

Review Article

Literature Review on Infectious Bovine Rhinotracheitis

Nagaro Damena*

Department of Veterinary Medicine, Wallaga University,
Oromia Regional State, Ethiopia

*Corresponding author: Nagaro Damena

Department of Veterinary Medicine, Wallaga University,
Oromia Regional State, Ethiopia.

Email: harawwee@gmail.com

Received: March 01, 2024

Accepted: April 05, 2024

Published: April 12, 2024

Summary

Infectious Bovine Rhinotracheitis is a disease of domestic and wild cattle and caused by BoHV-1. The disease was spread globally, but most of the European countries were eradicated it. The infection was manifested by signs of respiratory system, whereas the upper part is mostly affected and produce clinical sign of nasal discharge, reddening of the mouth and nose and conjunctivitis, elevated temperature, depression, loss of appetite, loss of milk yield and abortion. Genital tract also affected by the virus and produce the affection of male and female reproductive system. Isolation of virus from can be occurred from genital and nasal swabs for animals with reproductive problem and respiratory signs. The sample should be collected at the acute face of the infection. In severe cases, the sample also collected from different organs after slaughtering. Latency is the unique feature of the virus and the animals remain carrier of the virus throughout their life. The virus also further subtyped by molecular technique of DNA restriction analysis in to subtypes BoHV1.1 and BoHV1.2. subtype 1.2 can also subdivided in to 2a and 2b. Establishment of IBR depends more on the route of infection than on the subtypes of the BoHV 1. Indirect or blocking ELISA and virus neutralization test are most frequently used for antibody detection. Both attenuated live and inactivated vaccine are used. This vaccine is used to reduce clinical sign and the subsequent shedding of field virus. Moreover, to avoid its occurrence and spread of IBR, it is required to develop coordinated and systematic disease control with the involvement of all pertinent organizations and people.

Keywords: Infectious bovine rhinotracheitis; Cattle; BoHV-1

Introduction

Infectious Bovine Rhinotracheitis (IBR) is a contagious and infectious disease which is commonly distributed globally, with a mandatory notice by the International Health Agencies for Animal Health while disease outbreak arises in Infectious bovine rhinotracheitis-free member states or IBR-free Zones of a countries because of its importance in ways of health and worldwide exchange of livestock and their byproduct [1]. IBR is resulting from BoHV-1 infection, grouped under the family of Herpesviridae [2] of *Alphaherpesvirinae* subfamily; this is the primary problem of numerous respiratory and reproductive system disease within the buffaloes and cattle [3,4]. BoHV-1 has a large host variety that significantly harms products of animal fo exchange and brought losses of > 3 billion dollars yearly within the international livestock production [5].

The infection is usually a herd problem that impacts basically cattle on the age of above 6 months. There are 2 subtypes of BoHV-1 have been recognized. Subtype 1 includes strains that produce respiratory problems, which includes infectious bovine rhinotracheitis, particularly manifested by the presence of exudative rhinotracheitis which affects the bronchi of infected

livestock [6]. Subtype 2 consists of strains which produce reproductive problems such as IPV and IPB [7]. The transmission of virus can be indirect via infected equipment, people, semen, and via transfer of embryo. Directly the virus can be transmitted via air or contact with discharges of reproductive, ocular, or respiratory tracts of infected animals, by that increasing its distribution in dairy livestock [8]. Once the animal infected, they remain carrier throughout their life. while animal infected with latent BoHV-1 infection are encountered to different stress factors, which includes transport, intense weather situation, overcrowding, immunosuppressive remedies (e.g., dexamethasones), latent virus reactivation develop, causing the virus transmission to other animals [9].

Infectious bovine rhinotracheitis is an acute disorder and manifested by general uncomfortable, respiratory symptoms, hyperthermia to 42°C, and in appetite, production of milk and weight reduced [3]. Occasionally, this virus causes udder and uterus inflammation, abortion, loss of fertility, disturbed estrous cycle, and inflammation of epididymis in affected male by this virus [10]. Systemic and neurological diseases can be

produced by BoHV-1 infection as it develops infection, basically in sensorial neurons within the trigeminal ganglia or ganglia of dorsal root [9]. In addition to affecting the welfare and health of animal, it also produces losses to production as a result of elevated costs indirectly related to veterinary prognosis, weight reduction, dairy production discount and loss of life [11].

Enzyme linked immune sorbent assay was highly used for the investigation of sero-prevalence assessment of BoHV-1 antibodies in animal population within the world [12]. Both internal and external factors also affect the sero-prevalence of IBR infection in livestock production [13]. Findings across the world during the last 15 years recorded varying prevalence ranging from 35.9–77.5% in Europe and 37-60.8 % in Latin America [14].

