

Review Article

Nutritional Status of Patients with Malignant Obstructive Jaundice Evaluated by Nutritional Risk Screening and the Indocyanine Green Retention Test

Fang Liu¹; Chuang Yang^{2*}¹Mianyang Central Hospital, School of Medicine University of Electronic Science and Technology of China, Mianyang, Sichuan, PR China²Department of Hepatobiliary Surgery, Fokind Hospital, Tibet University, Tibet, PR China***Corresponding author: Chuang Yang**

Department of Hepatobiliary Surgery, Fokind Hospital, Tibet University, Tibet, No.14, Linkuo North Road, Lhasa City, Tibet Autonomous Region, PR China.

Tel: 13458018352

Email: ycdocor2023@vip.sina.cn

Received: July 02, 2024

Accepted: July 22, 2024

Published: July 29, 2024

Introduction

Jaundice is characterized by yellow discoloration of the skin, sclera, and mucous membranes resulting from the accumulation of excess bilirubin [1]. Bilirubin is formed by heme in the presence of heme oxygenase and biliverdin reductase [2,3]. Obstructive jaundice, also known as surgical jaundice, is a relatively common clinical condition usually caused by complete mechanical obstruction of the intra/extrahepatic bile ducts and obstruction of bile flow into the intestine, resulting in cholestasis or reverse flow of ester-type bilirubin into the blood, eventually leading to jaundice [4,5].

Malignant Obstructive Jaundice (MOJ) is caused by a primary malignancy or direct/indirect obstruction of the bile duct resulting from a distant malignancy with main clinical manifestations that include hyperbilirubinemia, yellow discoloration of tissues, and dilation of the bile duct [5,6]. Obstruction of the bile duct above the common hepatic duct is termed a high duct obstruction, which is usually caused by hilar cholangiocarcinoma, hepatocellular carcinoma, local infiltration and compression resulting from gallbladder carcinoma, and invasion of the hilar bile ducts [5-7]. Obstruction of the bile duct from the common hepatic duct to the duodenal papilla, known as low duct obstruction, is usually caused by malignant tumors of the ampulla and

Abstract

Malignant Obstructive Jaundice (MOJ) is typically caused by malignant tumors of hepatopancreatic biliary origin and less frequently by metastatic primary malignancies of non-hepatopancreatic biliary origin. MOJ is associated with an unusually high risk of malnutrition. Therefore, effective screening of nutritional status is especially important for early detection of MOJ. We recommend the indocyanine green retention test combined with the nutritional risk screening for assessment and dynamic real-time monitoring of nutritional status to improve the treatment outcomes of MOJ patients.

Keywords: Malignant obstructive jaundice; Dystrophy; Nutritional status; Liver reserve function; Indocyanine green

surrounding tissues of the pancreas, lower segment of the common bile duct, and duodenal papillae [8-10]. Previous studies have reported that MOJ in up to 14% of patients is due to metastatic cancers of the kidney, lung, and stomach [11,12]. MOJ is usually diagnosed in the late stage of disease because of the absence of symptoms. Progressive obstruction of the bile duct leads to complete obstruction of bile excretion. Therefore, most patients have relatively high bilirubin levels and abnormal liver function, requiring effective decoy therapy to create opportunities for surgery or to improve quality of life [13,14].

Impact of MOJ on Liver Function

The liver, which is the largest organ of the digestive tract, is involved in various functions that support metabolism, immunity, digestion, detoxification, and vitamin storage, among others, including rapid elimination of free bilirubin to maintain normal serum levels [15,17]. The liver has a rich lymphatic network that is a major component of the hepatic microcirculation and produces 25%–50% of the lymph fluid in the thoracic ducts [18]. The lymphatic network circulates along the bile ducts and portal veins, and the larger portal veins form bundles composed of 6–10 lymphatic vessels [18-21]. When obstructed, there is

a marked increase in pressure throughout the biliary system that can rupture the capillaries and ciliary ducts, which communicate directly with the lymphatic system [22,23], potentially causing the accumulation of bilirubin in the blood via the sinusoidal membranes or close junctions of the hepatocytes, resulting in hyperbilirubinemia [24,25]. MOJ can increase serum levels of ester-type bilirubin, which is not excreted in the feces, thereby reducing levels in the urine. Moreover, the lack of bilirubin entering the digestive tract results in pale, "putty" colored stools and an absence of urobilinogen in the urine [26-28]. Long-term MOJ can impair bilirubin uptake, transport, and esterification by the hepatocytes, and beta-glucuronidase in many tissues can promote the hydrolysis of ester-type bilirubin, resulting in increased serum concentrations of nonester-type bilirubin [27-30].

