20: 561-567.
- Sinha P, Clements VK, Fulton AM, Ostrand-Rosenberg S. Prostaglandin E2 promotes tumor progression by inducing myeloid-derived suppressor cells. Cancer Res. 2007; 67: 4507-4513.
- Gillespie MB, Moody MW, Lee FS, Poole LJ, Hornig JD, Lathers D, et al. Head and neck cancer recurrence and mortality in nonselective cyclooxygenase inhibitor users. Arch Otolaryngol Head Neck Surg. 2007; 133: 28-31.
- Liss C, Fekete MJ, Hasina R, Lam CD, Lingen MW. Paracrine angiogenic loop between head-and-neck squamous-cell carcinomas and macrophages. Int J Cancer. 2001; 93: 781-785.
- França CM, Batista AC, Borra RC, Ventiades-Flores JA, Mendonça EF, Deana AM, et al. Macrophage migration inhibitory factor and oral cancer. J Oral Pathol Med. 2013; 42: 368-373.
- 64. Baratelli F, Lin Y, Zhu L, Yang SC, Heuzé-Vourc'h N, Zeng G, et al. Prostaglandin E2 induces FOXP3 gene expression and T regulatory cell function in human CD4+ T cells. J Immunol. 2005; 175: 1483-1490.
- Diehl S, Chow CW, Weiss L, Palmetshofer A, Twardzik T, Rounds L. Induction of NFATc2 expression by interleukin 6 promotes T helper type 2 differentiation. J Exp Med. 2002; 196: 39-49.
- Dodge IL, Carr MW, Cernadas M, Brenner MB. IL-6 production by pulmonary dendritic cells impedes Th1 immune responses. J Immunol. 2003; 170: 4457-4464.
- 67. Pretscher D, Distel LV, Grabenbauer GG, Wittlinger M, Buettner M, Niedobitek G. Distribution of immune cells in head and neck cancer: CD8+ T-cells and CD20+ B-cells in metastatic lymph nodes are associated with favourable outcome in patients with oro- and hypopharyngeal carcinoma. BMC Cancer. 2009; 9: 292.
- 68. O'Donnell RK, Mick R, Feldman M, Hino S, Wang Y, Brose MS, et al. Distribution of dendritic cell subtypes in primary oral squamous cell carcinoma is inconsistent with a functional response. Cancer Lett. 2007; 255: 145-152.
- 69. Molling JW, Langius JA, Langendijk JA, Leemans CR, Bontkes HJ, van der Vliet HJ, et al. Low levels of circulating invariant natural killer T cells predict poor clinical outcome in patients with head and neck squamous cell carcinoma. J Clin Oncol. 2007; 25: 862-868.
- Wild CA, Brandau S, Lindemann M, Lotfi R, Hoffmann TK, Lang S, et al. Tolllike Receptors in Regulatory T Cells of Patients with Head and Neck Cancer. Arch Otolaryngol Head Neck Surg. 2010; 136: 1253-1259.
- Malm IJ, Bruno TC, Fu J, Zeng Q, Taube JM, Westra W, et al. Expression profile and in vitro blockade of PD-1 in HPV-negative head and neck squamous cell carcinoma. Head Neck. 2014; .
- 72. Drennan S, Stafford ND, Greenman J, Green VL. Increased frequency and suppressive activity of CD127(low/-) regulatory T cells in the peripheral circulation of patients with head and neck squamous cell carcinoma are associated with advanced stage and nodal involvement. Immunology. 2013; 140: 335-343.

- 73. Andrade MC, Ferreira SB, Gonçalves LC, De-Paula AM, de Faria ES, Teixeira-Carvalho A, et al. Cell surface markers for T and B lymphocytes activation and adhesion as putative prognostic biomarkers for head and neck squamous cell carcinoma. Hum Immunol. 2013; 74: 1563-1574.
- Millrud CR, Månsson Kvarnhammar A, Uddman R, Björnsson S, Riesbeck K, Cardell LO. The activation pattern of blood leukocytes in head and neck squamous cell carcinoma is correlated to survival. PLoS One. 2012; 7: e51120.