Literature Review

Etiology

Bovine herpesvirus 1 (BoHV-1) is a family member Herpesviridae, subfamily Alpha herpesvirinae Genus: Varicello virus [15]. Herpesviruses are double stranded, big, and enveloped virus. Herpes virus includes a center having linear double stranded DNA and icosadeltahedral capsid of approximately one hundred nanometer diameters containing 162 capsomeres [16]. The virus is related to IBR and IPB/ IPV (Brock *et al.*, 2020). Restrict endonuclease evaluation of DNA was used to observe the virus subtypes, including, BoHV1.1 and BHV1.2 [17,18]. BoHV-1 subtype 1.1 is related to rhinitis and respiratory signs, even as subtype 1.2 is related to IPV and IPB. Subtype 1.2 strains also classified in to BoHV1.2a and BoHV-1 a few subtype1.1 and 1.2a strains causes' abortion, as proven via near connection with clinical cases of abortion and via experimental infection of pregnant heifers [19]. Subtype 1.2b strains are usually connected with genital and respiratory illnesses however not associated with abortion [20].

Within the previous research, virus isolated from buffaloes and goat recognized as BoHV-1 by serological test have a special restrict enzyme profile to subtypes of BoHV-1, are actually seemed as separate viruses and were categorized as BoHV-2 and caprine herpesvirus, respectively. BoHV-5, which was formerly identified as BoHV-1.3 causes meningoencephalitis [21].

Epidemiology of Bovine Herpes Virus 1

For the primary time, the disease of BoHV-1 became recorded within the form of reproductive disorder as IPV in livestock at 1841 by Trommsdorff and Buchner in Germany. Viral affiliation with this contamination first stated at 1928 by Reimann and Reisinger. Within the 1950s, newly emergence of the respiratory form of the infection as infectious bovine rhinotracheitis became recognized in USA. In 1958, the virus became identified effectively for the primary time and its antigenic identification became discovered. Later this viral agent becomes categorized under the own family of Herpesviridae. BoHV-1 is presently wide spread everywhere in the world and confirmed as an example in USA, Canada, Zaire, Italy, Belgium, India, and Turkey [22].

Seroprevalence in livestock done in Turkey, exhibits that this virus is highly distributed some of the beef and dairy cattle in lots of parts of the country (Akca *et al.*, 2004). The past studies performed by Cabalar and Sahna [23] detected that the seropositivity rate of BoHV-1 was between 20% and 74% within the dairy livestock. BoHV-1 basically affects wild and domestic cattle, and further different ruminants can be also affected. Within

the past studies, cross species infection via caprine and bovine herpes virus have been found out [24]. Identification of antibody towards bovine herpesvirus1 in sheep shows that sheep can also play vital role within the epidemiology and upkeep of BoHV-1 within the surroundings [25]. However, cannot play any role within the BoHV-1 spread [26].

In unvaccinated herd, the seroprevalence of BoHV-1 has elevated up 78% within the UK (Woodbine *et al.*, 2009). Many Europeans effectively carried out the IBR eradication program. Others such as Sweden, Finland, Switzerland Denmark and Australia are formally freed from infectious bovine rhinotracheitis [27]. Like different countries, India was also suffering from this disease because the virus became diagnosed in numerous parts of the country, like Kerala [28], Putting a critical interest concerning uncommon bovine species like mithun and yak, the affiliation with BoHV-1 was recognized. The seroprevalence of infectious bovine rhinotracheitis in yak is lowest within the first year of life and highest over the age of 3 years. The area and gender of the yaks became no longer risk factor for the seroprevalence of the virus. The common occurrence in yak became detected to be 41% via VNT and AB-ELISA. The virus became also determined within the ocular swab of yaks infected by infectious bovine keratoconjunctivitis [29,30]. The geographical distribution and source of BoHV-1 infection is unknown because of its serological cross-reactivity with BoHV-1 [31]. In comparison, BoHV-5 infection and disease occurrence to be some often determined in Argentina and Brazil, in which several outbreaks were defined within the ultimate many years [32]. Cross protection via vaccination and natural infection can be happened among BoHV-5 and BoHV-1 (in endemic area) infection [33].

Economic Importance

Bovine herpes virus 1 may also causes production losses via its effect on welfare and fitness, manifested as Respiratory Problem (IBR) and Venereal Disease (IVP)/ (IPB); decreased fertility, lower in yield of milk production and abortion [34]. Annual losses inside the United Kingdom because of the disorder and its treatment had been predicted at as much as €3.1 million accompanied via reduced in weight [35]. An extra-large observe in Ireland predicted a discounted production of 250L/year for cows which had more than one calf within the herds testing positive for antibodies in milk samples. Similarly to the discount in milk production, minor results on herd fertility and death of cattle found; including the proof of developing that subclinical cases can produce ongoing losses in cattle [36].

Losses because of decreased milk production related to sub-clinical contamination were about estimated at 9.5L over incubation duration of fourteen days during a subclinical BoHV-1 contamination on a dairy herd [37]. Outbreaks within the collection center of semen may be very expensive, requiring removal of all bulls within the center [14]. Modeling of the records from 133 farms inside the Netherland indicated an average loss of 0.95kg of milk per cow per day over a 9-week duration following contamination [38]. Modeling of facts for an animal with a sub-clinical case within the United Kingdom mentioned an expected reduction in a milk yield in seropositive in relation to seronegative cows of 2.6kg/day over a 2-year duration [39].