As a result of increased pressure in the bile ducts and capillaries, the portal veins within Green's sheath are compressed, resulting in increased pressure in the sinusoidal sinusoids, decreased hepatic blood flow, fibrotic changes to the hepatic lobes, and prolonged cholestasis, resulting in biliary cirrhosis [31-33]. In a state of MOJ, the hepatic arteries are dilated, distorted, and the vascular plexus surrounding the bile ducts is markedly hyperplastic with irregular thickening of the arterial walls, which can result in crescent-shaped changes and narrowing of the arterial lumen and rupture or disappearance of the internal arterial elastic lamina, causing loss of arterial blood flow, eventually resulting in decreased hepatic blood flow and portal hypertension [33,34]. Cirrhosis is characterized by increased pressure in the portal veins, hyperplasia of the hepatic lymphatic system, and reabsorption of excess fluids in the liver and internal organs resulting in increased flow of hepatic lymph (up to 30 times the normal volume), which promotes ascites formation. Meanwhile, portal hypertension leads to gastrointestinal congestion, impairment of the gastrointestinal mucosal barrier and intestinal vascular barrier, formation of stress ulcers, translocation of the intestinal bacteria through the portal vein to the liver, leading to biliary infection and massive endotoxin production, which can result in disseminated intravascular coagulation [34-36].

Increasing evidence suggests that severe obstruction of the biliary tract leads to impaired liver function, immune dysfunction, coagulopathy, significant reductions to the volume of bile in the intestinal tract, and impaired feedback to the bile acid intestinal-hepatic circuits, resulting in impaired absorption of lipolytic vitamin K, which leads to vitamin K deficiency, further affecting the synthesis of hepatic coagulation factors II, VII, IX, and X, resulting in inhibition of the coagulation process and a tendency for severe bleeding of the nasal cavity, gingiva, and internal organs [36-39].

Effects of MOJ on Nutritional Status

Malnutrition affects 15% to 60% of older adults, up to 30% of hospitalized patients [40,41], and to varying degrees in patients with MOJ, with an incidence of malignant dystrophy as high as 50% [42,43]. In addition, malnutrition is associated with diminished bile flow into the gut, impaired circulation of bile acid in the intestines and liver, dysregulation of the intestinal microbiota, and promotion of inflammatory responses in the gut and liver, which further inhibits secretion of bile, disrupts the integrity of the intestinal mucosal barrier, and impairs digestion and absorption in the intestines [35,42,44]. In addition, intrahepatic accumulation of bile salts increases oxidative stress and apoptosis of hepatocytes by inhibiting production of cyto-

chrome P450 in addition to reductions in oxidative and aerobic metabolism. Inhibition of cytochrome P450 and microsomal mixed-function oxidase affects the metabolism of drugs and other substances [45,46].

MOJ leads to reduced liver function and subsequent decreased production of albumin, coagulation factors, and immunoglobulins, resulting in diminished humoral immunity [42,47-49]. As the most important plasma protein synthesized by the liver, albumin is an important biomarker of disease severity. Although changes to serum albumin are usually absent in the early stages of biliary obstruction [50], prolonged obstruction can lead to impaired hepatic production of albumin and poor protein intake due to intestinal bile deficiency and loss of appetite, resulting in decreased serum albumin levels to >30 g/L in 56% of patients. Hence, low serum albumin levels are an important predictor of severe impairment of liver function [37,51,52]. Even after the obstruction is resolved, improvement in serum albumin levels may require a relatively long period [53]. These findings indicate that MOJ adversely affects nutrient absorption and serum albumin levels, thereby highlighting the importance of monitoring nutritional status in clinical practice.

Indocyanine Green (ICG) Retention Test in Combination with Nutritional Risk Screening to Assess Nutritional Status

Rapid short-term (1–2 weeks) increases in bilirubin levels can lessen the effects of compromised nutrient absorption, although the risk of malnutrition may increase significantly over time. MOJ can severely impair nutrient absorption and adversely affect prognosis and prolong hospitalization [54,55]. In addition, percutaneous transhepatic common ductal drainage and endoscopic transnasal ductal drainage can improve patient survival but increases the length of hospitalization and associated costs [56-58]. The severity of malnutrition and serum bilirubin levels vary among patients, thus early detection and treatment of MOJ are particularly important [59].

