

Research Article

Neuropilin-2 Identifies Cardiovascular Precursor Cells and is Required for Vascular Differentiation in Murine Embryonic Stem Cells System

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Abstract

Neuropilins are cell surface receptors that bind Class III semaphorins and Vascular Endothelial Growth Factor (VEGF). While Neuropilin-1 is essential for vascular development, the role of Neuropilin-2 is less well defined. The objective of this study was to determine the role of Neuropilin-2 in early cardiac and vascular differentiation in embryonic stem cells. We report that expression of Neuropilin-2 coincides temporally with Brachyury expression in murine embryonic stem cells and that Neuropilin-2 expression occurs in the majority of Brachyury positive cells. In contrast to Neuropilin-1, we found very few Neuropilin-2 positive cells that co-express the hemangioblast marker Flk-1. Both Neuropilin-2 positive and Brachyury positive cells express mesoderm markers that can differentiate into cardiomyocyte, endothelial and smooth muscle cell types with equivalent efficiency. However, inhibition of Neuropilin-2 expression impairs the differentiation of cardiovascular precursors to endothelial cells and smooth muscle cells, but not cardiomyocytes. The onset of Neuropilin-2 expression identifies a sub-population of Brachyury positive cardiovascular precursors that can differentiate to endothelial, smooth muscle and cardiomyocyte cell types. Furthermore, Neuropilin-2 is functionally required for differentiation of vascular cell types but not cardiomyocytes.

Keywords: Neuropilins; Cardiovascular differentiation, Vascular cells

Introduction

Neuropilins (NRP) -1 and -2 are transmembrane receptors for class III semaphorin and Vascular Endothelial Growth Factors (VEGFs) [1,2]. NRPs are implicated in embryonic neurogenesis and vasculogenesis [3-5], with NRP-1 gene deletion leading to embryonic lethality due to cardiovascular malformations between E10.5 and E13.5 [4]. Embryos lacking NRP-1 display abnormal yolk sac vascular development and subsequent formation of the cardiac outflow tract and aortic arches is also disturbed [2,6]. In contrast, NRP-2 is a receptor for semaphorin IV and VEGF-C, and is required for embryonic vessel development [6-8]. While NRP-2 is expressed in embryonic blood islands in a pattern distinct from NRP-1, NRP-2 is preferentially expressed in the lymphatics and veins of more mature embryos [9]. NRP-2 knockout embryos exhibit defective vascular development with disordered small vessel formation [10], but does not result in early embryonic lethality, as found in VEGF and VEGFR-2 knockout embryos. Knockout of both NRP-1 and NRP-2 results in an even more pronounced phenotype, with severe impairment of yolk sac vascular development and early embryonic lethality [4,6]. These observations suggest a role for NRP-2 in embryonic vascular differentiation although the mechanisms involved underlying this process remain to be defined.

Brachyury (Bry), one of the most important mesoderm inducing factors [25] is a member of the T-box transcription factor gene family and is required for posterior mesoderm and notochord formation during vertebrate development [11]. In murine embryonic stem cells,

Bry expression is an important tool for tracking mesendodermal precursors [12]. To facilitate the use of Bry as a mesendodermal marker, the GFP gene was targeted to the Bry locus (Bry-GFP) in Murine Embryonic Stem Cells (mESC), with GFP expression correlating with endogenous Bry transcription. Consequently, developing mesendodermal progenitors can be identified and separated based on GFP expression [12].

Recently, cardiovascular progenitors were shown to differentiate from Bry⁺/Flk-1⁺ cells acquired from EBs of Bry-GFP mESCs [13]. We previously reported that NRP-1 expression identifies Bry⁺ and Flk-1⁺ vascular precursors in both murine and human embryonic stem cells [14]. However, the relationship between Bry and NRP-2, if any, are unknown. Since NRP-2 and Bry are involved in early aspects of cardiovascular development, we hypothesized that NRP-2 expression may temporally correspond to Bry expression, and may have an important role in the formation of cardiovascular tissues including cardiomyocytes, endothelial cells and smooth muscle cells. In this study, using Bry-GFP mESCs, we determined that the timing of NRP-2 expression coincides with Bry expression during mesoderm differentiation, and that NRP-2 is functionally required for the differentiation of mesodermal precursors into vascular cell types but not cardiac precursors.

Materials and Methods

mESC Culture

Bry/GFP knock-in murine embryonic stem cells (Bry-GFP mESCs) [12] were initially grown on mitomycin-c-treated

primary murine embryonic fibroblasts in ES Cell Qualified DMEM (EmbryoMax®, CHEMICON) supplemented with 1.5 x 10⁻⁴ M monothioglycerol (Sigma, St. Louis, MO), 15% Fetal Calf Serum (FCS) (Hyclone, Thermo Fisher Scientific, Waltham, MA), 10 mg/ml leukemia inhibitory factor (# L5158-5UG, Sigma), 10 mg/ml recombinant human Bone Morphogenetic Protein (BMP) 4 (#314-BP, R&D Systems, Minneapolis, MN), and transferred to feeder-free, serum-free conditions in N2B27 medium (Invitrogen, Carlsbad, CA) supplemented with 2 mM L-glutamine (Sigma), 0.1 mM β-mercaptoethanol (BDH), 10 mg/ml LIF and 10 mg/ml BMP4 as previously showed [15].