Immune Mechanism and Latency

Immune reaction to BoHV-1 infection: Cattles' immune reaction to BoHV-1 contamination is specific. Even though there are uniformities in immunity reaction to different Alpha herpes-

virus, in case of BoHV-1 there has been no identical immune reaction to the herpes simplex virus in mice and people, and does not correlate with laboratory BoHV-1 contamination in mice. The immune reaction to BoHV-1 contamination is activated while the virus starts to duplicate [40]. Antibody and adaptive cellular mediated immune reaction happen within 7 days after infection (Engels & Ackermann, 1996). Antibody reaction is concept to be vital in stopping infection and viral transmission, even as cellular-mediated immunity plays function in healing from infection [40].

Experimental in addition to infection by natural and vaccine discovered that the antibodies are prompted towards the all glycoprotein of the virus [41], defensive towards viremia and related extreme disorder [42]. The antibody reaction consists quite neutralizing antibodies, and participate to antiviral antibody-based cell cytotoxicity [45], that is frequently complement mediated [43]. Antibodies within the systems can be stayed for an endurable duration, like 3 years following BoHV-1 vaccination [26]. Maternal antibody to BoHV-1 remains for 123 days after weaning at months of age [44]. Newly born calf is protected from BoHV-1 infection after consuming colostrum through vaccine given for cow [42], despite the fact that antibody within the colostrum isn't effectively defensive because calves may have latent BoHV-1 infections at soon of their life within the presence of colostrum antibody too (SCAHW, 2000).

The immune reaction of cellular activated to BoHV-1 infections is under the manager of macrophages, interleukin-2, and interferon- γ production, natural killer cells and natural killer-like actions, proliferation of viral gC- and gD-unique CD4+ T-cells, and activation of cytotoxic T-lymphocyte action [41]. Interferon- γ , interferon- α , and interferon- β had been proven to prevention of viral transmission and to protection towards infection in laboratory infection in mice [46]. Responses to BHV-1 contamination are highly based and consist of both T helper cellular 1 and T helper cellular 2 responses [47].

Latency Development: The specific characteristic of BoHV-1 is the establishment of latency following first infection with natural strain or vaccination with an attenuated strain (Nandi *et al.*, 2009). Latency is thought to arise in nearly all animals which are inflamed with low or high doses of attenuated or virulent BoHV-1 [48]. Vaccine of attenuated strain can stay in a latent condition within the body and vaccination does no longer offer prevention towards establishment of a latent infection with a wild strain [49]. A live vaccine strain of BoHV-1 inoculation also can cause latency and vaccination in latently infected livestock does no longer protect again shedding of a wild strain too (SCAHW, 2000).

Younger aged calves could have latent infection and have immune reaction because of infection within the existence of colostrum antibody (SCAHW, 2000). The infection may provoke through the nasal and oral cavity, or ocular opens and the primary location for latency are the sensory neuron with in trigeminal ganglia. The livestock which is seronegative for BHV-1 antibodies can be latently inflamed with BoHV-1 [26]. Latency can also establish in non-neural components including lymphoid cells of tonsil [50], superficial blood and lymph nodes [51].

Clinical Presentation of Bovine Herpes Virus 1

Bovine herpes virus 1 causes contagious, acute disorder that has an effect on both reproductive and respiratory tract within the following form [52]:

Respiratory Infection: Within the livestock managed under intensive system (for instance feedlot), the respiratory form of the disorder is most often determined and sometimes isn't observed in livestock under grazing situation. The clinical sign and pathological disturbance of bovine herpes virus 1 infection of livestock isn't characterized and will simply be burdened with disorder happened in some of different pathogens and affirmation of the disorder require laboratory examination. The incubation duration extends from 4-7 days. If the infection isn't complicated, the disease can be slight with only mild serous nasal fluid and at increase in body temperature for 1 to 2 days. Many cases stay not noted. In more intense cases, there may be suggested pyrexia of 40 - 42°C, which may also stop for numerous days [53].

Affected animal is depressed with an elevated rate of respiratory and discount of milk production. The preliminary serous nasal discharge sometimes turns into muco-purulent within some days. The mucosas of the nares turns into redden and shallow erosion can be determined. A few animals establish excessive salivation. Oral lesions, that are unusual, include shallow erosion of the oral mucosa. A few animals produce one or both of eye conjunctivitis with clear ocular discharge, which later can also turn out to be muco-purulent. In feedlot or in different intensively managed livestock, there may be sever necrotizing laryngotracheitis and pneumonia which is complicated via contamination with bacteria secondarily. That infection is commonly appeared with in the first 3 to 4 weeks of animal coming into a feedlot. Occasionally, there may be outbreaks of intense pneumonia because of BoHV-1 contamination at later time after come to in to the feedlot. Abortion because of complicated with respiratory form of the BoHV-1 infection has been often determined [52].

Reproductive Infection: Reproductive infection with BoHV-1 happens in each sex and this form of infection is most often the manifestation of this herpes virus contamination in dairy livestock on pasture. This infection may be manifested within the establishment of vesicles, pustules and erosions or ulcers within the mucosa of the vagina or at the penis and prepuce [54].

Balanoposthitis – the disorder in bull is called IPB after 2-3 days incubation duration, pustules determined at the mucosal surface of the prepuce and penis. Those pustules can develop to ulcer with a muco-purulent discharge and not allow bull for mating. A percentage of infected bulls may also shed of virus via semen. In turn, contaminated semen can infect susceptible females via natural mating or artificial inseminations [52].