Poor nutritional status is common in hospitalized patients, especially those with chronic diseases, such as cirrhosis, advanced malignancies, and end-stage liver disease [60]. Hence, monitoring of nutritional status in a timely manner may be useful to predict morbidity and mortality in clinical practice, particularly for critically ill patients [61].

The Nutrition Risk Assessment Scale (NRS-2002) is a relatively simple, noninvasive, and low-cost tool that is commonly used in clinical practice to assess nutritional status and prevent poor treatment outcomes [62,63]. Since the degree of obesity varies among patients, the Body-Mass Index (BMI) alone is insufficient to accurately assess nutritional status, which is influenced by numerous factors, including sodium retention, edema, and limb vein thrombosis [64,65]. The presence of dystrophy in MOJ cannot be ruled out by a high BMI [66]. In addition, BMI is not a reliable biomarker for children, adolescents, and the elderly [62]. The NRS-2002 is reportedly appropriate for assessment of patients with early and mild liver disease, but has limited use for those with more severe liver disease [67-69]. Hence, combinations of tools can more accurately assess the nutritional status of high-risk patients and those in the intensive care unit [70].

The NRS-2002 combined with the ICG retention test more accurately reflects the nutritional status of patients with MOJ. At present, three methods are commonly used to assess liver reserve: (1) biochemical tests of serum biomarkers (i.e., aminotransferases, bilirubin, alkaline phosphatase, albumin, and pro-

thrombin); (2) composite scoring systems; and (3) quantitative tests of liver function [15,71,72]. However, biochemical tests of serum biomarkers reflect liver function only in one dimension with no predictive value for postoperative liver failure. In addition, the degree of variability in biomarkers is not completely consistent with the severity of liver disease and does not permit precise assessment of liver reserve function [15,73,74]. Moreover, comprehensive scoring systems, such as the Child–Pugh score, is subjective with poor comparability and does not directly reflect perfusion conditions, which are more likely to be influenced by extrahepatic factors [71,75]. As a quantitative liver function test, the ICG retention test is used to assess the dynamics of hepatic function over time in addition to physiologic status (i.e., effective hepatic or hepatic reserve function) and physiologic function (i.e., hepatocyte volume and effective infusion of hepatic blood flow [76,77]. The ICG retention test is a well-established indicator of liver reserve function, which is the most important determinant of the outcome of hepatectomy [76,78,79].

MOJ leads to decreases in the number of functional hepatocytes and hepatic blood flow, which can affect hepatic synthesis and metabolism of biomolecules, resulting in nutritional impairment [74,75,80]. However, ICG uptake is strongly influenced by hepatic blood flow and the collateral circulation, which might be useful to assess the degree of liver dysfunction and cirrhosis rather than screening of liver diseases or as a predictor of mortality.⁸⁰ Hepatocellular carcinoma is frequently associated with different degrees of cirrhosis. A recent study reported ICG-R15 values of 10%–23.7% in 183 patients with moderate cirrhosis, 10%–19% in 91 patients with severe cirrhosis, and >20% in 6.2% patients with severe cirrhosis [80,81], indicating that the risk of malnutrition is similar for patients with moderate and severe disease. Therefore, the combination of the NRS-2002 with the ICG retention test could more accurately reflect liver function and the nutritional status of patients with MOJ. Therefore, further studies are warranted to dynamically assess the nutritional status in patients with MOJ and high bilirubin levels, to investigate the effects of different bilirubin levels on the severity of malnutrition, and to explore potential relationships between serum bilirubin levels and ICG retention rates.

Summary

MOJ can lead to varying degrees of nutritional impairment, thus early detection and treatment could improve patient outcomes and lower hospital costs, while conforming to current healthcare policies. We suggest that the combination of the NRS-2002 with the ICG retention test to more accurately reflect liver function and the nutritional status of patients with MOJ as a guide to improve therapeutic outcomes.