Mesodermal differentiation

Bry-GFP mESCs were differentiated as EBs by trypsinization of mESCs and plating at a density of 1.5 x 10³ cells/cm² in 150 x 25 mm suspension culture dishes in N2B27 medium supplemented with BMP4 and basic fibroblast growth factor (bFGF), both at 5 mg/ml (# 233FB, R&D Systems).

Cardiovascular Differentiation: Day 3.5-EBs derived from Bry-GFP mESCs were dissociated to single cells by treatment with 0.25% trypsin (Invitrogen) and stained with antibodies for flow cytometry analysis as described below. NRP-2⁺/Flk-1- cells and Bry⁺/Flk-1- cells were sorted using a BD FACSAria and reaggregated for 20 hours at a density of 4 x 10⁵/ml in ultra low attachment 24-well plates (Costar, Thermo Fisher Scientific) containing N2B27 media supplemented with 5 mg/m rhVEGF (# 293VE, R&D Systems) and 30 ng/ml bFGF. Reaggregated cells were then trypsinized and plated in 24-well cell culture plates (Costar) at density of 2 x 10⁴ /cm², and cultured for an additional 5 - 7 days in N2B27 medium. For cardiomyocyte differentiation, medium was supplemented with 10 mg/ml rhBMP4 and 10 ng/ml rhVEGF; for endothelial cell differentiation, the medium was supplemented with 10 ng/ml rhVEGF; for smooth muscle differentiation, the medium was supplemented with 10 ng/ml rhPDGF-BB (# 220BB, R&D Systems).

Flow cytometry analysis

Day 3.5-EBs derived from Bry-GFP mESCs were dissociated to single cells by treatment with 0.25% trypsin. Cells were incubated with the following antibodies: APC conjugated anti-mouse Flk-1 IgG (# 17-5821-81, eBioscience, San Diego, CA) or co-incubated with goat polyclonal anti-NRP-2 IgG (# AF2215, R&D Systems) and Flk-1-APC for 40 minutes at 4°C. Cells were washed with 0.2 mM EDTA-, 1% FCS-PBS, centrifuged at 500g for 5 minutes then incubated with swine anti-goat IgG-phycoerythrin (# MBS674933, MYBIOSOURCE, San Diego, CA) for 30 minutes at 4°C. The quantity of Bry-GFP, Flk-1 and NRP-2 expressing cells was determined by flow cytometry analysis using a BD FACSCalibur.

Immunofluorescence microscopy

Reaggregated NRP-2⁺/Flk-1- and Bry⁺/Flk-1- cells were washed with 1X PBS and fixed in 4% paraformaldehyde (Electron Microscopy Sciences) for 10 minutes at room temperature. The following primary antibodies were applied to cells overnight at 4°C: cardiac lineage differentiation, Nkx2.5 (#sc-74692, Santa Cruz biotechnology), Islet-1 (#AF1837, R&D Systems), Troponin I (#sc-8118, Santa Cruz biotechnology) and Connexin 43 (#35-5000, Invitrogen); endothelial and smooth muscle cell differentiation, VE-Cadherin (#550548, BD

Pharmingen, San Diego, CA), CD31 (#14-0311-82, eBioscience) and SM-22 (#ab14106 Abcam, San Francisco, CA), α-smooth muscle actin (#A2547, Sigma). Cells were washed with 0.5% Tween 20 in PBS, and detected with donkey anti-goat, anti-rabbit, and anti-rat IgG conjugated Alexa fluor 488 or 555 (Invitrogen). Cells were imaged with an Olympus IX71 fluorescence microscope.

Reverse transcription-polymerase chain reaction (RT-PCR)

Total RNA was extracted from sorted NRP-2⁺ cells and Bry⁺ cells using RN easy Mini Kit (#74104, QIAGEN, Valencia, CA). First-strand cDNA was synthesized using M-MLV Reverse Transcriptase (#9PIM170, Promega, Madison, WI), followed by PCR using PuReTaq Ready-To Go PCR Beads (#27955701, GE Healthcare, Pittsburgh, PA). PCR cycle conditions were: 1 cycle of 94°C for 5 minutes; 35 cycles of 94°C for 20 seconds, 60°C for 20 seconds, 72°C for 40 seconds; and 1 cycle of 72°C for 7 minutes. GAPDH was used as an internal control. The sequences of primers used were (forward and reverse): PDGFR-β, GTCTGGTCTTTGGATCCT and AAGGCTGGTTACAGTTGGC;