Vulvovaginitis– is also called infectious pustular vulvovaginitis and is painful situation which can be detected after mating. Repeated micturition and erecting of the tail are the primary symptoms. There can be hyperemia or oedema of the vulva and the posterior third of the vagina. Small pink to white ulcers change in to pustules. There can be a thick yellow or white muco-purulent exudate, particularly when there was complication with secondary bacterial infection [54].

Conjunctivitis– this form of the BoHV-1 infection is quite unusual and resembles 'pinkeye'. There may be commonly involvement of the panophthalmitis and cornea. In a few cases, the most effective sign of contamination is irritation of conjunctiva [53].

Diagnosis of Bovine Herpes Virus 1

Presently, diagnostic methods are restricted to the labora-

tory and needs the usage of modified equipment via especially educated technicians. To discover the BoHV-1 subtypes, several strategies were performed, such as monoclonal antibody-specific antigen and DNA fingerprinting via detect the presence of the restrict portion (Vaz *et al.*, 2016). BoHV-1 may be routinely identified via cell culture, ELISA, VNT, and molecular techniques by using PCR [55]. Gold standard for detecting BoHV-1, it has drawbacks in terms of sensitivity, sperm cytotoxicity, time-taking, and cost. Numerous PCR techniques for identifying BoHV-1 were proven to be powerful [56]. LAMPA has been proposed as easy, short, and optional molecular pathogen test device for natural infection [57]. Isolation of BoHV-1 field strain from the strain in vaccine relying on single Nucleotide Polymorphisms, complete gene sequencing become lately required [20,58].

Serology: The indirect ELISA, competitive ELISA, and VNT are serological assessments often helped for the analysis of BoHV-1 antibodies within the serum [59]. Due to short-time it consumes to give a result, its comfort for testing more serum samples, and the fine indication of any serology used for IBR detection, the indirect ELISA is used most frequently. Moreover, due to the fact BoHV-1 viral latency is not unusual; figuring out serologically positive and in any other case healthful animals is probably a great predictor of infection at herd level. So antibodies positive animals have to be taken into consideration as BoHV-1 infected (with 2 exceptions: serological responses due to inactivated vaccine immunization or colostral antibodies) [52]. Consequently, IBR gE blocking ELISAs discriminate antibodies towards the absent antigen, permitting infected and vaccinated animals to be prominent (DIVA). Due to the fact the virus may reactivate throughout stress or illness, blood ought to be drawn for antibody testing all through the intense phase and once more 2 - 4 weeks later [60].

Polymerase Chain reaction: The PCR was applied to discover Bovine herpes virus 1 nucleic acid in sample [61,62]. Bovine herpes virus 1 and different herpes virus DNA can be isolated from 200 microliter of infected cell culture upper par using the Uniq-10 viral DNA extraction kit. The recovered DNA can be applied as a template for replica DNA synthesis using the Revert Aid™ first strand cDNA synthesis kit, which will be eluted in 50 microliter of nuclease free water, and this entire template can be kept at -70°C for later use [63]. Bovine herpes virus 1 may be identified by usage of RT-PCR [62] and its been proven to be the best method for examining for BoHV-1 abortion, also from affected fetus [64,65].

Differential Diagnosis

Many diseases can cause similar sign, which includes bacterial and viral infections that produce pneumonia, *Mycoplasma bovis*, Lung worm and Sunburns [66]. Occasionally disease like Malignant Catarrhal Fever and Foot and Mouth Disease can also cause similar clinical signs.

Treatment of Infectious Bovine Rhinotracheitis

Sick animal treatment may vary based on case by case. If a diagnosis of Infectious Bovine Rhinotracheitis made the veterinary practitioner may advice immediate isolation and vaccination of the sick and at risk animal with intranasal vaccination to reduce clinical signs and control spread of infection. Vaccination or any treatment cannot resolve latent infection from an animal. However, regular vaccination of latently infected animal can help to reduce reactivation and its spread to other animal. Non-steroidal anti-inflammatory drugs are perhaps the most

important part of IBR treatment as they minimize damage to the upper part of air ways and make the affected animals feel better [67].

Prevention and control of Infectious Bovine Rhinotracheitis

Inactivated and a live vaccine are to be had available on the market, consisting of products with DIVA properties. This performs a critical function within the implementation of prevention and control programmes [14]. The time of prevention is depending on the age of calf, antibody in the colostrum for vaccinated livestock and the kind of product used. The period of prevention and the booster interval is usually six months, even though for different vaccination regimes that is prolonged to twelve months, this can no longer be real in case of calves. Relying at the product, this can be vaccinated via the intramuscular, intranasal or subcutaneous route.

The crucial capacity hazard factors for the introduction of BoHV-1 are recognized [14,68]. They widely fall under the heading of trade (motion in or off) livestock, fomites, people, semen, ova and embryos, and airborne spread. There are to be had measures to overcome the routes of infection. Quarantine, alongside favorable serological screening can reduce the dangers related to exchange, basically if supplemented with information of the brought livestock status and their source herds. Those measures are mainly relevant for nations with authorized regional and country wide control and eradication programmes that want extra guaranties with appreciate to trade. Measures such as a non-go back policy and no longer collaborating in shows (quarantine implementation) and enough boundaries fencing also can restrict touch of animals with the ones in other herds. Aerosol transmission can also arise over a short distance [69].