References

- Ramez Bassari, Jonathan B Koea. Jaundice associated pruritis: A review of pathophysiology and treatment. *World J Gastroenterol.* 2015; 21: 1404–1413.
- O'Brien L, Hosick PA, John K, Stec DE, Hinds TD Jr. *Trends Endocrinol Metab.* 2015; 26: 212–220.
- Weaver L, Hamoud AR, Stec DE, Hinds TD Jr. Biliverdin reductase and bilirubin in hepatic disease. *Am J Physiol Gastrointest Liver Physiol.* 2018; 314: G668–G676.
- Hyun YW, Sung YH, Jeong H, Dong UK, Dong HB, et al. Role of endoscopic biliary drainage in advanced hepatocellular carcinoma with jaundice. *PLoS One.* 2017; 12: e0187469.
- Luke O'B, Peter AH, Kezia J, David ES, Terry DHJ. Biliverdin reductase isozymes in metabolism. *Trends Endocrinol Metab.* 2015; 26: 212–220.
- Jin H, Pang Q, Liu HC, Li ZK, Wang Y, Lu Y, et al. Prognostic value of inflammation-based markers in patients with recurrent malignant obstructive jaundice treated by reimplantation of biliary metal stents: A retrospective observational study. *Medicine (Baltimore).* 2017; 96: e5895.
- Chandrashekhara SH, Gamanagatti S, Singh A, Bhatnagar S. Current status of percutaneous transhepatic biliary drainage in palliation of malignant obstructive jaundice: A review. *Indian J Palliat Care.* 2016; 22: 378–387.
- Steven MS, Gao F, Dominic S, David CL, William GH, Fields R, et al. Jaundice: an important, poorly recognized risk factor for diminished survival in patients with adenocarcinoma of the head of the pancreas. *HPB (Oxford).* 2014; 16: 150–156.
- Chisaki I, Naohiko M, Akiko M, Yasuharu K, Tetsuya I, Kobayashi T, et al. Signet-ring cell carcinoma of the ampulla of Vater: a case diagnosed via repeated biopsies. *Clin J Gastroenterol.* 2020; 13: 607–614.
- Luchini C, Grillo F, Fassan M, Vanoli A, Capelli P, Paolino G, et al. Malignant epithelial/exocrine tumors of the pancreas. *Pathologica.* 2020; 112: 210–226.
- Takeshi Okamoto. Malignant biliary obstruction due to metastatic non-hepato-pancreato-biliary cancer. *World J Gastroenterol.* 2022; 28: 985–1008.
- Meyer JE, Messer RJ, Patel VC. Diagnosis and treatment of obstructive jaundice secondary to liver metastases. *Cancer.* 1978; 41: 773–775.
- Maitham AM, Max DL, Eric AR, Nicholas M, Saniay SR. Clinical and histological basis of adenosquamous carcinoma of the pancreas: A 30-year experience. *J Surg Res.* 2021; 259: 350–356.
- Fang Y, Gurusamy KS, Wang Q, Davidson BR, Lin H, Xie X, et al. Pre-operative biliary drainage for obstructive jaundice. *Cochrane Database Syst Rev.* 2012; 9: CD005444.
- Ge PL, Du SD, Mao YL. Advances in preoperative assessment of liver function. *Hepatobiliary Pancreat Dis Int.* 2014; 13: 361–370.
- Hoekstra LT, de Graaf W, Nibourg GA, Heger M, Bennink RJ, Stieger B, et al. Physiological and biochemical basis of clinical liver function tests. *Ann Surg.* 2013; 256: 27–36.
- Mousa OY, Kamath PS. A history of the assessment of liver performance. *Clin Liver Dis (Hoboken).* 2021; 18: 28–48.
- Masatake T, Yasuko I. Lymphatics in the liver. *Curr Opin Immunol.* 2018; 53: 137–142.
- Breslin JW, Yang Y, Scallan JP, Sweat RS, Adderley SP, Murfee WL. Lymphatic vessel network structure and physiology. *Compr Physiol.* 2018; 9: 207–299.
- Masatake T, Yasuko I. The hepatic lymphatic vascular system: Structure, Function, Markers, and Lymphangiogenesis. *Cell Mol Gastroenterol Hepatol.* 2016; 2: 733–749.
- Iwakiri Y. The lymphatic system: A new frontier in hepatology. *Hepatology.* 2016; 64: 706–707.
- Vollmar B, Wolf B, Siegmund S, Kasten D, Menger MD. Lymph vessel expansion and function in the development of hepatic fibrosis and cirrhosis. *Am J Pathol.* 1997; 151: 169–175.
- April O'B, Olga G, Gianfranco A, David Z, Anatoliy G, Glaser S. The role of lymphatics in cholestasis: A comprehensive review. *Semin Liver Dis.* 2020; 40: 403–410.