Cadherin 11, CCGACTTGTGAATGGGACTC and CTCCGCAGTCAGCTTCTTCT;

Flk-1, TCTGTGGTTCTGCGTGGAGA and GTATCATTTCCAACCACCCCT;

PDGFR-α, AATCCTGCAGACGAGAGCAC and GCCACCAAGGGAAAAGATTT;

GAPDH, TTGCCATCAATGACCCCTCA and CGCCCCACTTGATTGAA.

shRNA Knockdown of NRP-2 in Bry-GFP mESCs: Murine NRP-2 shRNA plasmid vectors (#sc-36041-SH, Santa Cruz biotechnology) were isolated from E.Coli using Endo Free DNA Maxi Kit (#12362, QIAGEN). NRP-2 shRNA vectors were then transfected into HEK293 cells plated at a density of 2.5 x 10⁵ cells/well on 6-well plates. Transfection was performed using DNA/Arrest-In reagent according to the manufacturer's instructions. Lent viral particles were collected from the cell supernatant and used for transfection of Bry-GFP mESCs. Bry-GFP mESCs were cultured at density of 1.5 x 10⁴/cm² in N2B27 medium for 24 hours. The medium was then replaced with 1 ml of the supernatant medium obtained from the transfected 293 cell supernatant medium, and after 48 hr another 1 ml N2B27 medium was added with culture at 37°C, 20% Oxygen, 5% CO₂. Transfected cells were selected by culture for 6 additional passages in N2B27 medium µg/ml containing puromycin1.

Statistical analysis

Experimental data were analyzed by unpaired 2-tailed t-test. Results are expressed as mean ± SEM. Differences were deemed significant when P < 0.05. Statistical analyses were performed using Prism, Version 4.00 (GraphPad Software, La Jolla, CA, USA).

Results

NRP-2 identifies Bry⁺ mesodermal precursor cells

Bry is an important regulator of early mesodermal cell commitment during development and in the differentiation of murine ESCs [16]. Consistent with previous findings [12,15], flow cytometry

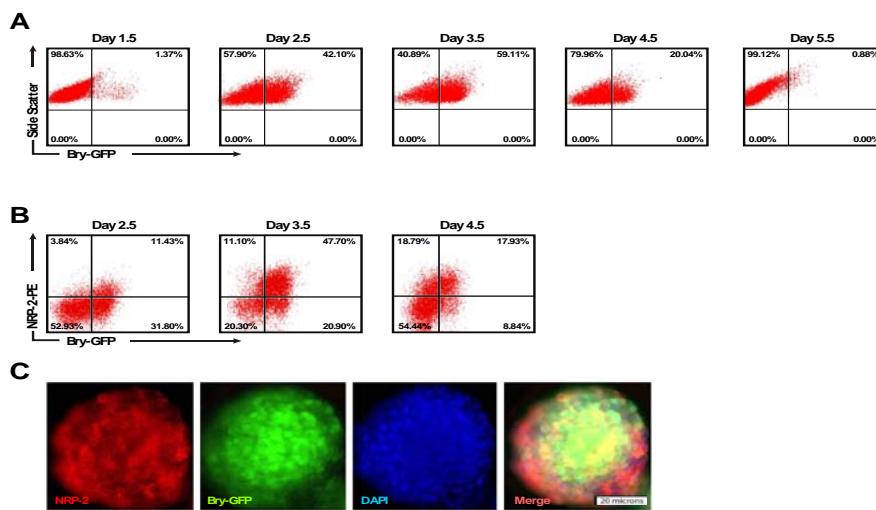


Figure 1: NRP-2 expression coincides with Bry in differentiating mESCs. EBs were dissociated at the indicated time points after differentiation and the number of Bry-GFP and NRP-2⁺ cells determined by flow cytometry. Panel A) Representative flow cytometry plots of Bry-GFP time course of expression. Panel B) Representative flow cytometry plots of NRP-2 and Bry-GFP expression over time. Panel C) Immunofluorescent photomicrographs of EBs stained with antibody against NRP-2, and endogenous Bry-GFP expression. Results shown are representative of at least five independent experiments.

analysis of GFP expression in EBs derived from Bry-GFP mESCs showed that Bry expression was up-regulated at 2.5 days, peaked at 3.5 days, decreased dramatically at 4.5 days, and was undetectable at 5.5 days (Figure 1A). To determine if NRP-2 was co-expressed in Bry⁺ cells, we performed flow cytometry analysis of dissociated EBs 3.5 days after differentiation in mesoderm-inducing conditions. The majority of Bry⁺ cells co-expressed NRP-2 (> 80%) at day 3.5 (Figure 1B). Immunofluorescence staining of NRP-2 of day 3.5 EBs derived from Bry-GFP mESCs confirmed that NRP-2 positive cells co-expressed Bry⁺ cells at that stage of differentiation (Figure 1C). Collectively, these results indicate that NRP-2 identifies the majority of Bry⁺ differentiated mESCs under mesoderm-inducing conditions.