Suitable disinfection measures, challenge of traffic and their frequency of contact with livestock and implementation of suitable disinfection system or provision of wearing farm specific boots and garb may be used to deal with the risk factors related to fomites and personnel. Bulls getting into semen-collection centers authorized for intra community trade ought to meet quarantine and next monitoring necessities, with semen and embryo imported from third countries issue to similar necessities [70]. Those measures are commonly identified as powerful. Latently infected carrier animal is covered within the food chain to determine the passing of livestock at anti-mortem and post-mortem examinations. Slaughter has to be done within the abattoir. To eliminate seropositive animal at herd stage, culling is best effective while the prevalence has fallen to low level and isn't always practiced within the face of an outbreak disease [69].

Infectious Bovine Rhinotracheitis status in Ethiopia

In Ethiopia, research concerning to IBR have been done however it is not thus far. Therefore, the status of the disease is not as a lot recognized in a country. After the entrance of infected bulls from Holland, the probability prevalence of IBR was diagnosed to be high. Probably unique strains of the virus might cause considerable health effect in susceptible livestock such as illness, abortion, stillbirth, neonatal dying and calf mortality thereby leading to excessive financial losses in livestock enterprise [71].

Even though hindrance of facts is existed, different research exhibits the presence of considerable increase in the prevalence of seropositive animals amongst non-infected livestock. reviews

of serological proof, the significance of the virus because the cause of genital tract problem in milk sheds of central, Southern, and Western Ethiopia has been assessed with a seroprevalence of 30.8%, 45.5% and 55.9% consecutively [72]. Within the latest, prevalence of 77.6% within the north western a part of Ethiopia was also estimated [73].