24. Chung C, Iwakiri Y. The lymphatic vascular system in liver diseases: its role in ascites formation. *Clin Mol Hepatol*. 2013; 19: 99–104.
25. Arrivé L, Monnier-Cholley L, Cazzagon N, Wendum D, Chambe-nois E, et al. Non-contrast MR lymphography of the lymphatic system of the live. *S Eur Radiol*. 2019; 29: 5879–5888.
26. David G Levitt, Michael D Levit. Quantitative assessment of the multiple processes responsible for bilirubin homeostasis in health and disease. *Clin Exp Gastroenterol*. 2014; 7: 307–328.
27. John Y, Chiang L. Bile acid metabolism and signaling. *Compr Physiol*. 2013; 3: 1191–1212.
28. Hamoud AR, Weaver L, Stec DE, Hinds TD Jr. Bilirubin in the liver to gut signaling axis. *Trends Endocrinol Metab*. 2018; 29: 140–150.
29. Eva Sticova, Milan Jirsa. New insights in bilirubin metabolism and their clinical implications. *World J Gastroenterol*. 2013; 19: 6398–6407.
30. Alan F, Hofmann, Lee R. Key discoveries in bile acid chemistry and biology and their clinical applications: history of the last eight decades. *J Lipid Res*. 2014; 55:1553–1595.
31. Lleo A, Maroni L, Glaser S, Alpini G, Marzioni M. Role of cholan-giocytes in primary biliary cirrhosis. *Semin Liver Dis*. 2014; 34: 273–284.
32. Vincenzo R, Clara M, Chiara M, Marco C, Ye HO, Invetnizzi P. Im-mune system and cholangiocytes: A puzzling affair in primary biliary cholangitis. *J Leukoc Biol*. 2020; 108: 659–671.
33. Fabris L, Spirli C, Cadamuro M, Fiorotto R, Strazzabosco M. Emerging concepts in biliary repair and fibrosis. *Am J Physiol Gastrointest Liver Physiol*. 2017; 313: G102–G116.
34. Juan PA, Rosa MM, Vijay HS. Gut–liver axis, cirrhosis and portal hypertension: the chicken and the egg. *Hepatol Int*. 2018; 2: 24–33.
35. Agustín A, Andrea de G, María R. The gut-liver axis in liver dis-ease: Pathophysiological basis for therapy. *J Hepatol*. 2020; 72: 558–577.
36. Matthew M, Yasuko I. Biology of portal hypertension. *Hepatol Int*. 2018; 12: 11–23.
37. Adnan M, Ani AK, Kais Z, Christopher LB, James HT, et al. Preven-tative care in cholestatic liver disease: Pearls for the specialist and subspecialist. *Liver Res*. 2019; 3: 118–127.
38. Asmita P, Anna KK, James PL. Role of the blood coagulation cas-cade in hepatic fibrosis. *Am J Physiol Gastrointest Liver Physiol*. 2018; 315: G171–G176.
39. Ameet D, Fouzia S, Quentin MA, Adam PL, Robert DG, Thursz MR. Thrombin and factor Xa link the coagulation system with liver fibrosis. *BMC Gastroenterol*. 2018; 18: 60.
40. McWhirter JP, Pennington CR. Incidence and recognition of mal-nutrition in hospital. *BMJ*. 1994; 308: 945–948.
41. Correia MITD. Nutrition Screening vs Nutrition Assessment: What’s the Difference?. *Nutr Clin Pract*. 2018; 33: 62–72.
42. Gong Q, Zhu P, Zhang BH, Shu C, Ding ZY, Wu J, et al. Safety and efficacy of n-3 fatty acid-based parenteral nutrition in patients with obstructive jaundice: a propensity-matched study. *Eur J Clin Nutr*. 2018; 72: 1159–1166.
43. Ma BQ, Chen SY, Jiang ZB, Wu B, He Y, Wang XX, et al. Effect of postoperative early enteral nutrition on clinical outcomes and immune function of cholangiocarcinoma patients with ma-lignant obstructive jaundice. *World J Gastroenterol*. 2020; 26: 7405–7415.
44. Tajana P, Stipe P, Nina B, Dominik K, Ivana M, Ivan L, et al. Gut peptide changes in patients with obstructive jaundice underg-ing biliary drainage: A prospective case control study. *World J Clin Cases*. 2022; 10: 5551–5565.