BMP4 signaling is required for NRP-2 and Bry expression during mesodermal differentiation

Our prior studies indicated that Bry⁺ cells are readily obtained under serum free conditions in the presence of BMP4 and bFGF [15]. However, the growth factor requirement for NRP-2 expression in differentiating mESCs is not known. To determine the growth factor requirements for NRP-2 and Bry expression we differentiated Bry-GFP mESCs as EBs under serum free conditions in the presence of the following growth factor conditions: no treatment, BMP4 and bFGF, bFGF, Activin A, or 15% FCS. The quantity of Bry-GFP⁺ and NRP-2⁺ cells was determined by flow cytometry analysis. We found that the combination of BMP4 and bFGF was as effective as 15% FCS for induction of Bry and NRP-2 expression, and that these growth factors induce NRP-2 expression in the majority of Bry⁺ cells (Figure 2A). To verify the requirement for BMP4 signaling, we antagonized BMP4 with Noggin. Noggin effectively blocked the differentiation of Bry-GFP mESCs, resulting in a significant decrease in the quantity of Bry⁺ or NRP-2⁺ cells (Figure 2A, B). Collectively, our findings indicate that the induction of both Bry and NRP-2 expression requires BMP4 signaling.

Both NRP-2⁺ and Bry⁺ cells express mesoderm-specific transcripts and the majority of NRP-2⁺ cells are Flk-1 negative

Our flow cytometry data indicated that NRP-2 is co-expressed with Bry in differentiating mESC, suggesting these cells were mesoderm precursors. To provide additional evidence that both NRP-2⁺ and Bry⁺ cells were equivalent mesoderm precursors, we used PCR analysis to determine if both cell types expressed mRNAs found in mesodermal cells. Transcripts encoding PDGFR- β , PDGFR- α , Cadherin 11 and Flk-1 mRNA were found in both Bry⁺ and NRP-2⁺ cells, but not in undifferentiated mESCs (Figure 3A).

Flk-1 expression in mesodermal progenitors identifies vascular and hematopoietic progenitor cells [12,15,17]. However, Flk-1 expression is typically observed in only 15 - 20% of cells differentiated under the conditions used in our studies [12,15,17]. To determine the proportion of NRP-2⁺ cells that express Flk-1, flow cytometry analysis was performed on dissociated single cells from EBs after

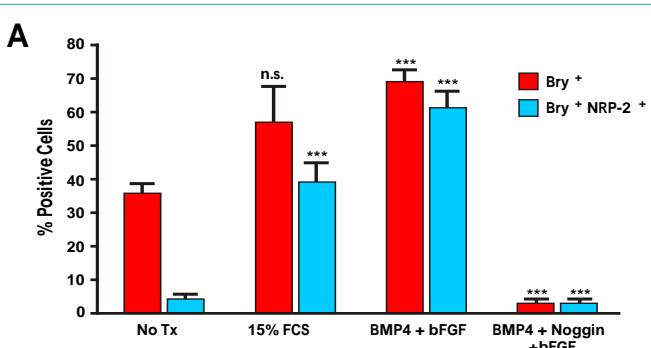


Figure 2A: NRP-2 identifies Bry-GFP⁺ cells in differentiating mESCs, and requires BMP4 for induction of NRP-2 expression. Panel A) The quantity of Bry-GFP⁺ and NRP-2⁺ cells in EBs grown in either 15% FCS, serum free conditions with BMP4 and bFGF, or with antagonism of BMP4 with Noggin in the presence of BMP4 and bFGF, as determined by flow cytometry analysis. Results shown are the means +/- SEM of five independent experiments. (Bry⁺: No Tx vs. 15% FCS: $35.15 \pm 3.0\%$, P = n.s. No Tx vs. BMP4 + bFGF: $71.75 \pm 2.9\%$, P = 0.001. BMP4 + bFGF vs. Noggin + BMP4 + bFGF: $7.25 \pm 1.5\%$, P = 0.001. Bry⁺/NRP-2⁺: No Tx vs. 15% FCS: $5.0 \pm 1.1\%$, P = 0.0009. No Tx vs. BMP4 + bFGF: $66.75 \pm 3.6\%$, P = 0.001. BMP4 + bFGF vs. Noggin + BMP4 + bFGF: $7.3 \pm 1.7\%$, P = 0.001).

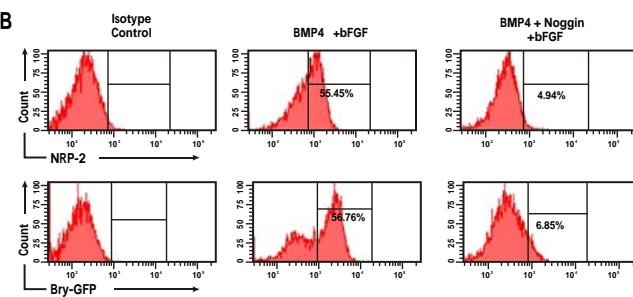


Figure 2B: Representative flow cytometry plots showing Bry-GFP and NRP-2 expression following culture with BMP4 and bFGF, or with Noggin, BMP4 and bFGF.