References

- Newcomer BW, Grady LC, Walz PH, Givens D. Prevention of abortion in cattle following vaccination against bovine herpesvirus 1: A meta-analysis. *Prev Vet Med.* 2017; 138: 1–8.
- Ring SC, Graham DA, Sayers RG, Byrne N, Kelleher MM, Doherty ML. Genetic variability in the humoral immune response to bovine herpesvirus-1 infection in dairy cattle and genetic correlations with performance traits. *J Dairy Sci.* 2018; 101: 6190–204.
- Duque D, Ramon Estevez JN, Abreu AM, Moncada MV, Durango JC, Molina DP. Aspectos sobre Rinotraqueítis Infecciosa Bovina. *J Agric Anim Sci.* 2014; 3: 58–71.
- Maresca C, Scoccia E, Dettori A, Felici A, Guarcini R, Petrini S, et al. National surveillance plan for infectious bovine rhinotracheitis (IBR) in autochthonous Italian cattle breeds: Results of the first year of activity. *Vet Microbiol.* 2018; 219: 150–3.
- Chen X, Wang X, Qi Y, Wen X, Li C, Liu X. Meta-analysis of prevalence of bovine herpes virus 1 in cattle in Mainland China. *Acta Trop.* 2018; 187: 37–43.
- Newcomer BW, Givens D. Diagnosis and control of viral diseases of reproductive importance: Infectious bovine rhinotracheitis and bovine viral diarrhoea. *Vet Clin North Am Food Anim Pract.* 2016; 32: 425–41.
- OIE. Infectious Bovine Rhinotracheitis/Infectious Pustular Vulvovaginitis. *Ch.* 2018; 3: 1139–57.
- Chase CCL, Fulton RW, Toole DO, Gillette B, Daly RF, Perry G. Bovine herpesvirus 1 modified live virus vaccines for cattle reproduction: Balancing protection with undesired effects. *Vet Microbiol.* 2017; 206: 69–77.
- Chothe S, Sebastian A, Thomas A, Nissly R, Wolfgang D, Byukusenge M. Whole-genome sequence analysis reveals unique SNP profiles to distinguish vaccine and wild-type strains of bovine herpesvirus-1 (BoHV-1). *J Virol.* 2018; 522: 27–36.
- Abad Zavaleta J, Utrera ÁR, Fernández JVR, Camacho AG, Martínez JPZ. Prevalencia de rinotraqueítis infecciosa bovina diarrhoea viral bovina en hembras entre épocas del año en la Zona Centro de Veracruz. *Nova Sci.* 2016; 8: 213–27.
- Maresca C, Scoccia E, Dettori A, Felici A, Guarcini R, Petrini S, et al. National surveillance plan for infectious bovine rhinotracheitis (IBR) in autochthonous Italian cattle breeds: Results of the first year of activity. *Vet Microbiol.* 2018; 219: 150–3.
- Roshtkhari F, Mohammadi G, A M. Serological evaluation of relationship between viral pathogens (BHV-1, BVDV, BRSV, PI-3V, and Adeno3) and dairy calf pneumonia by indirect ELISA. *Trop Anim Health Prod.* 2012; 44: 1105–10.
- Keneisezo K, Neithono K, Keneisevono K, Limasenla PKE, KS. Bovine herpes virus -1 (BoHV-1) in cattle: A review with emphasis on epidemiological parameters influencing the prevalence of bovine herpes virus -1 in cattle in India. *J Entomol Zool Stud.* 2019; 7: 284–90.
- Raaperi K, Orro T, Viltrop A. Epidemiology and control of bovine herpesvirus-1 infection in Europe. *Vet J.* 2014; 201: 249–56.
- Davison AJ, Eberle R, Ehlers B, Hayward GS, McGeoch DJ, Minson AC. The order Herpesvirales. *Arch Virol.* 2009; 154: 171–7.
- Harrison SC. Principles of virus structure. Chapter 3. 4th ed. Knipe DM, Howley PM, editors. 2001; 1: 53–85.
- Malla JA, Chakravarti S, Gupta V, Chander V, Sharma GK, Qureshi S, et al. Novel polymerase spiral reaction (PSR) for rapid visual detection of bovine herpesvirus 1 genomic DNA from aborted bovine fetus and semen. *Gene.* 2018; 644: 107–12.
- Dagalp SB, Farzani TA, Dogan F, Alkan F, Ozkul A. Molecular and antigenic characterization of bovine herpesvirus type 1 (BoHV-1) strains from cattle with diverse clinical cases in Turkey. *Tropical animal health and production.* 2020; 52: 555–64.
- Petrini S, Iscaro C, Righi C. Antibody responses to bovine alpha-herpesvirus 1 (BoHV-1) in passively immunized calves. *Viruses.* 2019; 11: 23.
- Fulton RW, d'Offay JM, Dubovi EJ, Eberle R. Bovine herpesvirus-1: Genetic diversity of field strains from cattle with respiratory disease, genital, fetal disease and systemic neonatal disease and their relationship to vaccine strains. *Virus research.* 2016; 223: 115–21.
- Smith GA, Young PL, Mattick JS. Bovine herpesvirus 1.1 an exotic disease agent? *Aust Vet J.* 1993; 70: 272–3.
- Rajkhowa S, Rajkhowa C, Rahman H, Bujabaruah KM. Seroprevalence of infectious bovine rhinotracheitis in mithun in India. *Rev Sci Tech.* 2004; 23: 821–9.
- Çabalar M, Can-Sahna K. Dogu ve Güneydogu Anadolu bölgesinde süt sigirlarında parainfluenza virus-3, bovine herpes virus-1 ve respiratory syncytial virüs enfeksiyonlarının seroepidemiolojisi. *YYÜ Vet Fak Dergisi.* 2000; 11: 101–5.
- Yesilbag K, Bilge-dagalap S, Okur-gumusova S, Gungor B. Studies on herpesvirus infections of goats in Turkey: Prevalence of antibodies to bovine herpesvirus 1. *Revue Med Vet.* 2003; 154: 772–4.
- Jetteur P, Thirty E, Pastoret PP. Serological survey concerning the IBR, CHV2, BVD, PI3, BRS and rinderpest viruses in small ruminants in Zaire. *Rev Elev Med Vet Pays Trop.* 1990; 43: 435–7.
- Hage JJ, Vellema P, Schukken YH, Barkema HW, Rijsewijk FA, Oirschot JT, et al. Sheep do not have a major role in bovine herpesvirus 1 transmission. *Vet Microbiol.* 1997; 57: 41–54.
- Ackermann M, Engels M. Pro and contra IBR eradication. *Vet Microbiol.* 2006; 113: 293–302.
- Bandyopadhyay S, Chakraborty D, Sarkar T, Pal B, Sasmal D, Biswas TK, et al. A serological survey of bovine herpes virus-1 antibodies in yaks (*Poephagus grunniens*). *Rev Sci Tec Off Int Epiz.* 2009; 28: 1045–50.
- Bandyopadhyay S, Chakraborty D, Sarkar T, Pal B, Sasmal D, Biswas TK. A serological survey of bovine herpes virus-1 antibodies in yaks (*Poephagus grunniens*). *Rev Sci Tec Off Int Epiz.* 2009; 28: 1045–50.
- Bandyopadhyay S, Das S, Baruah KK, Chakravarty P, Chakraborty D, Sarkar. Detection of bovine herpesvirus 1 sequences in yaks (*Bos grunniens*) with keratoconjunctivitis, using a highly sensitive nested polymerase chain reaction. *Rev Sci Tec Off Int Epiz.* 2010; 29: 695–703.
- Vogel FSF, Flores EF, Weiblen R, Kunrath CF. Atividade neutralizante anti-herpesvírus bovino tipos 1 (BHV-1) e 5 (BHV-5) no soro de bovinos imunizados com vacinas contra o BHV-1. *Ciênc Rural.* 2002; 32: 881–3.
- Rissi DR, Oliveira FN, Rech RR, Pierezan F, Lemos RB, Barros CLS. Epidemiologia, sinais clínicos e distribuição das lesões encefálicas em bovinos afetados por meningoencefalite por herpesvírus bovino-5. *Pesq Vet Bras.* 2006; 26: 123–32.