45. Robin Durník, Lenka Šindlerová, Pavel Babica, Ondřej Jurček. Bile acids transporters of enterohepatic circulation for targeted drug delivery. *Molecules*. 2022; 27: 2961.
46. Slobodan R, F. Peter G. Human cytochrome P450 enzymes 5-51 as targets of drugs and natural and environmental compounds: Mechanisms, Induction, and Inhibition—Toxic effects and ben-efits. *Drug Metab Rev*. 2018; 50: 256–342.
47. Assimakopoulos SF, Scopa CD, Vagianos CE. Pathophysiology of increased intestinal permeability in obstructive jaundice. *World J Gastroenterol*. 2007; 13: 6458–6464.
48. Serhat Oguz, Omer Salt, Abdil C Ibis, Saban Gurcan, Dogan Al-bayrak, et al. Combined effectiveness of honey and immuno-nutrition on bacterial translocation secondary to obstructive jaundice in rats: experimental study. *Med Sci Monit*. 2018; 24: 3374–3381.
49. Ohshio G, Furukawa F, Sekita K, Manabe T, Tobe T, et al. IgA containing circulating immune complexes and IgA anti-single stranded DNA antibodies in patients with obstructive jaundice. *Clin Exp Immunol*. 1985; 59: 435–441.
50. Ljungdahl M, Osterberg J, Ransjö U, Engstrand L, et al. Inflamma-tory response in patients with malignant obstructive jaundice. *Scand J Gastroenterol*. 2007; 42: 94–102.
51. Li S, Xu HX, Wu CT, Wang WQ, Jin W, et al. Prognostic value of γ -glutamyltransferase-to-albumin ratio in patients with pancreatic ductal adenocarcinoma following radical surgery. *Cancer Med*. 2019; 8: 572–584.
52. Luo L, Yao YT, Liao HT, Huang JW, Liao MH, et al. Cumulative damage effect of jaundice may be an effective predictor of com-plications in patients undergoing radical resection of Bismuth type II or above hilar cholangiocarcinoma. *Ann Transl Med*. 2021; 9: 861.
53. Simone S, Alberto F, Cosimo S, Cosimo S, Lorenzo V, et al. The Ra-tio of C-Reactive protein to albumin is an independent predictor of malignant intraductal papillary mucinous neoplasms of the pancreas. *J Clin Med*. 2021; 10: 2058.
54. Enver Z, Bilal I, Suad K, Dina Z, Omar Z, et al. Percutaneous biliary drainage for obstructive jaundice in patients with inoperable, malignant biliary obstruction. *Clin Exp Hepatol*. 2022; 8: 70–77.
55. Jiang WG, Puntis MC. Immune dysfunction in patients with ob-structive jaundice, mediators and implications for treatments. *HPB Surg*. 1997; 10: 129–142.
56. Shen ZY, Zhang J, Chen HD, Wang WS, Xu W, et al. Does pre-op-erative biliary drainage influence long-term survival in patients with obstructive jaundice with resectable pancreatic head can-cer?. *Front Oncol*. 2020; 10: 575316.
57. Semi P, Jeong YP, Moon JC, Jae BC, Seung WP, et al. The efficacy of endoscopic palliation of obstructive jaundice in hepatocellu-lar carcinoma. *Yonsei Med J*. 2014; 55: 1267–1272.
58. Jae KK, Chan HP, Ji HH, Jeong YP, Seung WP, et al. Endoscopic management of afferent loop syndrome after a pylorus preserv-ing pancreatoduodenectomy presenting with obstructive jaun-dice and ascending cholangitis. *Clin Endosc*. 2011; 44: 59–64.
59. James R, Jemma M, Felicity E, Kamarjit SM, Prashant P, et al. The outcomes of biliary drainage by percutaneous transhepatic cholangiography for the palliation of malignant biliary obstruc-tion in England between 2001 and 2014: a retrospective cohort study. *BMJ Open*. 2020; 10: e033576.