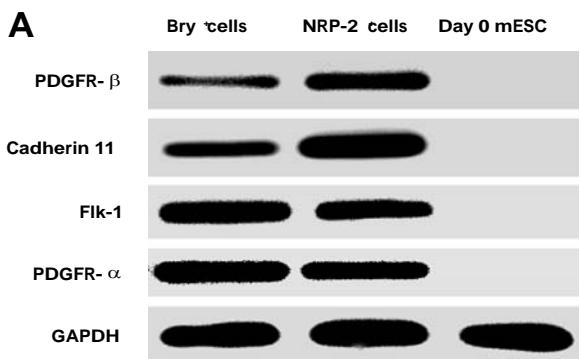


Figure 3A: Bry-GFP $^{+}$ and NRP-2 $^{+}$ cells express overlapping mesodermal markers. Bry-GFP $^{+}$ and NRP-2 $^{+}$ cells were sorted from EBs and mRNA obtained. RT-PCR was performed to detect mesodermal transcripts for PDGFR- β , Cadherin-11, FLK-1; PDGFR- α . GAPDH was included as an internal control. Results shown are representative of three independent experiments.

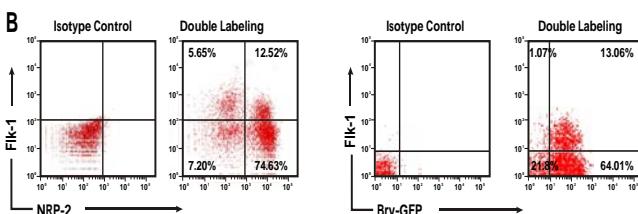


Figure 3B: Flow cytometry plots of Flk-1 expression in NRP-2 $^{+}$ and Bry $^{+}$ cells. Similar to Bry $^{+}$ cells, the majority of NRP-2 $^{+}$ cells did not express Flk-1 during early stage differentiation. Results are representative of three independent experiments.

differentiation in mesoderm-inducing conditions. We identified that the majority of NRP-2 $^{+}$ cells were negative for Flk-1. (Figure 3B). These data indicate that Bry $^{+}$ and NRP-2 $^{+}$ cells carry similar mesodermal transcript markers, and that NRP-2 $^{+}$ cells are Flk-1 negative. Collectively, these data also suggest that these cells may be cardiovascular progenitors.

NRP-2 $^{+}$ /Flk-1 $^{-}$ cells are cardiac and vascular progenitors

Our findings indicated that NRP-2 expression overlaps with that of Bry in mESCs differentiated under mesoderm-inducing conditions. However, the differentiation potential of either cell type remained to be determined. Given that NRP-2 $^{+}$ cells were largely Flk-1 $^{-}$ and, as prior work indicated, that cardiovascular precursors may be derived from Bry $^{+}$ /Flk-1 $^{-}$ cells differentiated under similar conditions to those

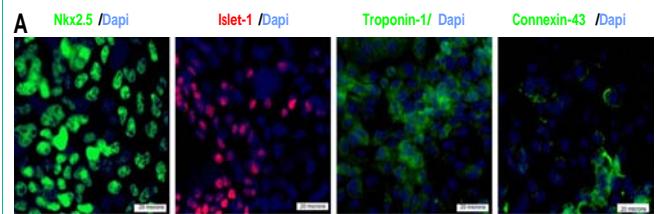


Figure 4A: NRP-2 $^{+}$ /FLK-1 $^{-}$ and Bry-GFP $^{+}$ /FLK-1 $^{-}$ cells differentiate into cardiomyocytes, endothelial cells and smooth muscle cells with equivalent efficiency. Panel A) Representative immunofluorescence photomicrographs showing cardiomyocyte differentiation of NRP-2 $^{+}$ /FLK-1 $^{-}$ cells following treatment with BMP4 and VEGF. Staining was with antibodies against Nkx2.5, Islet-1, Troponin-I, and Connexin-43.