33. Thiry J, Keuser V, Muylkens B, Meurens F, Gogev S, Vanderplassen A, et al. Ruminant alphaherpesviruses related to bovine herpesvirus 1. *Vet Res.* 2006; 37: 169–90.
34. Graham DA. Bovine herpesvirus-1 (BoHV-1) in cattle—a review with emphasis on reproductive impacts and the emergence of infection in Ireland and the United Kingdom. *Ir Vet J.* 2013; 66: 15.
35. Bennett R, Ijpelaar J. Updated estimates of the costs associated with thirty four endemic livestock diseases in Great Britain: a note. *Journal of Agricultural Economics.* 2005; 56: 135–44.
36. Sayers RG. Associations between exposure to bovine herpesvirus 1 (BoHV-1) and milk production, reproductive performance, and mortality in Irish dairy herds. *Journal of Dairy Science.* 2017; 100: 1340–52.
37. Hage JJ, Schukken YH, Dijkstra T, Barkema HW, Valkengoed PH, Wentink GH. Milk production and reproduction during a subclinical bovine herpesvirus 1 infection on a dairy farm. *Preventive Veterinary Medicine.* 1998; 34: 97–106.
38. Schaik G, Shoukri M, Martin SW, Schukken YH, Nielen M, Hage JJ, et al. Modeling the effect of an outbreak of bovine herpesvirus type 1 on herd-level milk production of Dutch dairy farms. *J Dairy Sci.* 1999; 82: 944–52.
39. Statham JM, Randall LV, Archer SC. Reduction in daily milk yield associated with subclinical bovine herpesvirus 1 infection. *Veterinary Record.* 2015; 177: 339.
40. Babiuk LA, Drunen LH, Tikoo SK. Immunology of bovine herpesvirus 1 infection. *Vet Microbiol.* 1996; 53: 31–42.
41. Tikoo SK, Campos M, Babiuk LA. Bovine herpesvirus 1 (BHV-1): biology, pathogenesis, and control. *Adv Virus Res.* 1995; 45: 191–223.
42. Mechor GD, Rousseaux CG, Radostits OM, Babiuk LA, Petrie L. Protection of newborn calves against fatal multisystemic infectious bovine rhinotracheitis by feeding colostrum from vaccinated cows. *Can J Vet Res.* 1987; 51: 452–9.
43. Rouse BT, Grewal AS, Babiuk LA, Fujimiya Y. Enhancement of antibody-dependent cell-mediated cytotoxicity of herpesvirus-infected cells by complement. *Infect Immun.* 1977; 18: 660–5.
44. Fulton RW, Saliki JT, Burge LJ, Payton ME. Humoral immune response and assessment of vaccine virus shedding in calves receiving modified live virus vaccines containing bovine herpesvirus-1 and bovine viral diarrhoea virus 1a. *J Vet Med B Infect Dis Vet Pub Health.* 2003; 50: 31–7.
45. Tikoo SK, Campos M, Popowych YI, van Drunen S, Little-van de H, Babiuk LA. Lymphocyte proliferative responses to recombinant bovine herpes virus type 1 (BHV-1) glycoprotein gD (gIV) in immune cattle: identification of a T cell epitope. *Viral Immunol.* 1995; 8: 19–25.
46. Abril C, Engels M, Liman A, Hilbe M, Albini S, Franchini M, et al. Both viral and host factors contribute to neurovirulence of bovine herpesviruses 1 and 5 in interferon receptor-deficient mice. *J Virol.* 2004; 78: 3653.
47. Mena A, Ioannou XP, Kessel A, Drunen LD, Popowych Y, Babiuk LA, et al. Th1/Th2 biasing effects of vaccination in cattle as determined by real-time PCR. *J Immunol Methods.* 2002; 263: 11–21.
48. Pastoret PP, Thiry E, Brochier B, Derboven G. Bovine herpesvirus 1 infection of cattle: pathogenesis, latency, consequences of latency. *Ann Rech Vet.* 1982; 13: 221–35.
49. Jones C, Newby TJ, Holt T, Doster A, Stone M, Ciacci Zanella J. Analysis of latency in cattle after inoculation with a temperature sensitive mutant of bovine herpesvirus. 2000; 1: 3185–95.
50. Winkler MT, Doster A, Jones C. Persistence and reactivation of bovine herpesvirus 1 in the tonsils of latently infected calves. *J Virol.* 2000; 74: 5337–46.
51. Mweene AS, Okazaki K, Kida H. Detection of viral genome in non-neural tissues of cattle experimentally infected with bovine herpesvirus 1. *Jap J Vet Res.* 1996; 44: 165–74.
52. Chatterjee A, Bakshi S, Sarkar S, Mitra J, Chowdhury S. Bovine herpes virus-1 and its infection in India - a review. *Indian J Anim Hlth.* 2016; 55: 21–40.
53. Gheorghita D, Paul-Adrian BOR, Mariana RUSU, Carmen Dana S, Diana O, Marina S, et al. Health impacts and control measures in infectious bovine rhinotracheitis scientific Works. *Series C Veterinary Medicine.* 2022; LXVIII.
54. Miller JM, Whetstone CA, van der Maaten MJ. Abortifacient property of bovine herpesvirus type 1 isolates that represent three subtypes determined by restriction endonuclease analysis of viral DNA. *Am J Vet Res.* 1991; 52: 458–61.
55. Biswas S, Bandyopadhyay S, Dimri U, Patra H. Bovine herpesvirus-1 (BoHV-1) a re-emerging concern in livestock: A revisit to its biology, epidemiology, diagnosis, and prophylaxis. *Vet Q.* 2013; 33: 68–81.
56. Kumar S, Leela N, Kota S, Narasimha P, Rajan S, Alwar V. Use of real-time polymerase chain reaction to detect bovine herpesvirus-1 in frozen cattle and buffalo semen in India. *Veterinaria Italiana.* 2011; 47: 313–22.
57. Suwancharoen D, Sittiwicheanwong B, Wiratsudakul A. Evaluation of loop mediated isothermal amplification method (LAMP) for pathogenic *Leptospira* spp. detection with *Leptospira* isolation and real-time PCR. *J Vet Med Sci.* 2016; 78: 1299–302.
58. Fulton R, d'Offay J, Eberle R, Moeller R, Campen H. Bovine herpesvirus-1: Evaluation of genetic diversity of subtypes derived from field strains of varied clinical syndromes and their relationship to vaccine strains. *Vaccine.* 2015; 33: 549–58.
59. Mahajan V, Banga H, Deka D, Folia G, Gupta A. Comparison of Diagnostic Tests for Diagnosis of Infectious Bovine Rhinotracheitis in Natural Cases of Bovine Abortion. *J Comp Pathol.* 2013; 149: 391–401.
60. Peter NA, Russell G. Update on infectious bovine rhinotracheitis. *Group BMJ Com.* 2017; 39: 255–72.
61. Rana SK, Kota SNLS, Samayam PNR, Rajan S, Srinivasan VA. Use of real-time polymerase chain reaction to detect bovine herpesvirus 1 in frozen cattle and buffalo semen in India. *Vet Ital.* 2011; 47: 313–22.
62. Thonur L, Maley M, Gilray J, Crook T, Laming E. One-step multiplex real time RT-PCR for the detection of bovine respiratory syncytial virus, bovine herpesvirus 1 and bovine parainfluenza virus 3. *BMC Vet Res.* 2012; 8: 37.
63. Peili H, Hongmei W, Guimin Z, Chengqiang H, Hongbin H. Rapid detection of infectious bovine Rhinotracheitis virus using recombinase polymerase amplification assays. *BMC Vet Res.* 2017; 13: 386.
64. Crook T, Benavides J, Russell G, Gilray G, Maley M, Willoughby K. Bovine Herpes Virus 1 abortion: current prevalence in the United Kingdom and evidence of haematogenous spread within the fetus in natural cases. *J Vet Diagn.* 2012; 24: 662–70.