60. Brooke C, Darren W, Bethany W, Marie S, Paul G, et al. Redefining Nutritional Requirements in End-Stage Liver Disease: Towards a Personalized Approach. *Nutrients*, 2023, 15: 4770.
61. Cathy A, Leah G, Naomi J, Khurshed J, Andrew GD. The relationship between nutritional intake and clinical outcomes in critically ill patients: results of an international multicenter observational study. *Intensive Care Med*. 2009; 35: 1728–1737.
62. Kondrup J, Allison SP, Elia M, Vellas B, Plauth M. ESPEN guidelines for nutrition screening 2002. *Clin Nutr*. 2003; 22: 415–421.
63. Elia M, Stratton R. On the ESPEN guidelines for nutritional screening 2002. *Clin Nutr*. 2004; 23: 131–132.
64. Joseph CW, Sheila ON, Belinda RB, Mark F, Soo KK Comparison of obesity and metabolic syndrome prevalence using fat mass index, body mass index and percentage body fat. *PLoS One*. 2021; 16: e0245436.
65. Yasar-Fisher C, Chen Y, Jackson AB, Hunter GR. Body mass index underestimates adiposity in women with spinal cord injury. *Obesity (Silver Spring)*. 2013; 21: 1223–1225.
66. Mirna HF, Ali IS, Ayman NT, Ghina B, Charif S. et al. Prognostic factors in patients with advanced cholangiocarcinoma: Role of surgery, chemotherapy and body mass index. *World J Gastroenterol*. 2008; 14: 3224–3230.
67. Pironi L, Arends J, Bozzetti F, Cuerda C, Gillanders L. ESPEN guideline on clinical nutrition in liver disease. *Clin Nutr*. 2019; 38: 485–521.
68. Pironi L, Arends J, Bozzetti F, Cuerda C, Gillanders L. ESPEN practical guideline: clinical nutrition in liver disease. *Nutr Hosp*. 2022; 39: 434–472.
69. Pankaj P, Radha KD, Sunil T, Puneeta T, Manuela M, et al. Nutrition in chronic liver disease: Consensus statement of the Indian national association for study of the liver. *J Clin Exp Hepatol*. 2021; 11: 97–143.
70. Canales C, Elsayes A, Yeh DD, Belcher D, Nakayama A et al. Nutrition Risk in Critically Ill Versus the Nutritional Risk Screening 2002: Are They Comparable for Assessing Risk of Malnutrition in Critically Ill Patients? *JPEN J Parenter Enteral Nutr*. 2019; 43: 81–87.
71. Sharma P. Value of liver function tests in cirrhosis. *J Clin Exp Hepatol*. 2022; 12: 948–964.
72. Sakka SG. Assessing liver function. *Curr Opin Crit Care*. 2007; 13: 207–214.
73. Sven MF, Giulio M, Achim K, Martine W, Rebecca D, et al. Non-alcoholic fatty liver disease: A patient guideline. *JHEP Rep*. 2021; 3: 100322.
74. Ruth B, Stefan G Bergthor B, Kristina H, Poya G, et al. Impact of post-hepatectomy liver failure on morbidity and short- and long-term survival after major hepatectomy. *BJS Open*. 2022; 6: zrac097.
75. Philip JJ, Sarah B, Chiaki K, Shinji S Mabel T, et al. Assessment of liver function in patients with hepatocellular carcinoma: A new evidence-based approach—The ALBI Grade. *J Clin Oncol*. 2015; 33: 550–558.
76. Steve H, Jordi C, Gregory TE. Non-Invasive assessment of liver function. *Curr Opin Gastroenterol*. 2015; 31: 199–208.
77. Faybik P, Hetz H. Plasma disappearance rate of indocyanine green in liver dysfunction. *Transplant Proc*. 2006; 38: 801–802.
78. Zheng JL, Xie W, Huang Y, Zhu YF, Jiang L. The technique of 3D reconstruction combining with biochemistry to build an equivalent formula of indocyanine green (ICG) clearance test to assess the liver reserve function. *BMC Surg*. 2020; 20: 283.
79. Wang YY, Zhao XH, Ma L, Ye JZ, Wu FX, et al. Comparison of the ability of Child-Pugh score, MELD score, and ICG-R15 to assess preoperative hepatic functional reserve in patients with hepatocellular carcinoma. *J Surg Oncol*. 2018; 118: 440–445.
80. Wu PC, Guo LZ, Yu S, Zeng N, Liu YC, et al. Noninvasive assessment of liver function reserve with fluorescent dosimetry of indocyanine green. *Biomed Opt Express*. 2022; 13: 1995–2005.
81. Gu J, Zhang EL, Liang B, Zhang ZY, Chen XP, et al. Effectiveness comparison of indocyanine green retention test with the cirrhotic severity scoring in evaluating the pathological severity of liver cirrhosis in patients with hepatocellular carcinoma and Child-Pugh grade A liver function. *World J Surg Oncol*. 2020; 18: 79.