- Heimdal JH, Aarstad HJ, Klementsen B, Olofsson J. Peripheral blood mononuclear cell (PBMC) responsiveness in patients with head and neck cancer in relation to tumour stage and prognosis. Acta Otolaryngol. 1999; 119: 281-284.
- Kim D, Kim J, Yoon JH, Ghim J, Yea K, Song P, et al. CXCL12 secreted from adipose tissue recruits macrophages and induces insulin resistance in mice. Diabetologia. 2014; 57: 1456-1465.
- Rosshart S, Hofmann M, Schweier O, Pfaff AK, Yoshimoto K, Takeuchi T, et al. Interaction of KLRG1 with E-cadherin: new functional and structural insights. Eur J Immunol. 2008; 38: 3354-3364.
- Fourcade J, Sun Z, Benallaoua M, Guillaume P, Luescher IF, Sander C, et al. Upregulation of Tim-3 and PD-1 expression is associated with tumor antigenspecific CD8+ T cell dysfunction in melanoma patients. J Exp Med. 2010; 207: 2175-2186.
- 79. Kalinski P. Regulation of immune responses by prostaglandin E2. J Immunol. 2012; 188: 21-28.
- Yu Y, Chadee K. Prostaglandin E2 stimulates IL-8 gene expression in human colonic epithelial cells by a posttranscriptional mechanism. J Immunol. 1998; 161: 3746-3752.
- Nakayama T, Mutsuga N, Yao L, Tosato G. Prostaglandin E2 promotes degranulation-independent release of MCP-1 from mast cells. J Leukoc Biol. 2006; 79: 95-104.
- Weller CL, Collington SJ, Hartnell A, Conroy DM, Kaise T, Barker JE, et al. Chemotactic action of prostaglandin E2 on mouse mast cells acting via the PGE2 receptor 3. Proc Natl Acad Sci U S A. 2007; 104: 11712-11717.
- Duluc D, Delneste Y, Tan F, Moles MP, Grimaud L, Lenoir J, et al. Tumorassociated leukemia inhibitory factor and IL-6 skew monocyte differentiation into tumor-associated macrophage-like cells. Blood. 2007; 110: 4319-4330.
- Hinz B, Brune K, Pahl A. Prostaglandin E(2) upregulates cyclooxygenase-2 expression in lipopolysaccharide-stimulated RAW 264.7 macrophages. Biochem Biophys Res Commun. 2000; 272: 744-748.
- 85. Mordasky Markell L, Pérez-Lorenzo R, Masiuk KE, Kennett MJ, Glick AB. Use of a TGFbeta type I receptor inhibitor in mouse skin carcinogenesis reveals a dual role for TGFbeta signaling in tumor promotion and progression. Carcinogenesis. 2010; 31: 2127-2135.
- Kesselring R, Thiel A, Pries R, Trenkle T, Wollenberg B. Human Th17 cells can be induced through head and neck cancer and have a functional impact on HNSCC development. Br J Cancer. 2010; 103: 1245-1254.
- Kryczek I, Banerjee M, Cheng P, Vatan L, Szeliga W, Wei S, et al. Phenotype, distribution, generation, and functional and clinical relevance of Th17 cells in the human tumor environments. Blood. 2009; 114: 1141-1149.
- Xiao M, Wang C, Zhang J, Li Z, Zhao X, Qin Z, et al. IFNgamma promotes papilloma development by up-regulating Th17-associated inflammation. Cancer Res. 2009; 69: 2010-2017.
- Ye ZJ, Zhou Q, Gu YY, Qin SM, Ma WL, Xin JB, et al. Generation and differentiation of IL-17-producing CD4+ T cells in malignant pleural effusion. J Immunol. 2010; 185: 6348-6354.
- Liu J, Duan Y, Cheng X, Chen X, Xie W, Long H, et al. IL-17 is associated with poor prognosis and promotes angiogenesis via stimulating VEGF production of cancer cells in colorectal carcinoma. Biochem Biophys Res Commun. 2011; 407: 348-354.

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