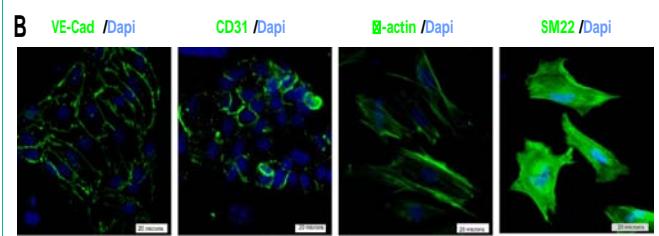
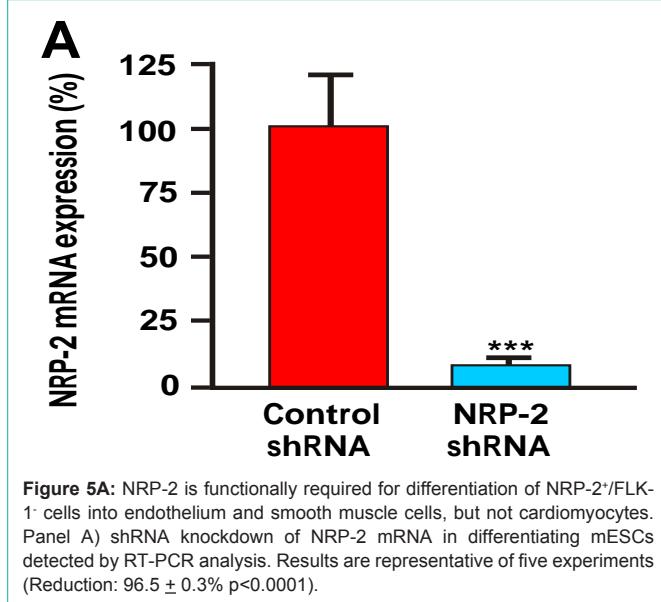
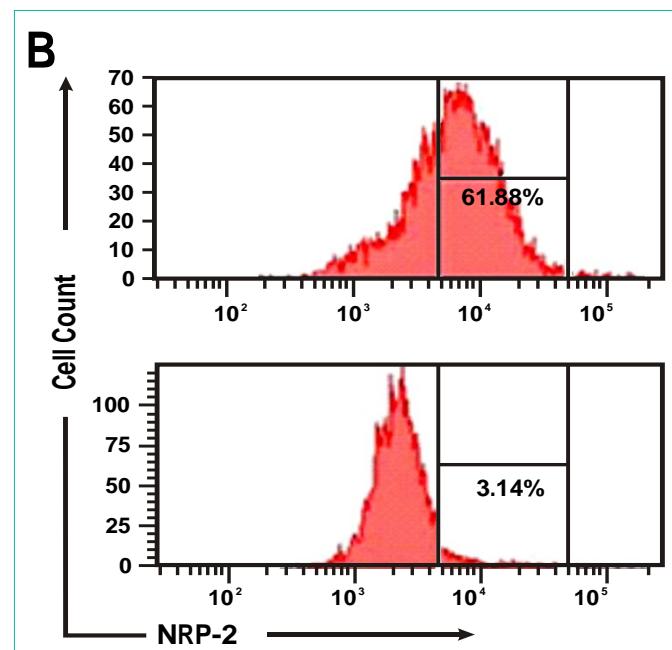
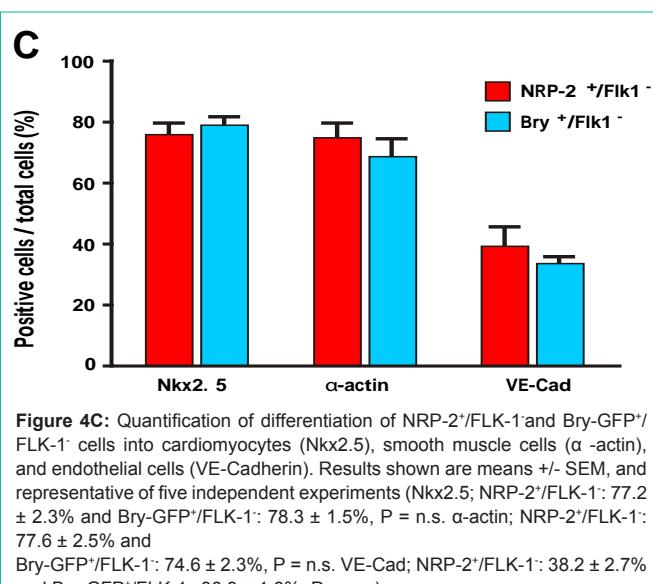


Figure 4B: NRP-2 $^{+}$ /FLK-1 $^{-}$ cells differentiated into endothelial cells following treatment with VEGF and smooth muscle cells after treatment with PDGF-BB. Staining is shown with antibodies against endothelial specific VE-Cadherin and CD31, and smooth muscle cell specific α -actin and SM22.