65. Wernine K, Hoffmann B, Kalthoff D, König P, Beer M. Development and validation of a triplex real time-PCR assay for the rapid detection and differentiation of wild-type and glycoprotein E-deleted vaccine strains of Bovine herpesvirus type 1. *J Virol Methods*. 2011; 174: 77–84.
66. Radostits OM, Gay CC, Hinchcliff KW, Constable PD. Infectious Bovine Rhinotracheitis Infection. In: *Veterinary Medicine*. Philadelphia: Saunders Elsevier. 2007; 1349–61.
67. Muylkens B, Thiry J, Kirten P, Schynts F, Thiry E. Bovine herpes virus 1 infection and infectious bovine rhinotracheitis. *Vet Res*. 2007; 38: 181–209.
68. EFSA. Opinion of the Scientific Panel on Animal Health and Welfare related to the “Definition of a BoHV-1-free animal and a BoHV-1-free holding, and the procedures to verify and maintain this status. *EFSA Journal*. 2006; 4: 2903.
69. Simon M, Anette B, Andrew B, Paolo C, Klaus D, Sandra E. Assessment of listing and categorisation of animal diseases within the framework of the Animal Health Law. *EFSA J*. 2017; 15: 04947.
70. Mars MH, Jong MC, Maanen C, Hage JJ, Oirschot JT. Airborne transmission of bovine herpesvirus 1 infections in calves under field conditions. *Veterinary Microbiology*. 2000; 76: 1–13.
71. Hagos A, Kuastros MB. Qualitative Risk Analysis of IBR Introduction to Ethiopia via the Legal Importation of Bulls from Netherlands. *Animal and Veterinary Sciences*. 2021; 9: 80–7.
72. Sibhat B, Ayelet G, Skjerve E, Gebremedhin EZ, Asmare K. Bovine herpesvirus-1 in three major milk sheds of Ethiopia: Serostatus and association with reproductive disorders in dairy cattle. *Prev Vet Med*. 2018; 150: 126–32.
73. Zewde D, Tadesse T, Alemu S. Sero Status and Presumed Risk Factors Assessment for Bovine Herpesvirus-1 in North Western, Ethiopia. *Austin J Vet Sci & Anim Husb*. 2021; 8: 1-8.
74. Schwarz AJ, York CJ, Zirbel LW, Estela LA. Modification of IBR virus in tissue culture and development of a vaccine, *Proc. Soc Exp Biol Med*. 1957; 96: 453–8.
75. Suman B, Samiran B, Umesh D, Pabitra HP. Bovine herpesvirus-1 (BHV-1) a re-emerging concern in livestock: a revisit to its biology, epidemiology, diagnosis, and prophylaxis. *Veterinary Quarterly*. 2013; 33: 68–81.