used here [12,15,17], we hypothesized that the Bry $^{+}$ /Flk-1 $^{-}$ or NRP-2 $^{+}$ /Flk-1 $^{-}$ cells may be cardiovascular precursors. To test this hypothesis, we performed flow cytometry-based cell sorting of Bry $^{+}$ /Flk-1 $^{-}$ and NRP-2 $^{+}$ /Flk-1 $^{-}$ cells from day 3.5 EBs. We then differentiated these cells under reaggregation conditions that promote cardiovascular differentiation and determined the proportion of cells expressing markers of cardiac, endothelial and smooth muscle cell types (Figure 4A). We identified that Nkx2.5 $^{+}$ /islet-1 $^{+}$ /troponin-I $^{+}$ /connexin-43 $^{+}$ cardiomyocyte progenitors could be derived from both NRP-2 $^{+}$ /Flk-1 $^{-}$ (Figure 4A, C) and Bry $^{+}$ /FLK-1 $^{-}$ cells (data not shown) in presence of VEGF and BMP4. Moreover, VE-Cadherin $^{+}$ /CD31 $^{+}$ endothelial progenitors were induced by VEGF alone, while SM22 $^{+}$ / α -actin $^{+}$ smooth muscle progenitor cells were induced by PDGF alone from both the NRP-2 $^{+}$ /Flk-1 $^{-}$ (Figure 4B, C) and Bry $^{+}$ /Flk-1 $^{-}$ populations (data not shown). Collectively, our findings indicate that Bry $^{+}$ /Flk-1 $^{-}$ and NRP-2 $^{+}$ /Flk-1 $^{-}$ cells are cardiovascular precursors with differentiation potential to cardiomyocyte, endothelial and smooth muscle cell types.

NRP-2 is functionally required for differentiation of endothelial and smooth muscle cells, but not cardiomyocytes

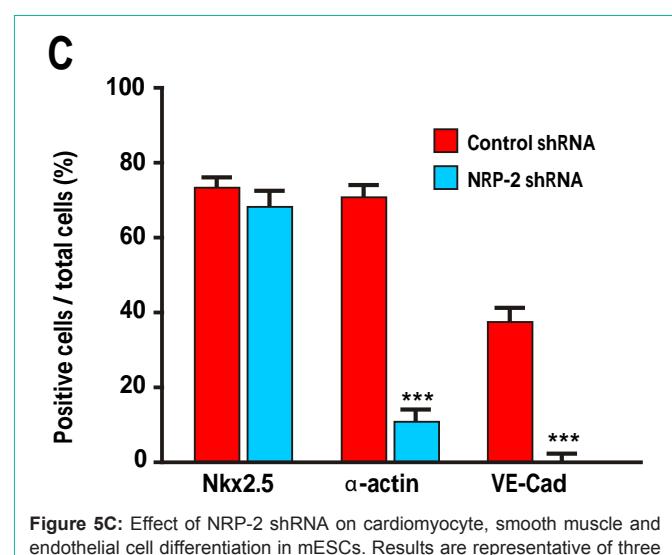
To determine the function of NRP-2 in the differentiation of cardiovascular precursors to cardiomyocytes, endothelial and smooth muscle cells, we transfected Bry-GFP mESCs with NRP-2 shRNA vectors to suppress NRP-2 expression. We found that NRP-2 shRNA had no effect on the expression of Bry-GFP, but efficiently suppressed NRP-2 mRNA and protein expression (Figure 5A, B). We then isolated Bry $^{+}$ /FLK-1 $^{-}$ -cardiovascular precursors from NRP-2 shRNA-transfected Bry-GFP mESCs, and determined their differentiation efficiency to endothelial, smooth muscle and cardiomyocyte cell types. We found that transfection with NRP-2 shRNA resulted in a



significant decrease in the number of endothelial and smooth muscle cells, but had no effect on the quantity of cardiomyocytes (Figure 5C, D). These data indicate a requirement for NRP-2 in the differentiation of vascular cell types, but not for cardiomyocyte differentiation from Bry⁺/Flk-1⁺ cardiovascular precursors.

Discussion

Previously we reported that the onset of NRP-1 expression is an early event in mesoderm formation and corresponds with Bry and Flk-1 expression in murine and human embryonic stem cells, and that NRP-1 is functionally required for the differentiation of mesodermal precursors into endothelial cells [14]. The goals of the current study were to determine if NRP-2 expression displayed a similar temporal expression pattern during mesoderm formation, determine the differentiation potential of NRP-2⁺/Flk-1⁺ cells, and determine the function of NRP-2 in the murine ESC differentiation model.



Our time course studies in differentiating mESCs indicate that NRP-2 expression is dependent on BMP4 stimulation and coincides with Bry. Surprisingly, under these differentiation conditions NRP-2 expression is found in the majority of Bry⁺ cells, but does not appear to be functionally required for Bry expression as transfection with NRP-2 shRNA had no effect on the differentiation capacity of Bry-GFP mESCs. These findings provide novel information on the timing and distribution of NRP-2 expression in mesoderm precursors. At slightly later stages of embryonic development, NRP-2 expression is found predominantly in veins, lymphatic's and in the heart of murine embryos [9,18-20]. Very few prior studies have assessed the onset of

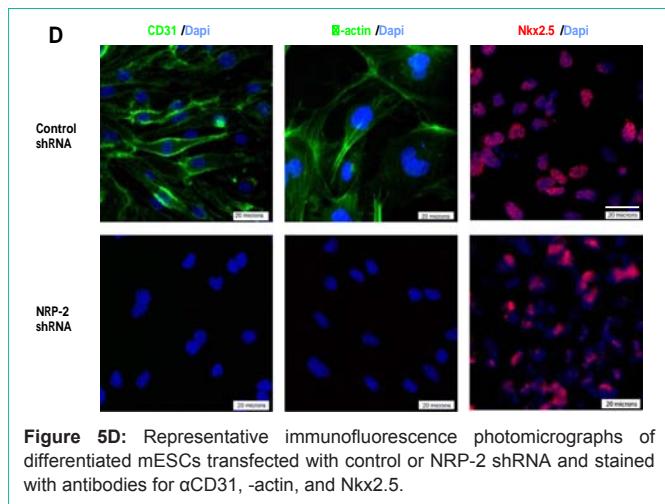


Figure 5D: Representative immunofluorescence photomicrographs of differentiated mESCs transfected with control or NRP-2 shRNA and stained with antibodies for α CD31, -actin, and Nkx2.5.

expression of the neuropilins near the time of mesoderm formation in early development. In the chick embryo, NRP-2 expression is found in the blood islands, as well as the primitive streak and lateral plate mesoderm, and provides an independent verification that our findings in this ESC model accurately replicate *in vivo* events [9].

We found that the cellular potential of the NRP-2 $^{+}$ /Flk-1 $^{-}$ cells was consistent with a population of cardiovascular precursors with similar differentiation capacity to Bry $^{+}$ /Flk-1 $^{-}$ cells [13]. We were able to obtain differentiation to endothelial, smooth muscle and cardiomyocyte cell types from NRP-2 $^{+}$ /Flk-1 $^{-}$ -mesodermal precursors, illustrating the overlapping differentiation potential of these two cell types (Figure 6). While NRP-2 expression marked a population of cells with differentiation capacity to cardiac and vascular cell types, NRP-2 was only functionally important for differentiation towards endothelial and smooth muscle cell fates, but not to cardiomyocytes. These data are also consistent with findings in murine knockout models where the NRPs are functionally required during the early stages of vascular development [6,21].

The fact that NRP-2 is expressed in the majority of Bry $^{+}$ cells under mesoderm-inducing differentiation conditions and is not functionally required for cardiomyocyte formation implies a role for NRP-2 during primitive streak formation and potentially in the migration and patterning of mesodermal cells [17]. Potential candidate molecules to direct primitive streak migration may include the Semaphorins. Semaphorins are major regulators of morphogenesis in the nervous system and in malignant cellular transformation [22,23]. Semaphorin expression in early mammalian development is not well characterized, but in Xenopus embryos Semaphorins 3A, 3F and 4B are localized to somatic mesoderm, making it plausible that Semaphorins may play a role in the guidance and migration of NRP-2 $^{+}$ mesoderm progenitors during gastrulation [20,24].

The similar expression profiles of NRP-2 and Bry provides a convenient means to identify mesoderm precursors based on the cell surface expression of NRP-2. This correlation in marker expression may be of importance for two clinically relevant applications: tissue biomarkers and tissue engineering of stem cells. Thus, the correlation of Bry and NRP-2 expression may facilitate the identification of cells with cardiovascular differentiation potential from ESCs, induced pluripotent stem cells and other cell sources, including those from

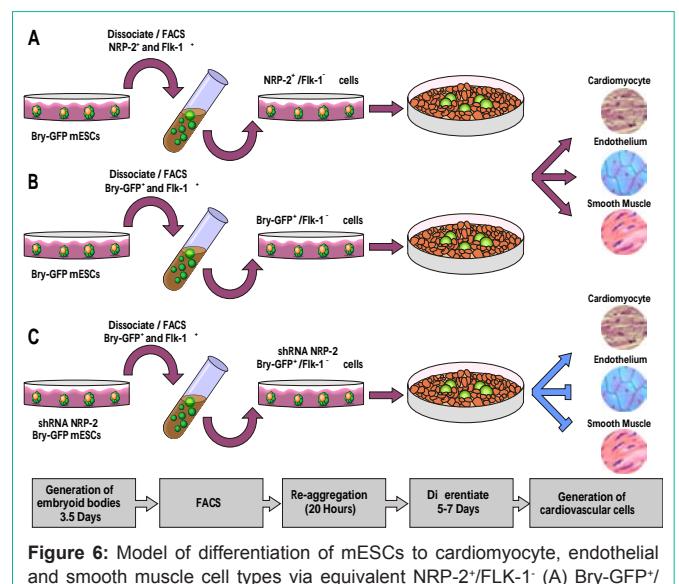


Figure 6: Model of differentiation of mESCs to cardiomyocyte, endothelial and smooth muscle cell types via equivalent NRP-2 $^{+}$ /FLK-1 $^{-}$ (A) Bry-GFP $^{+}$ /FLK-1 $^{-}$ precursors and also NRP-2 shRNA-treated Bry-GFP $^{+}$ /FLK-1 $^{-}$ cells.

non-genetically modified cell lines. The further extension of these findings using these cells will facilitate the translation of cardiovascular precursors as a therapy for cardiovascular disease.

Acknowledgement

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